A standard clot method for the assay of plasminogen activators, anti-activators, and plasmin

CHRISTINE M. HAWKEY AND J. L. STAFFORD

From the Haematology Department, St. George’s Hospital Medical School, London

SYNOPSIS. Details are given of the selection and standardization of reagents for use in a standard clot method for estimating plasminogen activators, anti-activators, and plasmin.

Substrate standardization is necessary for the assay of components of the fibrinolytic system, not only to permit reproducibility in the laboratory but, in particular, to facilitate comparison of results recorded in different centres during the development of this relatively new subject. Difficulties experienced with reproducibility are due mainly to the fact that commercially available fibrinogens and thrombins are contaminated in varying degree by other components of the fibrinolytic system, whilst human plasminogen is difficult and expensive to prepare. Procedures involving a standard clot have already been used (Christensen, 1949; Johnson and Tillett, 1952; Fletcher, 1954; Baumgarten, Ciminera, and Cole, 1960). The purpose of this communication is the detailed description of a method found to be suitable for general laboratory use, incorporating reagents readily available in Europe: the results obtained during standardization are recorded and the selected commercial preparations specified.

The standard clot comprises fibrinogen and thrombin, to which plasminogen and activator may be added if appropriate. It is made up to a fixed volume with buffer and thus provides ‘reagent’ space so that the components to be tested are actually included in the fibrin clot and their intrinsic effect on the subsequent lysis time can be measured. The method depends on the fibrin clot so formed being reproducible, stable, of constant volume, and uniformly susceptible to the action of plasmin. It offers a number of advantages over others in current use. The selection of fibrin as the substrate more nearly reproduces physiological conditions than does that of casein or the synthetic esters. It is shorter and easier to use quantitatively than the fibrin plate method (Astrup and Mullertz, 1952) which has the further disadvantage that the test substance cannot actually be incorporated in the clot. Neither the special apparatus required for the 125I-labelled clot procedure nor the preparations involved in the standard thrombus (Blix, 1962) are necessary.

METHOD

Only those materials easily obtainable in Europe or prepared in the laboratory have been investigated and, when possible, more than one batch of each reagent has been tested to assess variation from batch to batch. Throughout, meticulous attention has been paid to storage and the provision of optimal conditions necessary for the maintenance of stability.

Other workers have already selected 1.1 ml. as a standard volume and this provides a conveniently sized test system. Basically, the clot consists of 0.2 ml. fibrinogen and 0.1 ml. thrombin, each in a pre-standardized solution, leaving a reagent space of 0.8 ml. into which plasminogen, activator, and test substances can be introduced as desired, the volume being made up by saline phosphate buffer (Appendix 1).

The test is carried out in new 75 × 12 mm. glass test-tubes which have been washed in detergent, rinsed, soaked overnight in 2% HCl, and re-rinsed thoroughly with tap and then distilled water. Tubes and reagents are placed in an ice-bath. Reagents are added as rapidly as possible in the sequence saline buffer, plasminogen, fibrinogen, activator, and, finally, thrombin; several tubes can be set up together, providing that the total length of time spent in preparation is not more than one and a half minutes. A stop-watch is started as the thrombin is added, the contents of the tube are quickly mixed, preferably by means of an automatic mixer, and the tubes transferred to a water-bath at 37°C. One minute later they are examined to confirm that clotting has taken place; thereafter they are examined every 30 seconds, or at regular intervals determined by experience of the particular test being carried out. The end-point is taken as the time of complete disappearance of the clot and is recorded as the lysis time in minutes.

INTERPRETATION OF RESULTS. The lysis times of serial dilutions of the test substances are plotted against...
concentration. A 'standard clot' unit of activity is the concentration required to produce complete lysis in an arbitrarily fixed time. It is convenient to select 10 minutes for this purpose.

REAGENTS

Two points in the selection of reagents merit emphasis. With all protein substrates, the varying amount of contamination must be assessed and, if possible, the cause identified (Appendix 2). The choice of buffer can be critical: either as a result of interaction with the different proteins or because of its effect upon ionic calcium, the solution used can exert an influence on results. Calcium inhibits some phases of the fibrinolytic mechanism (Bruce, 1964) and so a saline phosphate buffer (pH 7.5) has been used in the tests described in this paper because ionic calcium is removed as calcium phosphate. The inclusion of a trace of merthiolate in the buffer has not been found to influence the results of the test, and is useful in preventing the growth of contaminating moulds and algae. In the preparation of streptokinase dilutions, gelatin is added to the buffer solution because the provision of a gel serves to stabilize the streptokinase during the time taken to complete the procedures (Appendix 3).

