Dysfibrinogenaemia and liver cell growth

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SUMMARY From the evidence presented, it is proposed that ‘dysfibrinogenaemia’ represents the production of normal fetal fibrinogen by rapidly proliferating liver cells in both regenerating and neoplastic tissue. Prolongation of the reptilase clotting time, which was formerly believed to reflect dysfibrinogenaemia, may be rather the result of hepatocytic death.

In the context of these studies, dysfibrinogenaemia (DF) is defined as an excess of immunoassayable fibrinogen, of 1 g per litre or more, over thrombin-clottable protein in the plasma. On this basis, DF was shown to be a reliable biological marker of clinically overt primary hepatocellular carcinoma (PLC) in adult black Africans (Barr et al., 1976). Furthermore, this finding was associated with a prolonged reptilase clotting time (RT) in these patients.

Against this background, a study in Scotland was initiated in an attempt to detect preclinical PLC in adult patients with chronic liver disease, which, for this purpose, was defined as the persistence of any evidence of hepatic disorder for longer than six months. These patients were classified histologically as chronic alcoholic liver disease, chronic active hepatitis, haemochromatosis, or primary biliary cirrhosis.

Material and methods

Plasma samples were prepared as previously described (Barr et al., 1976). Reptilase clotting times were performed by the method of Gralnick et al. (1971) with one modification; each vial of reagent containing 20 Batroxobin units (approximately 50 μg of enzyme) was reconstituted with 1·5 ml rather than 1·0 ml of distilled water.

Thrombin-clottable fibrinogen was assayed by the method of Ogston and Ogston (1966) and heat-precipitable fibrinogen by the method of Millar et al. (1971). In the latter method stored plasma samples were mixed with washed homologous group O red cells to give a haematocrit of approximately 50%.

Immunoassayable fibrinogen was determined by a radial immunodiffusion technique based on that described by Fahey and McKelvey (1965). Antihuman fibrinogen antiserum was incorporated in the agar gel, and multiple aliquots of a human plasma protein solution were applied to appropriate wells for the construction of a standard graph for each plate.

An identical method was used for the estimation of serum alpha-fetoprotein, using serial dilutions of a human alpha-fetoprotein preparation for standardisation. Behring diagnostic reagents for these immunoassays were obtained from Hoechst Pharmaceuticals, Hounslow, Middlesex.

Serum fibrin-fibrinogen degradation products were measured by a standard tanned red cell haemagglutination-inhibition technique (Wellcome FDP kit).

All estimates were performed in duplicate and the mean result was recorded in each instance.

Results

Control data were obtained from 20 healthy colleagues in Aberdeen (Table 1). We then examined samples from eight Scottish patients with PLC. Of these, seven revealed a prolonged RT, but DF was detectable in none.

<table>
<thead>
<tr>
<th>Table 1 Control data (median ± 2 SD)</th>
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<tr>
<td>Reptilase clotting time(s)</td>
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<tr>
<td>Normal range</td>
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<tr>
<td>Thrombin-clottable fibrinogen (g/l)</td>
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<tr>
<td>Normal range</td>
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<tr>
<td>Heat-precipitable fibrinogen (ml/100 ml)</td>
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<tr>
<td>Normal range</td>
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<tr>
<td>Immunoassayable fibrinogen (g/l)</td>
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<td>Normal range</td>
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Results from patients with chronic liver disease were grouped according to four patterns (Table 2).
Table 2  Patients with chronic liver disease

<table>
<thead>
<tr>
<th>Group</th>
<th>A</th>
<th>B</th>
<th>C</th>
<th>D</th>
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<tbody>
<tr>
<td>Samples</td>
<td>22</td>
<td>18</td>
<td>14</td>
<td>21</td>
</tr>
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A = dysfibrinogenaemia and prolonged reptilase time;  
B = prolonged reptilase time alone;  
C = dysfibrinogenaemia alone;  
D = normal reptilase time and no dysfibrinogenaemia.

The combined abnormality (DF plus prolonged RT) was demonstrable in as many samples as those on which the results were normal; and almost as many showed DF alone as prolonged RT alone. Hence DF and prolonged RT do not appear to reflect the same phenomenon, and both occurred too frequently to be indicative of the presence of PLC. In no patient could the prolonged RT be explained by hypo-  
dysfibrinogenaemia or raised levels of fibrin-fibrinogen  
degradation products.

In a further group of 10 patients with carcinoma metastatic to the liver, only one had DF and a prolonged RT demonstrable. This exceptional patient was an alcoholic. These findings further support the specific association of acquired DF with primary hepatic disease, which is predictable since fibrinogen is produced selectively in the liver (Straub, 1963).

Discussion

Abnormal fibrin monomer polymerisation has been described in 50% of a large series of patients with chronic parenchymal liver disease (Green et al., 1976). This abnormality does not seem to be due to the presence of inhibitory activity (Green et al., 1976), factor XIII deficiency (Lane et al., 1977), or a major abnormality of the molecular structure of fibrin monomers (Lane et al., 1977). The incidence of the defect is essentially identical with that obtained for DF in chronic liver disease in the present study (Table 1). Moreover, DF (as defined by us) and defective fibrin monomer polymerisation have been described in the same PLC patient (Verhaeghe et al., 1972). This suggests that DF and abnormal fibrin monomer polymerisation in liver disease represent the same basic defect.

