Odontogenic myxoma: Ultrastructural and histochemical studies

J. D. HARRISON

From the Department of Oral Pathology, King’s College Hospital Dental School, London

SYNOPSIS  An odontogenic myxoma of a maxilla has been examined. Histochemistry of the mucosubstance indicated that hyaluronic acid and chondroitin sulphate were present. On ultrastructural examination many of the cells in the myxomatous tissue were seen with prominent rough endoplasmic reticulum, suggesting a secretory function, and possibly the myxomatous ground substance was produced by these cells. Cells containing collagen fibres were found. Epithelial islands with intercellular spaces producing an arrangement similar to the stellate reticulum of the enamel organ were found, and possibly had developed from odontogenic epithelial rests. These islands were surrounded by a clear zone, outside which were cells with an increased prominence of adenosine-triphosphatase, nucleoside-diphosphatase, thiamine-pyrophosphatase, and often arylamidase reaction products. This probably represents a reaction of the mesodermal tissue to the epithelial islands. Possibly the epithelium exerts an inductive effect on the mesodermal tissue, and the myxomatous appearance may be the result of an aberrant development of mesodermal cells into ‘myxoblasts’ which secrete the myxomatous ground substance.

A considerable number of odontogenic myxomas have been studied by means of routine histological techniques (Zimmerman and Dahlin, 1958; Barros, Dominguez, and Cabrini, 1969; Gorlin, 1970; Pindborg, 1970; Whitman, Stewart, Stoopack, and Jerrold, 1971; Cawson, 1972). Two more specialized studies have been a thorough study of the chemistry and histochemistry of the mucosubstance by Hodson and Prout (1968) and a histochemical study by Sedano and Gorlin (1965), both these studies being primarily of single cases. In the present investigation a maxillary tumour which was found on preliminary biopsy to be an odontogenic myxoma has been studied after removal by means of routine histological techniques, enzyme and mucosubstance histochemistry, and electron microscopy.

Materials and Methods

The tumour occupied the right maxilla of an otherwise fit woman of 21 years who had noticed swelling of the right face for at least three years. The case history will be reported elsewhere (Harrison and Eggleston, 1973). Upon removal of the tumour a pyramid of tissue extending to the centre of the tumour was removed. The cut surface was white and the tissue was of a rubbery consistency. The apical part of this pyramid was preserved in several ways.

MUCOSUBSTANCE HIStOChemistry AND ROUTINE HISTOLOGY

A piece of tissue was fixed overnight at 0-4° in 10% formalin with 0-9% saline and 2% calcium acetate and was subsequently wax-embedded. Sections were cut at 5 μ.

Mucosubstances containing vic-glycols were demonstrated by the periodic-acid Schiff technique (Stoward, 1967, but with Schiff’s reagent from G. T. Gurr). Glycogen was detected by the disappearance of its staining with the periodic-acid Schiff technique after incubating sections for one hour at 37° in an aqueous solution of 0-1% malt diastase (Sigma).

Acidic mucosubstances were demonstrated by staining with 0-05% Alcian Blue 8GX at pH 2-5 (Mowry, 1956) or by the Alcian Blue critical electrolyte concentration technique at pH 5-8 with 0-05, 0-2, 0-4, 0-6, or 0-9 M MgCl₂ (Scott and Dorling, 1965).

Some sections were incubated with 0-1% staphylococcal hyaluronidase (Organon Laboratories Ltd)
Odontogenic myxoma: ultrastructural and histochemical studies

571
to remove hyaluronic acid (Linker, Meyer, and Hoffman, 1956; Linker, Hoffman, Meyer, Sampson, and Korn, 1960; Spicer, Horn, and Leppi, 1967) or with 0-1% bovine testicular hyaluronidase (type I, Sigma) to remove chondroitin sulphates A and C and hyaluronic acid (Linker et al, 1960; Spicer et al, 1967). The enzymes were dissolved in normal saline and the incubations were for three hours at 37°. Control sections were incubated in normal saline. These sections were then stained either by Alcian Blue at pH 2-5 or at pH 5-8 with 0-05 M MgCl₂. The removal of stainable material by the enzymes was observed by comparing sections stained after enzyme treatments with those stained after treatment with saline alone.

For histological examination, sections were stained with haematoxylin and eosin or the van Gieson technique for collagen (Fullmer, 1965).