FIBRINOGEN It is, of course, preferable to use human fibrinogen as a substrate and that manufactured by Kabi (Sweden) is to be recommended. But this is relatively expensive and, as fibrinogen is a basic component of the standard clot, it is important to have a source from which it can be obtained readily and cheaply. For this reason Armour bovine plasma fraction I has been selected, although it is known to contain several contaminant proteins which vary considerably from batch to batch, and a substantial proportion of the dry powder is citrate.

The freeze-dried fraction I is stored at 4°C, and weighed out as required. It is dissolved in saline phosphate buffer at 37°C for 15 minutes and then kept in an ice-bath where it remains stable for at least four hours. If a precipitate forms at this temperature (‘profibrin’) it can be re-dissolved by warming the solution to 37°C for a few minutes.

Three batches have been tested: each was found to contain differing amounts of clottable protein, plasminogen activator, plasminogen itself, and inhibitors (Table I). The trace of plasminogen activator detectable on the fibrin plate was not enough to cause lysis of a standard clot in 24 hours and was, therefore, not considered relevant. The unpredictable effect of the plasminogen could be eliminated by the deliberate addition of excess plasminogen from human serum euglobulin (see later). Even though the effect of plasminogen contamination was nullified in this way, variation of the clot lysis time still occurred when 1% solutions (w/v) of different batches of fraction I were used in the preparation of streptokinase-activated standard clots. Moreover, this could not be corrected by separating out contaminant antiplasmins by euglobulin precipitation. Calibration curves for streptokinase made with fraction I from the various batches were, however, always found to be parallel when the plasminogen level was controlled (Fig. 1); from this observation it was concluded that the concentration of substrate was itself influencing the result, and estimation of clottable protein in each batch confirmed that this varied. By adjusting the amount

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**TABLE I**

RESULTS OF TESTS FOR CONTAMINANTS IN REAGENTS

<table>
<thead>
<tr>
<th>Material Tested</th>
<th>Batch No.</th>
<th>Strength of Solution Tested</th>
<th>Plasmin</th>
<th>Plasminogen Activator</th>
<th>Pro-activator</th>
<th>Anti-activator</th>
<th>Anti-plasmin</th>
</tr>
</thead>
<tbody>
<tr>
<td>Armour plasma fraction I (bovine)</td>
<td>DB 1770</td>
<td>1%</td>
<td>0</td>
<td>+</td>
<td>±</td>
<td>0</td>
<td>±</td>
</tr>
<tr>
<td></td>
<td>FA 1770</td>
<td>1%</td>
<td>0</td>
<td>+</td>
<td>±</td>
<td>0</td>
<td>±</td>
</tr>
<tr>
<td></td>
<td>EF 0573</td>
<td>1%</td>
<td>0</td>
<td>+</td>
<td>±</td>
<td>0</td>
<td>±</td>
</tr>
<tr>
<td>Kabi fibrinogen (human)</td>
<td>71063</td>
<td>1%</td>
<td>0</td>
<td>+</td>
<td>0</td>
<td>0</td>
<td>±</td>
</tr>
<tr>
<td></td>
<td>71035</td>
<td>1%</td>
<td>0</td>
<td>+</td>
<td>0</td>
<td>0</td>
<td>±</td>
</tr>
<tr>
<td></td>
<td>64462</td>
<td>1%</td>
<td>0</td>
<td>+</td>
<td>0</td>
<td>0</td>
<td>±</td>
</tr>
<tr>
<td>Warner Chilcott fibrinogen</td>
<td>0688091A</td>
<td>1%</td>
<td>0</td>
<td>+</td>
<td>0</td>
<td>0</td>
<td>+</td>
</tr>
<tr>
<td>Parke, Davis thrombin topical (bovine)</td>
<td>086127A</td>
<td>100 N.I.H. units per ml.</td>
<td>±</td>
<td>+</td>
<td>±</td>
<td>0</td>
<td>±</td>
</tr>
<tr>
<td></td>
<td>080449A</td>
<td></td>
<td>±</td>
<td>+</td>
<td>0</td>
<td>±</td>
<td>±</td>
</tr>
<tr>
<td>Leo thrombin reagent (bovine)</td>
<td>P 162021</td>
<td>100 N.I.H. units per ml.</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>Kabi plasminogen (human)</td>
<td>1127</td>
<td>250 units per ml.</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Merck Sharp &amp; Dohme plasminogen (human)</td>
<td>Not given</td>
<td></td>
<td>0-01%</td>
<td>0</td>
<td>+</td>
<td>0</td>
<td>+</td>
</tr>
<tr>
<td>Human serum euglobulin</td>
<td>All batches prepared</td>
<td>Standardized as described</td>
<td>0</td>
<td>+</td>
<td>±</td>
<td>±</td>
<td>0</td>
</tr>
</tbody>
</table>
then calculated: found same speed. DB 1770
STANDARDIZATION 1770 FA
Fraction I
Concentration of other
No. of time which would provide similar
evaluated, using this
thereafter, this
FIG. 1. Streptokinase dilution curves using 1% solutions
of different batches of fraction I.
 EF 0573, ○○○ FA 1770, × DB 1770.
The streptokinase units referred to are manufacturer's
units.
(w/v) of fraction I in the standard clot and thus controlling
the ultimate amount of fibrin formed, it was found that
unknown batches of fraction I could be standardized
against a pre-selected control batch.
Standardization Initially a streptokinase dilution curve was plotted for each of the three fibrinogen batches
examined, using a standard clot mixture incorporating
1% fibrinogen solution (Fig. 1). The batch providing the
fastest lysis time (Armour EF 0573) was selected and,
thereafter, 0-2 ml. of a 1% solution of this batch (50 mg.
in 5 ml. saline phosphate buffer) was used in the pre-
paration of every standard clot.
Although batch EF 0573 fibrinogen was established as
the reference for all subsequent estimations, the con-
centration of other batches of fraction I was determined which would provide similar results. Standard clots
incorporating serial dilutions of the unknown fraction I
were prepared in parallel with the reference fibrinogen.
For this particular experiment, streptokinase in a
concentration of 10 units/ml., providing a 10-minute
lysis time, was diluted to 1 unit/ml., thus prolonging the
time of complete lysis to 20 to 23 minutes (Table II).