Gralnick et al. (1975) have reported ‘defective’ fibrin monomer polymerisation with normal human fetal fibrinogen. Since this finding is also the commonest abnormality in congenital dysfibrinogenaemia (Gralnick and Finlayson, 1972), it suggests that this latter entity may rather be due to a persistence of fetal fibrinogen production.

Fetal proteins may be re-expressed postnatally in human tissues, not only in association with malignancy (Fishman, 1976) but also with simple regeneration (Jerry et al., 1976). Thus a marked rise in fetal haemoglobin is not only typical of juvenile chronic myeloid leukaemia (Maurer et al., 1972) but has also been described in association with the early phase of successful bone marrow transplantation (Alter et al., 1976). Indeed, in the case of alpha-fetoprotein, serum levels may be as high in neonatal hepatitis as are ever recorded in PLC (Kang et al., 1972; Zeltzer et al., 1974).

On the basis of these interrelationships, it can be postulated that DF in liver disease results from derepression, in hepatocytes, of fetal genes coding for fibrinogen production; and, further, that this occurs only in proliferating liver tissue, whether malignant or benign, although it may require that the constituent cells have a short mean generation time and high growth fraction. In this regard it is of considerable interest that, in an experimental hepatoma model, the serum alpha-fetoprotein concentration was found to relate directly to the growth rate of the tumour (Sell et al., 1974), and it has been proposed that the detection of raised levels of serum alpha-fetoprotein, in human non-neoplastic hepatic disease, may result from liver cell regeneration (Zeltzer et al., 1974; Bloomer et al., 1975).

Thus may be explained our observation that DF is not a feature of clinically overt PLC in Scotland but is associated with the tumour in Africans, in whom the disease is more rapidly progressive (Perves, 1976). Furthermore, in benign intrinsic liver disease, DF regresses with clinical resolution, as has been our experience and that of others (Soria et al., 1970; Aiach et al., 1973; Green et al., 1976). By contrast, DF is not a feature of ‘secondary’ liver disease, for example metastatic carcinoma. In this circumstance, employing accepted criteria (Popper, 1954), convincing histological evidence of liver cell regeneration, adjacent to the metastases, was not detected in a survey of postmortem material from 15 patients who had secondary carcinoma in the liver. Indeed, the dominant features in those hepatocytes adjacent to the expansile tumour deposit appeared to be degenerative in nature.

Prolongation of the RT is more difficult to explain. We suggest that this represents a separate though possibly related abnormality, and we postulate that it reflects the release, from dead and dying hepatocytes, of material which does not exhibit fibrin-fibrinogen related antigen but which inhibits the action of reptilase on fibrinogen. Support for this hypothesis is provided by the finding, in the present study, that the concentration of serum aspartate aminotransferase was significantly higher in chronic liver disease patients with prolonged RT than in patients in whom the RT was normal ($p < 0.01$,  

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Wilcoxon’s one-tailed test). By contrast there was no demonstrable association between DF and serum aspartate aminotransferase concentration.

Thus a prolonged RT occurs in the majority of patients with chronic parenchymal liver disease (Green et al., 1976), in many patients with secondary hepatic damage who do not have DF (Green et al. 1976), and in virtually all patients with PLC irrespective of the presence of DF. Again, resolution of benign liver disease has been accompanied by restoration of the RT to normal (Soria et al., 1970; Green et al., 1976) in accordance with our own findings.

Although neither DF nor a prolonged RT is indicative of PLC, a pattern of progressive abnormality on serial sampling may point to this diagnosis. We have studied one such patient in whom a histological diagnosis of micronodular cirrhosis was made in 1975. In June 1976 he presented with a considerable increment in hepatomegaly. $^{99m}$Tc sulphur colloid and $^{75}$Se selenomethionine scanning and repeated ultrasonography suggested the presence of PLC, and a probable tumour circulation was demonstrable by hepatic arteriography. Unfortunately this could not be confirmed by percutaneous and laparoscopic biopsies. Results of repeated estimations on this patient are shown in the Figure.

![Graph showing the relationship between Plasmin fibrinogen (grams per litre) and Replicate clotting time in seconds.](image)

**Figure** Results of investigations in one patient, June-November 1976: **RCT** = reptilase clotting time; **IAF** = immunoassayable fibrinogen; **TCF** = thrombin clottable fibrinogen; **HPF** = heat-precipitable fibrinogen.

Alpha-fetoprotein has never been demonstrable in his serum by immunodiffusion nor by immuno-electrophoresis. Indeed, no patient in the present study had a serum alpha-fetoprotein level detectable by the former technique, which has a sensitivity of 3000 ng per ml (Bloomer et al., 1975).

We suggest that all patients with chronic parenchymal liver disease, but particularly males with haemochromatosis and alcohol-induced organ damage, be subjected to routine serial screening for DF and prolonged RT. In the event that progressive abnormalities are identified, every effort should be made to define and localise the PLC with a view to early and curative surgical removal of the tumour in suitable patients.

Our ongoing studies are directed to this end and to the further evaluation of these techniques, in patients with acute hepatitis, as potential predictors of clinical disease evolution.

We acknowledge the generous gift of Reptilase-R from Paines and Byrne Ltd, and the co-operation of our colleagues in allowing us to study their patients. The results of these investigations were presented in part to the American Association for Cancer Research, Denver, Colorado, May 1977; and to the Pathological Society of Great Britain and Ireland, Aberdeen, July 1977.

References