The hyaluronidase digestions and staining with Alcian Blue at pH 2-5 were also applied to the following tissues: a myxoma removed from the gingiva associated with the lower first and second molars of a woman of 35 years; a myxoma associated with the follicle of an unerupted upper canine in a girl of 10 years; an odontogenic fibroma situated on an edentulous lower alveolus in a woman of 68 years; two umbilical cords; and buccal salivary glands.

ENZYME HISTOCHEMISTRY

Pieces of tissue of about 3 mm diameter were fixed for 16 hours at 0-4° in 4% formaldehyde (freshly prepared from paraformaldehyde) in 0-08 M cacodylate buffer pH 7-2 with 7-5% sucrose and 0-05% calcium chloride, and were then rinsed in 0-05 M cacodylate buffer pH 7-2 with 7-5% sucrose for about 60 hours at 0-4° before quenching in hexane cooled by solid carbon dioxide. This tissue will be referred to as 'prefixed' tissue. Other pieces of tissue were quenched upon removal without prior fixation. The quenched pieces were stored at -70°.

Cryostat sections of about 10 to 12 μ were mounted on slides or coverslips. The following enzyme histochemical techniques were applied to sections of prefixed tissue and to sections of unfixed tissue which had been fixed in the formol-sucrose fixative for five minutes at 0-4° (postfixed tissue): adenosine triphosphatase using a modification of the technique of Wachstein and Meisel (1957) with 1-8 mM lead nitrate and 20 mM magnesium sulphate; nucleoside diphosphatase using inosine diphosphate (Smith and Farquhar, 1970); thiamine pyrophosphatase using similar concentrations to those used by Smith and Farquhar (1970); alkaline phosphatase (Stutte, 1967); acid phosphatase (azo-dye and metal-precipitation techniques of Barka and Anderson, 1962); beta-glucuronidase (Hayashi, Nakajima, and Fishman, 1964); non-specific esterase (Davis, 1959) with either 0-03 mM eserine to inhibit cholinesterase activity (Koelle, 1955) or 0-01 mM E600 to inhibit organophosphate-sensitive esterases (Bulmer and Fisher, 1970); and peroxidase (Graham and Karnovsky, 1966). Arylamidase activity (Patterson, Hsiao, and Keppel, 1963) was demonstrated in prefixed and unfixed sections by a technique originally considered specific for leucine-aminopeptidase (Nachlas, Monis, Rosenblatt, and Seligman, 1960). Techniques for the following enzymes were applied to unfixed sections: cytochrome oxidase (Burstone, 1961); succinic dehydrogenase (Nachlas, Tsou, de Souza, Cheng, and Seligman, 1957); and NADH- and NADPH-diaphorases using NADH and NADPH (as in the method of Scarpelli, Hess, and Pearse, 1958) and nitro-BT (as suggested by Nachlas et al, 1957).

ELECTRON MICROSCOPY

Pieces of tissue of about 1 mm diameter were immersed for two hours at room temperature in a fixative consisting of 4% acrolein, 5% glutaraldehyde, and 4% formaldehyde (freshly prepared from paraformaldehyde) in 0-08 M cacodylate buffer pH 7-2 with 0-05% calcium chloride (based on Karnovsky, 1965, and Rodriguez, 1969). The pieces were rinsed, osmicated, dehydrated, and embedded in araldite. Ultrathin sections were examined in an electron microscope after staining with uranyl acetate and lead citrate or with lead citrate alone. Thicker sections were stained with Toluidine Blue and examined light microscopically.

Results

HISTOLOGY

The bulk of the tumour was composed of a faintly basophilic myxomatous ground substance in which cells of various size and shape were dispersed (figs 1, 2, 3, and 4). A variable number of fibres was found coursing throughout the ground substance (figs 1, 2, 3, and 4). 'Cavities' with a poorly defined lining of fibres were found (fig 2). Thin-walled vessels, occasionally containing red blood corpuscles, were frequently associated with 'cavities', either within or adjacent to them (fig 2).

Epithelial islands were found at intervals throughout the tissue (figs 1, 2, 3, and 4). In some islands no intercellular spaces were seen (fig 1), whilst in most there were conspicuous intercellular spaces (figs 2, 3, and 4). The islands with intercellular separation were surrounded by a clear zone relatively poor in fibres and cells (figs 2, 3, and 4). Such a zone was not usually seen around islands without intercellular separation (fig 1).
Fig 1  Section shows myxomatous tissue and an epithelial island with negligible intercellular separation. Haematoxylin and eosin, × 410.