| TABLE II |
| STANDARDIZATION OF FIBRINOGEN (ARMOUR BOVINE PLASMA |
| FRACTION I) |

<table>
<thead>
<tr>
<th>Fraction I</th>
<th>Lysis Time in Minutes</th>
</tr>
</thead>
<tbody>
<tr>
<td>Batch No.</td>
<td>Volume of 1% Fraction I Solution in Standard Clot</td>
</tr>
<tr>
<td></td>
<td>0:3</td>
</tr>
<tr>
<td>EF 0573</td>
<td>52</td>
</tr>
<tr>
<td>FA 1770</td>
<td>82</td>
</tr>
<tr>
<td>DB 1770</td>
<td>36</td>
</tr>
</tbody>
</table>

The amount of fraction I in the clot which lysed at the
same speed as that containing reference fibrinogen was
then calculated: using a solution containing this quantity
in 0-2 ml., the calibration curve for streptokinase was
found to coincide exactly with the curve from the
reference batch within the range 50 to 0.5 units strepto-
kine per millilitre.

The standardization of Armour batches FA 1770 and
DB 1770 (Tables II and III) showed that these could be
used in the preparation of a standard clot as follows:—
FA 1770: 0.2 ml. of a solution containing 37.5 mg. in
5 ml. SP buffer (0.75%, w/v).

DB 1770: 0.2 ml. of a solution containing 25 mg. in
5 ml. SP buffer (0.5%, w/v).

Other sources Warner bovine fibrinogen has been
found to be free from significant contamination with
activator although the batch tested contained some
plasminogen (Table I). Batches of Kabi human fibrinogen,
which should be standardized as described for use in the
standard clot, have also been found to be free from activa-
tor and, if necessary, the plasminogen can be removed by
the method of Mosesson (1962).

Howell (1963) refers to the use of time-expired bank
blood as a convenient laboratory source of human
fibrinogen which can be used when plasminogen con-
tamination does not matter: a plasma-euglobulin
preparation gives a fibrinogen-plasminogen mixture free
from anti-plasmin activity.

THROMBIN Two commercial thrombin preparations,
both of bovine origin, were assessed: Thrombin Topical
(Panke, Davis) and Thrombin Reagent (Leo Laboratories).
A thrombin/fibrinogen curve, prepared for each batch,
showed the thrombins to be equal in their power to clot
fibrinogen. The results of tests for contamination with
components of the fibrinolytic system are given in Table I.
These show that each of the two batches of Thrombin
Topical tested contained contaminants likely to influence
the results of tests of fibrinolytic activity; this has also
been found by other workers (McNicol, 1962; Innes,
1962). Only one batch of Thrombin Reagent was avail-
able: this was found to be remarkably free from contami-
nants and this fact, together with an assurance from the
manufacturers that this batch will be used as a refer-
ence standard for all future batches, leads us to recom-
 mend the Leo preparation for use in the standard clot.

