Serum tube identification of *Candida albicans*

D. W. R. MACKENZIE

*From the Department of Microbiology, The Queen’s University of Belfast*

SYNOPSIS  The production of filaments (germ tubes) by cells of *Candida albicans* in serum tubes permits presumptive identification to be made within two to three hours. The proportion of yeast-like cells forming filaments is progressively decreased with increasing cell concentration. The test is effective over a comparatively wide range of temperatures and using different types and concentrations of sera.

Routine identification of *Candida albicans* in diagnostic laboratories is usually dependent on the production on certain media of the characteristic refractile chlamydospores. Several other techniques have been described where the criteria are less specific, but which may have considerable practical value in permitting a rapid presumptive diagnosis. Such a test has been recently described by Taschdjian, Burchall, and Kozinn (1960), by which a rapid (two to three hours) provisional identification of *C. albicans* can be made by observing changes in the morphology of yeast cells in serum or serum derivatives. This paper deals with studies on the value of the ‘serum tube’ test in identifying *C. albicans*.

**MATERIAL AND METHODS**

YEASTS  The following species of yeast capable of growth at 37°C. were used in the investigations:—

One hundred and sixty-three isolates of *C. albicans* from seven diagnostic laboratories in the United Kingdom. All produced chlamydospores on Oxoid cornmeal agar.

*C. brumptii*, *C. rugosa*, *C. utilis*, *Saccharomyces fragilis*, and *Schizosaccharomyces versatilis* were obtained from the National Collection of Yeast Cultures; *C. krusei*, *C. guilliermondii*, *C. macedoniensis*, *C. pseudotropicalis*, *C. stellatoidea* from the Mycological Reference Laboratory, London School of Hygiene and Tropical Medicine; and *C. tropicalis*, *Trichosporon capitatum*, and *Geotrichum sp.* were isolated at this laboratory from human sources during routine mycological examinations.

TECHNIQUE  The test organism is inoculated into 0.5 ml. of serum in a small tube or container (Kahn, Durham tube, Bijoux bottle), and incubated at 37°C.

RESULTS

The rounded or short-oval cells of *C. albicans* rapidly give rise to filamentous outgrowths and within two or three hours numerous ‘hand mirror’ forms (Hu, Livingood, Johnson, and Pomerat, 1954) are visible (Fig. 1). These are characteristic of *C. albicans*, and under appropriate conditions (see below) are produced by over 95% of the cells. They have been termed ‘germ tubes’ (Mackenzie, 1958; Taschdjian et al., 1960), thus emphasizing the close morphological similarities to the initial hypha of true filamentous fungi. Conditions affecting their formation have been studied as follows:

![FIG. 1. Germ tubes of Candida albicans in bovine serum after three hours at 37°C. × 640.](image-url)
Taschdjian et al. (1960) reported that germ tubes were formed in fresh and inactivated human serum and in 'deep-frozen' stored material. Dog serum also proved satisfactory. These observations were confirmed and extended to include rabbit, guinea-pig, horse, and bovine sera. The duration and temperature of the storage of serum did not affect the incidence of germ tubes or their rate of appearance and subsequent development. They were not formed in heat-coagulated serum. With a standardized inoculum ($8 \times 10^8$ cells) and estimations of percentage germ tubes at each dilution based on counts of 500 cells, the incidence of filamentation is highest in undiluted serum and decreases with decreasing concentration of the serum (Fig. 2). The rapidity of the test makes sterilizing the glassware unnecessary.

The age of the inoculum is apparently not insignificant in determining the presence or absence of hyphal development. Cultures which had been stored at room temperature for up to six months formed germ tubes when they were inoculated into serum although the percentage of cells forming filaments naturally decreased as the cells became increasingly less viable.

In contrast, germ tube formation in serum is directly affected by the concentration of cells in the inoculum (Fig. 3). When the number exceeded $10^7$ per ml, the incidence of germ tubes decreased until at concentrations of $5 \times 10^6$ cells per millilitre, and at greater concentrations they were virtually absent. The percentage of germ tubes derived from counts of 500 cells at each concentration after three hours at $37^\circ$C., using fresh inactivated guinea-pig serum and a standardized volume of inoculum, and the relationship between length of strands and concentration of inoculum, are shown in Figure 3. Other sera tested showed a more marked inhibition of germ tubes at high cell densities.