Fig 2  Section shows myxomatous tissue and several thin-walled vessels (V), one being within a cavity. An epithelial island is seen with considerable intercellular separation, and it is surrounded by a clear zone relatively poor in fibres and cells. Haematoxylin and eosin, × 180.
Odontogenic myxoma: ultrastructural and histochemical studies

Fig 3  Light-microscopical araldite section adjacent to section viewed electron-microscopically in figure 16. There is some intercellular separation in the epithelial island, which is surrounded by a clear zone. Bundles of fibres are elsewhere conspicuous in the myxomatous tissue. Toluidine Blue, × 360.

Fig 4  Light-microscopical araldite section adjacent to section viewed electron-microscopically in figure 17. There is considerable intercellular separation in parts of the epithelial island, which is surrounded by a clear zone. Few fibres are present in the myxomatous tissue. Toluidine Blue, × 460.

MUCOSUBSTANCE HISTOCHEMISTRY
With the periodic-acid Schiff technique there was faint diastase-resistant staining of the sections. Glycogen was demonstrated in many cells in the myxomatous tissue and in many cells in the epithelial islands.

With the Alcian Blue critical electrolyte concentration technique the sections were stained moderately with 0-05 M MgCl₂, weakly with 0-2 M, very weakly with 0-4 M, faintly with 0-6 M, and were not stained with 0-9 M. Mast cells were not seen.

Sections were moderately stained by Alcian Blue at pH 2-5 (fig 5). When sections were incubated in saline alone or with the hyaluronidases before staining with Alcian Blue at pH 2-5, it was found that there was a slight decrease in staining after saline alone (fig 6), a greater decrease after staphylococcal hyaluronidase (fig 7), and an absence of staining after testicular hyaluronidase (fig 8). After the various incubations and staining with Alcian Blue at pH 5-8 with 0-05 M MgCl₂ there were no unequivocal decreases in staining.

Similar results were achieved when sections of the myxoma associated with the gingiva and of the two umbilical cords were subjected to the incubations and staining with Alcian Blue at pH 2-5. However, the myxomatous material associated with the unerupted canine showed weak staining with Alcian Blue at pH 2-5, a slight increase after incubating in saline, a slight decrease after staphylococcal hyaluronidase when compared with the saline-treated sections, the intensity being similar to that of untreated sections, and a complete loss of staining after testicular hyaluronidase. In the
Fig 5  Untreated section stained with Alcian Blue at pH 2·5. Moderate staining is seen throughout. × 160.

Fig 6  Adjacent section to fig 5, incubated in saline alone before staining with Alcian Blue at pH 2·5. There is some reduction in staining when compared with figure 5. × 160.

Fig 7  Adjacent section to fig 6, incubated with staphylococcal hyaluronidase before staining with Alcian Blue at pH 2·5. There is a considerable reduction in staining when compared with figure 6. × 160.

Fig 8  Adjacent to fig 7, incubated with testicular hyaluronidase before staining with Alcian Blue at pH 2·5. There is no staining. × 160.
Odontogenic myxoma: ultrastructural and histochemical studies

odontogenic fibroma only the faintest trace of stain was seen with Alcian Blue at pH 2.5, and the intensity of staining was unaffected by saline or staphylococcal hyaluronidase, whilst no stain was seen after testicular hyaluronidase. The staining of the mucin in the buccal salivary glands was not affected by the various incubations.

ENZYME HISTOCHEMISTRY

The distribution of reaction products from the techniques for adenosine triphosphatase, nucleoside diphosphatase, and thiamine pyrophosphatase was similar. Reaction product was seen in many cells in the myxomatous tissue and was usually faint or weak, although some moderately stained cells were seen, and such cells were concentrated around epithelial islands with intercellular separation, a clear zone usually separating these cells from the island (fig 9). Faint staining of the cells in the islands was often seen.

Faint alkaline-phosphatase activity was infrequently seen outlining cells in the myxomatous tissue and rarely outlining cells in epithelial islands. Activity was not seen in the cells surrounding islands.