Thrombin can be stored for up to two months at
-35°C. In measured aliquots of a solution containing
500 NIH. units/ml. saline in plastic containers. For use
it is diluted 1:10 with saline phosphate buffer; this
solution will remain active for eight hours in an ice-bath.

It is important to use enough thrombin to convert all
the fibrinogen present in the system rapidly to fibrin. If
conversion is incomplete or if there is time for significant
fibrinogenolysis to occur, the final amount of fibrin substrate in the test will be reduced. By using un
taminated thrombin it is permissible to add an excess to the
standard clot system, thus ensuring complete and
rapid fibrin formation. Thrombin Reagent is best used
in a concentration of 50 N.I.H. units/ml and 0-1 ml of
this solution will bring about clotting in a 1-1 ml.
standard clot in less than 30 seconds.

PLASMINOGEN
Although it is stable in blood, pure plasminogen is difficult and expensive to prepare and is
unstable. It is present in Cohn plasma fraction III but,
unfortunately, human fraction III is not readily available
in Great Britain. Of the commercial preparations tested,
Kabi human plasminogen was found to be partially
activated to plasmin. The purity of the Merck Sharp &
Dohme preparation would make it ideal for use in the
standard clot but it is very unstable in solution and must
be kept on ice: it is best used as 0-1 ml of a solution
containing 1 mg. plasminogen/ml. saline phosphate
buffer. Using a concentrated solution the marked
instability can be ignored for the period of the experiment.

Human serum euglobulin, prepared from pooled plasma taken from time-expired banked acid-citrate-
dextrose blood, is a satisfactory source of plasminogen available in all clinical laboratories (Appendix 4). Serum,
rather than plasma, is used to preclude inaccuracy due to
the presence of further fibrinogen altering the final
centralization of substrate: antiplasmins remain in the
supernatant during euglobulin precipitation and can be
rejected, and although anti-activators formed during the
collection of blood (Flute, 1960) are precipitated with the
euglobulin, their effect can be diluted out. Some activa-
tor activity persists (Table I) but this is insufficient to
lyse a standard clot in 12 hours and can thus be regarded
as insignificant. Aliquots of the euglobulin, re-dissolved
in saline phosphate buffer, can be stored at −35°C. in
plastic containers: full activity is retained for at least
three months.

Standardization of human serum euglobulin
Serial dilutions of stock euglobulin solution are incorporated in the
standard clot as follows:—

| Euglobulin + buffer ........... | 0-7 |
| Fibrinogen ..................... | 0-2 | 1-1 ml. |
| Streptokinase ................. | 0-1 |
| Thrombin ...................... | 0-1 |

(For details of reagents see Appendix 1.)

Typical lysis results are given in Table IV: the dilution
producing the shortest lysis time is selected for use. A
solution calculated to contain the required amount of
plasminogen in 0-1 ml is prepared by dilution of the
stored stock solution. Unlike purified plasminogen, this
solution will remain stable for eight hours in an ice-bath,
and shows no tendency towards autoactivation.

PLASMINOGEN ACTIVATORS
The activators of plasmino-
gen most generally used in laboratory estimations are
streptokinase and urokinase. At present streptokinase is
the cheaper and more readily available.

Streptokinase Several commercial preparations are
available, each of which is suitable for use in the standard
clot, provided that standardization is carried out. In the
present study Varidase (Lederle) has been used: this is
most satisfactory as long as precautions are taken to
prevent deterioration of prepared solutions.

Varidase is supplied as freeze-dried material in vials
containing 100,000 manufacturer’s units. For the pre-
paration of a stock solution 10 ml of ice-cold normal
saline is added to the vial giving a solution containing
10,000 units per ml. After mixing, the streptokinase
solution is distributed in measured aliquots into plastic
tubes which are immediately frozen at −35°C. One tube
is retained for standardization which is carried out as
soon as possible. Each vial must be standardized, but the
stock solution frozen at −35°C. has been found to
retain its full activity for at least one month.

