Measurement of serum folate: experience with 75Se-selenofolate radioassay

IAN JOHNSON, HUGH GUILFORD, AND MICHAEL ROSE

From the Haematology Department, St George's Hospital, Tooting, London SW17 and The Radiochemical Centre, Amersham, Bucks

SUMMARY Three hundred and eighty-two serum samples were assayed for folate content by a standard Lactobacillus casei microbiological assay and by radioassay using the 75Se-selenofolate radioassay kit from the Radiochemical Centre, Amersham. Results showed encouraging concordance. Explanations were discovered for the few disparities. The radioassay procedure is simple and an assay batch can be completed in one working day.

We wish to document our results from measurements of serum folate levels by both microbiological assay and radioassay.

Samples

Three hundred and eighty-two successive samples of clotted blood, submitted over a 25-week period to the haematology laboratory at St George's Hospital for serum folate assay, were separated and stabilised for purposes of radioassay by the addition of about 5 mg of sodium ascorbate per millilitre of serum. Ascorbic acid was added to the samples for microbiological assay. The folate content of these samples was measured both by a radioassay method and by microbiological assay. Each assay was conducted in ignorance of the result obtained by the other method by another operator.

Forty-five blood samples in sequestrene were assayed for their red cell folate content. Samples for microbiological assay were prepared as a 1:10 dilution of whole blood in 1% aqueous ascorbic acid, with a drop of Tween 80 solution. Those samples for radioassay were prepared as a 1:8 dilution in glass-distilled water and, after complete haemolysis, sodium ascorbate was added to a final concentration of 5 mg/ml.

Methods

MICROBIOLOGICAL FOLATE ASSAY

The Lactobacillus casei method of Waters and Mollin (1961) was used for serum folate assay. Red

75Se-selenofolate was added 100 μl of each standard or sample are pipetted into polypropylene assay tubes, diluted with lysine buffer, pH 10.5, and vortex mixed. All tubes are heated for 5 minutes in a boiling water bath to denature endogenous binding protein, and then cooled to ambient temperature. To each clear solution is added 100 μl of a solution of 75Se-selenofolate in a 0.1M phosphate-EDTA-albumin buffer. The radio-selenium label is incorporated into a cysteine moiety which replaces glutamate to produce pteroyl-l-methylselenocysteine 75Se. Then 100 μl of porcine serum in the same buffer is added. Porcine serum provides plentiful and avid folate binding proteins (Mantzos, 1975). The folate binding behaviour of porcine serum is more reproducible than milk or

Received for publication 6 December 1976
milk extracts. After vortex-mixing, the tubes are incubated at ambient temperature for 30 minutes. A suspension of albumin-coated charcoal (200 µl) is added. This charcoal reagent is prepared by mixing 5 parts charcoal with 1 part bovine serum albumin in 10 parts distilled water. After centrifugation the albumin-coated charcoal is resuspended in 0.1M phosphate-EDTA-buffer containing 0.1% bovine albumin. This is desiccated, and the 200 µl of reconstituted charcoal dispensed into each mixture is coated with 5 mg of albumin. The tubes are centrifuged at >1000 g for 15 minutes. The supernates containing protein-bound folate are decanted into counting tubes, leaving 'free' folate adsorbed on the charcoal.

The radioactivity in each counting tube is quantified in a gamma counter set for selenium-75. A 5-point standard curve is constructed by plotting counts for each of the duplicate standards against the N5-methyltetrahydrofolate concentration of each. The folate level of each of the unknown samples can be interpolated from this curve.

These reagents, and detailed protocol for their use, can be obtained in kit form from The Radiochemical Centre, Amersham.

Results

REPRODUCIBILITY
Human serum controls based on N5-methyltetrahydrofolate were included in all the assays in this study. The between-assays variation for these sera was as follows (7 batches of kits; 2 operators):

- Serum (a) Mean value (n = 16) 3.4 µg/l, CV = 6.6%
- Serum (b) Mean value (n = 14) 6.0 µg/l, CV = 12.7%

The within-assay variation on 10 replicates, mean value 3.37 µg/l, was 4.0%. (This sample was not one of the controls used to determine inter-assay variation.)