Intense acid-phosphatase, beta-glucuronidase, and non-specific esterase activities were seen in some cells in the myxomatous tissue, and these were frequently arranged around vessels (fig 10). Strong activities were demonstrated in foamy macrophages, and these were found in 'cavities' (fig 11). Activity was not seen in cells surrounding epithelial islands. In the cells of the epithelial islands, faint acid-phosphatase activity was sometimes seen, faint non-specific esterase activity was rarely seen, and beta-glucuronidase activity was not seen. The non-specific esterase activity appeared to be completely E600-resistant, except in the foamy macrophages, where it was entirely, or almost entirely, E600-sensitive.

Moderate arylamidase activity was seen in cells which appeared to correspond to those with intense acid-phosphatase, beta-glucuronidase, and non-specific esterase activities. Activity was seen in other cells in the myxomatous tissue and was usually faint, such cells often being seen around epithelial islands with intercellular separation in a similar distribution to the cells with moderate adenosine-triphosphatase, nucleoside-diphosphatase, and thiamine-pyrophosphatase staining. Faint activity was occasionally seen in cells in the islands.

Peroxidase, cytochrome-oxidase, succinic-dehydrogenase, and NADH- and NADPH-diaphorase activities were not demonstrable. Control sections of a human submandibular gland were

![Fig 9](http://jcp.bmj.com/)...Section incubated in the inosine-diphosphatase medium for one hour at room temperature. The nuclei are counterstained with methyl green. Moderately stained cells are concentrated around an epithelial island, and are separated from it by a peri-insular clear zone. Several moderately stained vessels (V) are present. × 70.

![Fig 10](http://jcp.bmj.com/)...Section incubated in the non-specific esterase medium with eserine for 10 min at room temperature. Cells with intense activity and sections of a thin-walled vessel (V) with some granules of reaction product in the walls are seen. × 300.
incubated in the media for the respiratory enzymes and strong activity was found in the ducts.

**ELECTRON MICROSCOPY**

Sections were cut through seven pieces of tissue and were examined by light microscopy in order to find epithelial islands and 'cavities'. On the basis of these findings, sections at several different levels in four pieces were cut for electron-microscopical examination.

The ground substance of the tumour consisted of amorphous material of low electron density and variable numbers of particles of high electron density (fig 12). The size of the particles varied considerably, but they were usually less than 700 Å diameter. Many of the particles had fibrillar projections of a smaller diameter, which were frequently branching. In some regions where there were concentrations of particles, the fibrils were prominent and formed a network communicating with other fibrils and particles.

Variable amounts of banded collagen fibrils with a periodicity of about 660 Å were present. Zones of ground substance were seen with no collagen fibrils whilst in other regions bundles of collagen fibrils were predominant.

On examination of the cells of the tumour a wide spectrum of appearance was found, and classification of cells was often difficult or impossible.

Many of the cells in the myxomatous tissue could be considered as secretory cells on account of prominent rough endoplasmic reticulum. In some the cisternae were narrow, whilst in others they were dilated and contained amorphous material (figs 13 and 14). A well developed Golgi apparatus was often seen (figs 13 and 14). These cells varied considerably in shape and size, cells whose longest dimension was above 20 μ being seen (fig 13).

Several 'cavities' were found in one piece of tissue. They contained a considerable amount of amorphous material which was continuous with that in the surrounding ground substance, but electron-dense particles were seldom seen. The 'cavities' were surrounded by collagen fibrils which were intermixed with the contents at the periphery (figs 13 and 14). Cells which were considered as secretory cells (figs 13 and 14), cell debris, and macrophages filled
Fig 13 Electron micrograph showing a large cell containing prominent rough endoplasmic reticulum with some dilated cisternae. Several Golgi apparatuses (G) are present. The cell is situated at the periphery of a cavity which contains a considerable amount of amorphous material. Collagen fibrils are seen surrounding the cavity at the top right, and some are seen in the cavity. Uranyl acetate and lead citrate, × 6000.

Fig 14 Electron micrograph showing a cell with prominent rough endoplasmic reticulum with dilated cisternae containing amorphous material. A prominent Golgi apparatus is seen (G). The cell is at the periphery of a cavity, and amorphous material, collagen fibrils, and a little membranous debris are seen. Uranyl acetate and lead citrate, × 12 500.
with membrane-bound granules of varying density, which appeared to correspond to light-microscopical foamy macrophages, were seen in the 'cavities'.

Cells containing banded collagen fibrils with a periodicity of about 660 Å were seen in the myxomatous tissue (fig 15). The fibrils were usually in bundles, and were present in membrane-bound structures, which often contained amorphous material and occasionally electron-dense debris. Communication between these structures and the exterior was occasionally seen. These cells possessed a variable, but usually small, amount of rough endoplasmic reticulum, sometimes with moderately dilated cisternae containing amorphous material.