Standardization All dilutions are made in gel buffer,

| pH 7-5, and all solutions kept in an ice-bath. Dilutions

| are prepared as follows:—

<table>
<thead>
<tr>
<th>Streptokinase (100 manufacturer’s units/ml.)</th>
<th>Gel Buffer (ml.)</th>
<th>Final Strength of Streptokinase (manufacturer’s units/ml.)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1-0</td>
<td>4-0</td>
<td>20</td>
</tr>
<tr>
<td>0-75</td>
<td>4-25</td>
<td>15</td>
</tr>
<tr>
<td>0-5</td>
<td>4-5</td>
<td>10</td>
</tr>
<tr>
<td>0-25</td>
<td>4-75</td>
<td>5</td>
</tr>
<tr>
<td>0-05</td>
<td>4-95</td>
<td>1</td>
</tr>
</tbody>
</table>

Each dilution is tested in the following standard clot
mixture:

| Saline phosphate buffer .................... | 0-6 |
| Euglobulin ................................ | 0-1 |
| Fibrinogen ................................ | 0-2 | 1-1 ml. |
| Streptokinase dilution ................... | 0-1 |
| Thrombin .................................. | 0-1 |

(For details of reagents see Appendix 1.)

The lysis time in minutes is plotted against the con-
centration of streptokinase on double logarithmic graph
paper. This will give a straight line. The dilution giving
a lysis time of 10 min. is read from the graph; by

TABLE IV

<table>
<thead>
<tr>
<th>Saline Phosphate Buffer (ml.)</th>
<th>Serum Euglobulin (ml.)</th>
<th>Streptokinase (10 units/ml.)</th>
<th>Standard Fibrinogen (ml.)</th>
<th>Standard Thrombin (ml.)</th>
<th>Lysis Time (min.)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>0-7</td>
<td>0-1</td>
<td>0-2</td>
<td>0-1</td>
<td>17</td>
</tr>
<tr>
<td>0-2</td>
<td>0-5</td>
<td>0-1</td>
<td>0-2</td>
<td>0-1</td>
<td>14</td>
</tr>
<tr>
<td>0-4</td>
<td>0-3</td>
<td>0-1</td>
<td>0-2</td>
<td>0-1</td>
<td>12</td>
</tr>
<tr>
<td>0-5</td>
<td>0-2</td>
<td>0-1</td>
<td>0-2</td>
<td>0-1</td>
<td>11</td>
</tr>
<tr>
<td>0-6</td>
<td>0-1</td>
<td>0-1</td>
<td>0-2</td>
<td>0-1</td>
<td>11</td>
</tr>
<tr>
<td>0-65</td>
<td>0-05</td>
<td>0-1</td>
<td>0-2</td>
<td>0-1</td>
<td>11</td>
</tr>
<tr>
<td>0-675</td>
<td>0-025</td>
<td>0-1</td>
<td>0-2</td>
<td>0-1</td>
<td>&gt;12 hours</td>
</tr>
</tbody>
</table>

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definition this will contain 1 ‘standard clot’ unit of streptokinase in 0·1 ml. (to be distinguished from the manufacturer’s unit). This dilution is then prepared for use from the stock solution.

It is particularly important that dilutions should be made in gel buffer and kept at all times in an ice-bath. If these conditions are strictly observed the solutions will remain stable for up to four hours.

Urokinase This is a suitable activator for use in all tests except those where proactivator is to be estimated. It is obtainable from Leo Pharmaceutical Products in ampoules containing 25,000 Ploug units. Standardization may be carried out as for streptokinase, and each new vial must be standardized. The material is more stable than streptokinase and solutions kept in an ice-bath retain full activity for eight hours. Stock solutions containing 10,000 units per ml. of saline should be stored at —35°C.

APPLICATIONS OF STANDARD CLOT METHOD

The purpose of the standard clot method is not to supplant biochemical standardization, which is necessarily a highly technical commitment, but to augment the facilities of the average clinical laboratory. Its principal virtue is the ease with which the basic method can be adapted to measure the activity of plasmin, antiplasmin, activator, or anti-activator (the latter including the antagonistic effect of anti-streptococcal antibodies). It is also possible to measure plasminogen concentration but as this involves the use of a different fibrinogen preparation it will be reported elsewhere.

ACTIVATOR ACTIVITY ASSAY The method is not suitable for the measurement of intrinsic plasma activator because for this it is necessary to remove all naturally occurring inhibitors from the test plasma. It is, however, very useful for estimating the activity of artificial activators; this is carried out as described for the standardization of streptokinase. The method can also be used for assessing the activity of tissue activator which does not normally occur in association with inhibitors. By the construction of a graph, as described, the amount of activator which will cause the lysis of a 1·1 ml. standard clot containing excess plasminogen can be determined; this is defined as one standard clot unit of activator.