The concentration of cells required to suppress germ tube formation by 50% was determined for guinea-pig, bovine, and horse sera with the results shown in Table I. If the inoculum of a serum tube

![Graph](http://jcp.bmj.com/)

**TABLE I**

<table>
<thead>
<tr>
<th>Serum (Inactivated)</th>
<th>50% Inhibition (cells per ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Guinea-pig</td>
<td>$4 \times 10^7$</td>
</tr>
<tr>
<td>Horse</td>
<td>$6.5 \times 10^7$</td>
</tr>
<tr>
<td>Ox</td>
<td>$7.5 \times 10^7$</td>
</tr>
<tr>
<td>Calf</td>
<td>$9.5 \times 10^7$</td>
</tr>
</tbody>
</table>

![Graph](http://jcp.bmj.com/)

**FIG. 3.** Germ tube incidence and growth in relation to concentration of inoculum.

results in too great a concentration of yeast cells germ tube formation might be inhibited to the extent of invalidating the test. In practice, however, complete inhibition is unlikely to be achieved although marked suppression is an established sequel to an excessively large inoculum.

Ten laboratory workers were asked to inoculate serum tubes from slant cultures of *C. albicans*, and the final concentration of cells was determined. This ranged from $2 \times 10^6$ to $6.4 \times 10^7$ cells per millilitre, corresponding to an anticipated incidence of germ tubes of 25 to 95% in most of the sera tested. Even
at the lower figure, strand formation is sufficiently distinct to permit ready identification of *C. albicans* or *C. stellatoidea*. In one batch of calf serum, however, only 8% of cells in the inoculum formed strands at a concentration of $6.4 \times 10^7$ cells per millilitre. Clearly a 92% inhibition of germ tubes is undesirable and cannot be expected to give satisfactory results. When small quantities of serum are employed, there is a distinct possibility of the test being ineffectual. The best results are obtained when small quantities of inoculum are used, imparting no more than a faint turbidity to the serum substrate. Tests with common laboratory agar media (glucose-peptone, 2.5% malt extract, malt tellurite, blood, glucose-nutrient, and 2% peptone) showed no correlation between the percentage of germ tubes in serum and the type of medium supporting growth of the inoculum. Washing with normal saline before inoculation did not affect the ultimate percentage of germ tubes, and is considered to be unnecessary.

**TEMPERATURE** Tests performed between 30°C and 42°C at 1° intervals showed that germ tubes were formed between 31°C and 41°C.

**SPECIFICITY** The formation of filaments in serum or serum derivatives is, like the production of chlamydoспорes, characteristic of *C. albicans* but is not confined to this species alone. It has been noted in *C. stellatoidea*, *C. utilis*, *C. rugosa*, and *Schizosaccharomyces fragilis*, whilst the presence of elongated buds or pseudomycelial cells in inocula of *C. brumptii* and *C. pseudotropicalis* may mimic the germ tubes of *C. albicans*.

Specificity is relative, i.e., although the above species of yeasts form strands when grown in serum, they are (with the exception of *C. stellatoidea*) insignificant members of the population of sites likely to yield *C. albicans*. The value of the test lies in its rapidity and ease of execution, and it is best regarded as a screening test for yeasts of possible medical significance. It should be applied only to those isolates capable of growth at 37°C. whose growth on primary isolation is essentially yeast-like, i.e., rounded, budding cells. Where microscopic examination shows the predominance of elongated or thread-like cells in young cultures it can be concluded that the isolate is not *C. albicans*.

**CONCLUSIONS**

It is concluded that serum tube tests permit a rapid presumptive diagnosis of *C. albicans*. Chlamydoспор formation and biochemical characteristics (fermentation, assimilation, etc.) remain the specific criteria for positive identification and differentiation from other species of *Candida*. In a random series of 163 isolates identified as *C. albicans* in various laboratories in the United Kingdom, it was found that eight (5%) failed to produce germ tubes and were shown by further investigation to be incorrectly identified. Although less rapid than agglutination tests with specific antisera, a positive result can nevertheless be obtained in two hours compared with overnight incubation which is required for the detection of chlamydoспорes.

This study was supported by a grant from the Medical Research Council. Acknowledgements are made of the technical assistance received from Mr. D. L. Corkin and Miss Lesley Rusk.

**REFERENCES**


Serum tube identification of Candida albicans

D. W. R. Mackenzie

doi: 10.1136/jcp.15.6.563

Updated information and services can be found at:
http://jcp.bmj.com/content/15/6/563

These include:

Email alerting service
Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

Notes

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/