Three large epithelial islands were examined (figs 16 and 17). In one of these (fig 16) and in parts of the other two (fig 17), the intercellular spaces were small or moderately sized and usually appeared empty apart from a little amorphous material and membranous debris. Particles were not seen in these spaces. In parts of two of the islands (fig 17) there was considerable separation between epithelial cells with ground substance extending throughout large intercellular spaces. In some places there was loss of continuity of the peripheral layer of epithelial cells, and the basement membrane was seen separating large intercellular spaces from the peri-insular region, although occasionally the basement membrane was incomplete with communication between large intercellular spaces and the peri-insular region. Invaginations of the surrounding tissues lined by basement membrane were seen in regions with little intercellular separation (figs 16 and 17).

The epithelial cells contained little endoplasmic reticulum, usually with diluted empty cisternae and few attached ribosomes. Golgi apparatuses were seldom seen and were small. Many of the cells contained tonofilaments. Variable quantities of glycogen granules were seen. In the regions with little or moderate intercellular separation desmosomes and microvilli were seen. A clear zone relatively poor in fibres and cells and containing ground substance was seen around the islands, particularly where there was much intercellular separation (figs 16 and 17).
Odontogenic myxoma: ultrastructural and histochemical studies

Fig 16  Electron micrograph of adjacent section to figure 3. An epithelial island is seen with small, and some moderately sized, intercellular spaces which are largely empty. A basement membrane surrounds the island and lines invaginations (I). A clear zone relatively poor in collagen fibrils and cells and containing ground substance is seen around the island. Lead citrate, × 1800.

Discussion

Hodson and Prout (1968) found that of the total mucopolysaccharide present in a myxoma of the mandible 80% was hyaluronic acid and 20% chondroitin sulphate, and they suggested that a high hyaluronic acid content may be a characteristic feature of myxomas. In the three myxomas of the present investigation, the reduction in staining with Alcian Blue at pH 2.5 after staphylococcal hyaluronidase when compared with saline-treated sections and the abolition of staining after testicular hyaluronidase indicate the presence of hyaluronic acid and chondroitin sulphate A and/or C (Linker et al, 1956, 1960; Spicer et al, 1967), whilst in the odontogenic fibroma little chondroitin sulphate A and/or C and no hyaluronic acid were demonstrated.

In two of the myxomas in the present investigation, in the umbilical cords, and in the myxoma studied by Hodson and Prout (1968) incubation in saline alone appeared to cause a loss of acidic mucosubstance from the sections. One feature common to all these tissues appears to be the presence of hyaluronic acid, and possibly some of this is lost during the saline incubation. Although the presence of chondroitin sulphate A and/or C appears to be another feature common to these tissues, the odontogenic fibroma which contained no demonstrable hyaluronic acid but some chondroitin sulphate A and/or C did not show any decrease in staining after incubation in saline. Suggesting that chondroitin sulphate A and/or C is not removed by saline. Hyaluronic acid appeared to be present in the myxoma associated with the unerupted canine, although there was an increase in staining after incubation in saline. Possibly this increase is due to reactive sites on acidic mucosubstances being exposed by the saline incubation. A similar unexplained increase in stain-
ing was seen by Spicer et al (1967) in mouse and rat cornea after incubation in 0·1 M phosphate buffer pH 5·5.

Although a decrease in staining intensity with Alcian Blue at pH 2·5 was seen after the enzyme incubations, it was not seen with Alcian Blue at pH 5·8 with 0·05 M MgCl₂. This retention of staining is probably due to proteins which are weakly negative at pH 5·8 (Scott and Dorling, 1965) still being stained in the medium containing 0·05 M MgCl₂, and masking the loss of mucosubstance. This suggests that the reduction in staining with the critical electrolyte concentration technique between 0·05 M and 0·2 M may not be entirely due to hyaluronic acid, which has a critical electrolyte concentration in the region of 0·1 M MgCl₂ (Scott and Dorling, 1965). However, the staining at 0·4 M which was lost at 0·9 M is probably due to chondroitin sulphate which has a critical electrolyte concentration in the region of 0·5 M MgCl₂ (Scott and Dorling, 1965).