ANTI-ACTIVATOR ASSAY Inhibition of plasminogen activation may be brought about by two different mechanisms. Blood itself is able to exert an inhibitory effect on activators, which is probably equally potent against the intrinsic (plasma) activator and such artificial activators as streptokinase and urokinase. This inhibition by blood is not intrinsically present in the circulation, but is developed rapidly in shed blood, the rate of development probably being dependent upon surface contact, temperature, and the presence of platelets and calcium (Flute, 1960, 1962). The other form of inhibition against plasminogen activation to be found in whole blood is due to the presence of antibodies developed as a result of stimulation by heterologous protein antigens: the most important of these in humans is the anti-streptokinase antibody resulting from incidental streptococcal infections. The urokinase used so far has always been of human origin and anti-urokinase antibodies have not been reported in man, although human urokinase is probably antigenic to other species (Celander and Guest, 1960). The inhibitory effect of antibodies is not influenced by those factors governing the production of the blood anti-activator but is present in the circulating blood: by varying the conditions of collection of the blood sample it is therefore possible to assay the inhibitory effect of antibody alone or the total inhibition due to antibody plus blood inhibitor. Measurement of antibody activity is carried out on citrated, platelet-poor plasma collected and prepared with a meticulously observed cold technique, using plastic apparatus in order to prevent activation of the blood inhibitor. For the assay of total inhibition the test sample is collected and prepared under conditions which give maximum opportunity for activation of the inhibitor.

An important application of this method is to assess the inhibition which must be overcome before thrombolytic treatment with infusion of activator can become effective. Such inhibition is due to antibody against the activator being used; the demonstrable inhibitor in shed blood can be ignored because it is not active in the circulation. If urokinase is the activator of choice antibodies will not be present but when streptokinase is used, interference by antibodies is inevitable. Any assay must be carried out on cold-treated, non-activated, platelet-poor citrated or E.D.T.A. plasma, in order to calculate the initial dose of streptokinase which will neutralize the antibodies already present.

MEASUREMENT OF ANTI-STREPTOKINASE ANTIBODY CONCENTRATION Venous blood is collected with a plastic syringe which has been cooled to 4°C. (the needle is not cooled). It is placed in a pre-cooled plastic tube containing 1/10 volume of 3·8% trisodium citrate or 1/100 volume of 10% E.D.T.A. and immediately centrifuged at 3,000 r.p.m. (3,700 g) for 10 minutes at 4°C. The platelet-poor plasma is transferred to a second polystyrene tube in an ice-bath by means of a polystyrene Pasteur pipette, and the test set up immediately.

While the blood is being centrifuged, streptokinase of the type to be used for the treatment is standardized as described previously and a solution containing 10
standard clot units per millilitre prepared. The test plasma is diluted 1 in 5 with saline phosphate buffer and further dilutions are made from this as shown in Table V, all tubes being kept in an ice-bath.

**TABLE V**

<table>
<thead>
<tr>
<th>Tube</th>
<th>Dilution</th>
<th>Test Plasma or Serum 1 in 5 (ml.)</th>
<th>Saline Phosphate Buffer</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>1:1</td>
<td>0:2</td>
<td>—</td>
</tr>
<tr>
<td>2</td>
<td>1:2</td>
<td>0:1</td>
<td>0:1</td>
</tr>
<tr>
<td>3</td>
<td>1:4</td>
<td>0:05</td>
<td>0:15</td>
</tr>
<tr>
<td>4</td>
<td>1:8</td>
<td>0:02</td>
<td>0:2</td>
</tr>
<tr>
<td>5</td>
<td>1:16</td>
<td>0:01</td>
<td>0:2</td>
</tr>
<tr>
<td>6</td>
<td>1:32</td>
<td>0.01 of test 1 in 50</td>
<td>0:2</td>
</tr>
<tr>
<td>7</td>
<td>—</td>
<td>—</td>
<td>0:2</td>
</tr>
<tr>
<td>8</td>
<td>—</td>
<td>—</td>
<td>0:25</td>
</tr>
<tr>
<td>9</td>
<td>—</td>
<td>—</td>
<td>0:25</td>
</tr>
</tbody>
</table>

To tubes 1-7 add 0.1 ml. streptokinase, 10 standard clot units per ml.