SERUM FOLATES
The distribution of values from these two methods is shown in the correlation diagram in Figure 1. The correlation coefficient is 0.82. A few of these points show a significant disparity between the biological and radio assay. Of these, the ringed results were from subjects who, upon enquiry, were discovered to be receiving either antimicrobial or cytotoxic preparations at the time of sampling. The bioassay result would thus be expected to be falsely

![Graph](https://example.com/graph.png)

**Fig. 1** Results of serum folic acid assay by radioisotope dilution method and by microbiological method from 382 samples. The ringed results were on samples known to be from subjects taking broad-spectrum antibiotics.
Measurement of serum folate: experience with $^{75}$Se-selenofolate radioassay

647

depressed. The correlation coefficient for duplicate samples upon radioassay ‘in-batch’ is 0.97.

RED CELL FOLATES

The correlation between bioassay and radioassay values for a small number of samples run in a preliminary study of red cell folate determination using the radioassay are shown in Fig. 2; the correlation coefficient is 0.64, which clearly leaves much to be desired. The correlation coefficient for duplicate samples upon radioassay ‘in-batch’ is 0.96.

![Graph showing correlation between radioisotope assay and microbiological assay](image)

Fig. 2 Results of red cell folate assay by both methods on 45 samples. The ringed results were from two samples taken from a subject receiving cytotoxic treatment.

Discussion

Microbiological assays for serum folate have been used to identify clinical states due to folate depletion since their advent (Tepley and Elvehjem, 1945). Although a red cell folate level is a more reliable determinant of folate status, being less susceptible to short-term fluctuation than serum folate levels (Chanarin, 1969), serum folate determinations provide a valuable adjunct to clinical diagnosis.

Radioisotopic assays for folate measurement have been explored since folate binders were identified in milk (Ghitis, 1967) and in animal (especially porcine) sera (Mantzos et al., 1974). A variety of methods have been documented (Waxman et al., 1971; Rothenberg et al., 1972; Dunn and Foster, 1973; Mantzos, 1975) recently, all of which suffer from the shortcomings of scintillation counting associated with the use of tritiated folic acid, which appears to prohibit their widespread application for clinical diagnostic purposes (Coulson and Rose, 1974; International Society of Haematology, London, 1975). Further development of a method, using a gamma-emitting radiofolate adhering to basic principles of saturation analysis, has yielded a simple, reliable, and effective method which has a number of advantages compared with the L. casei assay.

(i) It is not susceptible to the caprices of the organism, which occasionally undergoes mutation with alterations in growth patterns. Results are not influenced by contaminating organisms and antibiotics, nor by most drugs which the subject may have been taking at the time of sampling. Large doses of methotrexate, however, may give rise to circulating levels in serum sufficient to raise the apparent folate level artificially.

(ii) The time necessary for an operator to be involved directly with the microbiological assay of a batch of 45 samples is at least 7 to 9 hours spread over three days. Even making a generous allowance for the time of counting the radioactivity, the results from the radioassay can be available the same day the assay was run. Direct technical involvement is necessary for about 2.5 hours, assuming an autogamma counter is available. If a radioassay for vitamin B$_12$ is already installed with available counting apparatus there seems no clear reason to persist with microbiological methods for folate assay, unless the cost of radioassay materials becomes prohibitive.

A recent survey of commercial assay kits was highly critical of the quality of the products and the reliability of the results (Kubasik et al., 1975). The performance, however, of the $^{75}$Se-selenofolate kit under routine conditions at St George’s Hospital illustrates that, for the purposes of serum folate assay, it is reliable, reproducible, and accurate. It is a rapid and simple procedure without the difficulties of $\beta$-counting which have bedevilled some of its antecedents.

Our preliminary findings, measuring red cell folate, suggest that the method lends itself to this purpose, and modifications are being explored to improve the reproducibility and correlation with bioassays (Johnson and Rose, 1977). While it is widely recognised that some drugs affect the microbiological assay (Reizenstein, 1965; Beard and Allen, 1967) we are in the process of determining more comprehensively which commonly administered drugs interfere with the microbiological assay (Rose and Johnson, in preparation).

We thank Mrs J. Tooze for performing the microbiological folate determination on the folate samples submitted during this study.
References


Ian Johnson, Hugh Guilford, and Michael Rose


Measurement of serum folate: experience with 75Se-selenofolate radioassay.
I Johnson, H Guilford and M Rose

J Clin Pathol 1977 30: 645-648
doi: 10.1136/jcp.30.7.645

Updated information and services can be found at:
http://jcp.bmj.com/content/30/7/645

These include:

Email alerting service
Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

Notes

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/