The particles of high electron density seen in the ground substance at the ultrastructural level are somewhat similar in appearance to particles which appeared to contain acidic mucosubstance which was probably chondroitin sulphate found in cartilage matrix by Matukas, Panner, and Orbison (1967).

Hodson and Prout (Hodson and Prout, 1968; Prout and Hodson, 1968) suggested that the cell responsible for the myxoma is a neoplastic mesenchymal cell which is secreting hyaluronic acid in excessive amount, and suggested that the term 'myxoblast' would be suitable to distinguish it, metabolically if not morphologically, from the collagen-producing fibroblast. The ultrastructural findings support the view of Hodson and Prout that an active secretory cell is responsible for the myxoma, for many cells with the characteristics of secretory cells were found. The finding of secretory cells in 'cavities' suggests that an accumulation of secretory material from such cells produces these 'cavities'.

The absence of demonstrable respiratory enzyme activity in the tumour indicates a low level of respiratory activity, and is compatible with the slow growth of the tumour, which had been present at least three years.

The intense acid-phosphatase, beta-glucuronidase, and E600-resistant esterase activities found in some cells are probably lysosomal enzyme activities. Possibly these cells are active macrophages, and the lower intensity of these enzyme activities in foamy macrophages possibly reflects a functional aging as macrophages become filled with engulfed material. A similar observation has been made in other pathological situations in man (J. D. Harrison, in preparation) and in cat (Harrison, 1972). The
Odontogenic myxoma: ultrastructural and histochemical studies

Nylen, 1966) to islands with widely separated cells similar to the stellate reticulum of the enamel organ (Fischlschweiger, Provenza, and Sisca, 1967; Sisca, Provenza, and Fischlschweiger, 1967). These similarities strongly suggest that the islands consist of odontogenic epithelium.

Although the basement membrane usually maintained the outline of the islands where there was loss of continuity of the peripheral cells, it appeared to be missing in some places and continuity between intercellular spaces and the surrounding region was seen. This contrasts with the situation in the human enamel organ where the continuity of the basement membrane is maintained when there is loss of cellular continuity in the outer enamel epithelium in the intermediate bell stage. The basement membrane in such areas is relocated into and through the outer enamel epithelium and extends between stellate-reticulum cells, the invaginations containing dental sac material (Fischlschweiger et al., 1967). The appearance of these invaginations is similar to that of invaginations in regions with little separation between cells in the islands examined ultrastructurally.

Possibly there is a development of epithelial rests to produce a stellate-reticulum arrangement with an eventual loss of continuity between epithelial cells. There may sometimes be a relocation of the basement membrane, whilst at other times there is no relocation and the basement membrane breaks down, allowing an ingress of ground substance which surrounds widely separated epithelial cells.

The epithelial islands with separation between cells appear to produce a reaction in the surrounding tissue. This was manifested by the clear zone surrounding such islands and the increased prominence of reaction product from the techniques for adenosine triphosphatase, nucleoside diphosphatase, and thiamine pyrophosphatase, and often arylsulphatase, in cells around these islands. Although islands with a stellate-reticulum-like arrangement and peri-insular clear zones do not appear to have been previously found in odontogenic myxomas, they are found in ameloblastic fibroma (Pindborg, 1970). Pindborg (1970) suggested that the presence of immature mesodermal tissue in this tumour may be explained as the result of an inductive effect on the part of odontogenic epithelium. The same suggestion may be applied to the odontogenic myxoma in the present investigation, and the myxomatous appearance may be the result of an aberrant development of mesodermal cells into 'myxoblasts'. The finding of similar features in the present odontogenic myxoma and the ameloblastic fibroma supports the concept (Pindborg, 1970) that, rather than distinct and separate types, there is a continuous spectrum of odontogenic tumours.
I gratefully acknowledge the financial support of the Wellcome Trust; the technical assistance of Mr P. S. A. Rowley with the preparation of the material for electron-microscopical examination and with the photographs, of Mr K. J. Davies with the photographs, and of Miss M. R. Atkins with the histological procedures; and the gift of staphylococcal hyaluronidase from Organon Laboratories Ltd. I am grateful to Mr M. Harris for the specimen.

References


Odontogenic myxoma: Ultrastructural and histochemical studies

J. D. Harrison

*J Clin Pathol* 1973 26: 570-582
doi: 10.1136/jcp.26.8.570

Updated information and services can be found at:
http://jcp.bmj.com/content/26/8/570

**Email alerting service**

Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

**Notes**

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/