To tubes 8 and 9 add 0.05 ml. streptokinase, 10 standard clot units per ml.

All tubes are incubated in an ice-bath for 10 minutes to allow the inhibitor to act. At the end of this time the following components of the standard clot are added to each tube as rapidly as possible:

- Saline phosphate buffer .............. 0.5 ml.
- Plasminogen ................................ 0.1 ml
- Fibrinogen ................................ 0.2 ml
- Thrombin ................................ 0.1 ml

(For details of reagents see Appendix I.)

A stop-watch is started as the thrombin is added and the tubes shaken and transferred immediately to a 37°C water-bath. They are examined at 30-second intervals and the lysis times recorded. Tube no. 7, containing 1 standard clot unit of streptokinase, should lyse in 10 minutes and is the streptokinase control.

**MEASUREMENT OF TOTAL INHIBITION OF ACTIVATOR**

Venous blood is collected with a glass syringe and placed in a glass tube. The blood is allowed to clot at 37°C and incubated at this temperature for a further 20 minutes. Serum is obtained by centrifugation and stored in a glass tube in an ice-bath until used.

The test is set up in the same way as for the assay of antibody activity. Inhibition against other extrinsic and artificial activators can be estimated by their inclusion in the test system instead of streptokinase.

**INTERPRETATION OF RESULTS**

The lysis time of the clots in tubes 1-6 is plotted against the dilution of test plasma or serum on double logarithmic graph paper. By reference to this curve the dilution of test giving a lysis time corresponding to the lysis time given by the 1 unit control (tubes 8 and 9) is found. This dilution multiplied by 5 (for original test dilution) gives the number of inhibition units per millilitre of the test. 1 unit of inhibition being defined as the amount which will inhibit 50% of the activator activity in a standard clot. From the result of the streptokinase antibody test and knowledge of the patient’s plasma volume, the total amount of antibody present can be calculated and the initial dose of streptokinase prescribed accordingly (Johnson and McCarty, 1959).

**PLASMIN ACTIVITY ASSAY**

The standard clot method is not used for estimating plasma plasmin activity because, as for the estimation of endogenous activator, inhibitors must be removed to unmask the fibrinolytic effect. It could be used to estimate fibrinolytic activity in pathological fibrinolytic states where the inhibitor activity has been overcome, but other tests suitable for this are available. The principal value of the standard clot method in this respect is in the assay and standardization of proprietary brands of plasmin: it should be noted that most of these are prepared by the addition of activator—usually streptokinase—to plasminogen and therefore inevitably contain some free activator (Baumgarten et al., 1960). Unless a plasminogen-free assay system is used, the rate of lysis of the standard clot will be a measure of the fibrinolytic potential rather than a direct function of the plasmin added.

One standard clot unit of plasmin may be defined as the amount which will lyse a standard clot in 10 minutes. Dilutions of the test material are made in saline-phosphate buffer to contain 500, 100, 50, 40, 30, 20, 10, 1, and 0.1 manufacturer’s units. Each dilution is tested in the following standard clot mixture:

Saline phosphate buffer ............. 0.6 ml
Plasminogen* .......................... 0.1 ml
Fibrinogen .......................... 0.2 ml
Thrombin .......................... 0.1 ml

*Plasminogen is replaced by saline phosphate buffer if a plasminogen-free system is being used for the assay of plasmin alone.

The lysis time given by each plasmin dilution is noted. Lysis times are plotted against dilution of test material on double logarithmic graph paper. From the graph the dilution which will lyse a standard clot in 10 minutes is determined; this will contain one standard clot unit of fibrinolytic activity.

**APPENDIX 1**

**BASIC MIXTURE FOR STANDARD CLOT TESTS**

Saline phosphate buffer + test substance(s) ........... 0.6 ml
Plasminogen (pre-standardized human serum euglobulin) ............... 0.1 ml
Fibrinogen (pre-standardization bovine plasma fraction I) ........... 0.2 ml
Actinogen (10 standard clot units/ml) .................. 0.1 ml
Thrombin (Leo Laboratories. Solution containing 50 N.I.H. units/ml) ........... 0.1 ml
Total volume .................. 1.1 ml

**APPENDIX 2**

**METHODS USED FOR THE IDENTIFICATION OF CONTAMINANTS IN THE REAGENTS**

1. **PLASMIN** Heated fibrin plates (Lassen, 1952).
2. **PLASMINOGEN** Heated fibrin plates. Test material mixed with an equal volume of streptokinase (1,000, 100,
A standard clot method for the assay of plasminogen activators, anti-activators, and plasmin

and 10 units/ml.) and human serum (1/100) or human activation product as a source of procactivator (Mullertz and Lassen, 1953; Hawkey, 1963).

3 PLASMINOGEN ACTIVATOR

Unheated fibrin plates and plates prepared from a euglobulin precipitate of bovine plasma fraction I. The euglobulin precipitate contains fibrinogen and plasminogen and plates prepared from it are more sensitive to minor degrees of plasmin activity than are standard unheated fibrin plates because the anti-plasmin level is reduced (Kowalski et al., 1959).

4 PROACTIVATOR

Unheated fibrin plates. Test material mixed with an equal volume of streptokinase (1,000, 100, and 10 units/ml).

5 ANTI-ACTIVATOR

Modified standard clot method as described in the text. When fibrinogen was the test material it was first clotted with thrombin and the fibrin clot removed with a wooden applicator stick, care being taken to squeeze out as much as possible of the liquid trapped in the clot. This was done in order that the presence of extra fibrinogen would not influence the final result of the test.

6 ANTI-PLASMINS

Modification of the method described by Howell (1963) using a more dilute plasmin solution. Fibrinogen solutions were treated as for the measurement of anti-activators.

All fibrin plates were prepared from bovine plasma fraction I (Armour) and Thrombin Reagent (Leo). Tests were set up in triplicate and repeated at least twice. Appropriate controls were carried out in each case. Results obtained on fibrin plates were confirmed by the standard clot method.

APPENDIX 3

BUFFERS

Saline phosphate buffer has been used in the tests described in this paper. It removes ionic calcium which exerts an inhibitory effect on the fibrinolytic system.

**SALINE-PHOSPHATE BUFFER**

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<tr>
<th>Component</th>
<th>Quantity</th>
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<tbody>
<tr>
<td>KH2PO4</td>
<td>13.6 g</td>
</tr>
<tr>
<td>NaCl</td>
<td>18.0 g</td>
</tr>
<tr>
<td>Merthiolate</td>
<td>200 mg</td>
</tr>
<tr>
<td>5 N NaOH</td>
<td>18 ml</td>
</tr>
</tbody>
</table>

Make up to 2,000 ml. with distilled water. Adjust to pH 7.5. This buffer is used to dissolve all reagents except the streptokinase.

**GEL BUFFER**

<table>
<thead>
<tr>
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<th>Quantity</th>
</tr>
</thead>
<tbody>
<tr>
<td>KH2PO4</td>
<td>27.2 g</td>
</tr>
<tr>
<td>NaCl</td>
<td>20.0 g</td>
</tr>
<tr>
<td>Gelatin (Davis)</td>
<td>10 g</td>
</tr>
<tr>
<td>Merthiolate</td>
<td>200 mg</td>
</tr>
<tr>
<td>5 N NaOH</td>
<td>35 ml</td>
</tr>
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</table>

Make up to 2,000 ml. with distilled water. Adjust to pH 7.5. This buffer is used for the preparation of streptokinase dilutions, and gelatin has been added because the provision of a gel serves to stabilize the streptokinase during the time taken to complete the procedure.

APPENDIX 4

PREPARATION OF HUMAN SERUM EUGLOBULIN AS A SOURCE OF PLASMINOGEN

Human serum euglobulin is prepared from time-expired bank blood in the following way. Citrated (A.C.D.) plasma from 10 donors is pooled; the plasma may have been stored at −35°C if necessary. One thousand N.I.H. units of Leo thrombin reagent are added per 120 ml. plasma, and the plasma incubated at 37°C for 30 minutes to ensure complete conversion of fibrinogen to fibrin. The fibrin clot is removed and discarded, as much liquid as possible being squeezed out beforehand. One part of serum is added to 19 parts of glass-distilled water at 4°C. and the pH adjusted to 5.3 by the addition of 0.2 parts of 1% acetic acid. The mixture is left at 4°C for 30 minutes to allow for complete precipitation of the euglobulins. The precipitate is obtained by centrifugation at 1,500 r.p.m. (1,000 g) for 15 minutes, and redissolved to one quarter of the original volume of serum in saline phosphate buffer. The euglobulin solution is stored in aliquots in plastic containers at −35°C and must be standardized before use.

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A standard clot method for the assay of plasminogen activators, anti-activators, and plasmin

Christine M. Hawkey and J. L. Stafford

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