Plasma blood group changes in gastrointestinal tract carcinoma

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SUMMARY The levels of A, H, I, and i plasma antigens and of anti-B, anti-I, and anti-T antibodies were measured in 70 subjects with colonic or gastric carcinoma. These studies showed a significant increase in A plasma activity of the A subjects, and in H plasma activity of the O subjects, while 25% of the tested subjects showed increased I plasma activity. There was no difference in i plasma activity between cancer patients and healthy subjects. These results take into account the marked polymorphism acquired by neoplastic tissue, which is capable also of producing a greater quantity of antigens than that of healthy subjects. Nevertheless this heterogeneity forms a barrier to the clinical measurement of these plasma antigens for screening neoplasms. The significant fall in the amount of anti-T antibodies seemed to be secondary to the absorption of these antibodies on the surface of the tumour cells.

Many authors (Salmon and Malassenet, 1960; Saeed and Fine, 1968) have reported an excess of A, B, and H substances in the plasma of patients with gastrointestinal tract neoplasms. The A, H, Leα, and T glycosyltransferases were proved to be present in colonic tumour cells on culture (J. P. Cartron, 1978, personal communication).

On the other hand, Davidsohn et al. (1971) and Feizi (1978) demonstrated the disappearance of A, B, and H substances in the tumour cells of the gastrointestinal tract, whereas the I substance seemed to be present in an abnormally high quantity (Feizi, 1978). (I antigen could be a precursor of the biosynthesis of H antigen.) Stellner et al. (1973) showed in adenocarcinoma cells, the disappearance of enzyme activities capable of transforming H₁ glycolipids into A or B glycolipids.

The present study was undertaken to find out if the modifications seen in tumour tissue from the gastrointestinal tract had any effect on the level of A, H, I, and i substances and anti-B, anti-I, and anti-T antibodies in the serum in order to investigate the potential value of their use in cancer screening.

Material and methods

The titration of A, H, I, and i plasma antigens is based on the inhibition of haemagglutination. The basic agglutination must be as weak as possible, but at the same time detectable in the used system. The agglutination obtained with a low-antibody concentration can thus be inhibited by a low-substance concentration.

Increasing dilutions of plasma were mixed for 2 hours at 20°C with a constant antibody concentration (inhibition phase). After fixation the amount of free antibodies was measured by their ability to agglutinate specific red cells. The agglutination phase of non-treated red cells was performed in a manifold combined with an autoanalyser (Rouger et al., unpublished observation (Fig.1).

Plasma dilutions mixed with antibodies were pumped and flowed in a reaction coil. A red blood cell solution and PVP were added. This mixture reacted for 30 minutes. The reaction coil was at 10°C or 15°C, according to the system used (Table 1). After double decantation, the residual red cell concentration was determined in a continuous flow counter, and the results were recorded with a pen recorder.

From the results obtained, we calculated the agglutination percentages for the chosen dilutions. The agglutination percentage curve, expressed in terms of logarithms of tested plasma dilutions, was a sigmoid, which was linearised by probit-logarithm transformation. Each tested plasma was referred to a standard inhibitory solution. The reference solution contained 100 arbitrary inhibition units.

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Fig. 1 A schematic flow diagram of the circuit used for the titration of A, H, I, and i antigens.

(AIU). The comparison obtained between the tested plasma and the reference solution allowed the determination of the number of AIUs present in the tested plasma. The reproducibility of these tests was confirmed by repeated measurements of the same plasma at the same dilutions. The variation observed did not exceed 20%.

The concentrations of anti-B and anti-I antibodies were measured in an AutoAnalyzer using bromelin-treated red cells (B cells at 15°C for anti-B activity and O red cells at 4°C for anti-I activity). The concentration of antibodies was measured in the same way with the help of neuraminidase-treated O:i red cells at 20°C.

The secretor status of each subject was determined by the study of A, B, or H substances in the saliva. Thus 70 subjects with gastric or colonic cancers without metastasis were tested. The diagnosis was confirmed by histological examination after surgery. Each pathological population was compared with a healthy population (180 subjects). Both the healthy and cancer populations were identical as far as age and sex were concerned (patients were aged between 25 and 65; 50% were men and 50% were women).

Table 1 Some technical aspects of A, H, I, and i antigen titration

<table>
<thead>
<tr>
<th>Plasma activities</th>
<th>A</th>
<th>H</th>
<th>I</th>
<th>i</th>
</tr>
</thead>
<tbody>
<tr>
<td>Temperature (°C)</td>
<td>20</td>
<td>20</td>
<td>20</td>
<td>20</td>
</tr>
<tr>
<td>Inhibition</td>
<td>15</td>
<td>10</td>
<td>15</td>
<td>15</td>
</tr>
<tr>
<td>% PVP</td>
<td>1-5</td>
<td>1</td>
<td>1-5</td>
<td>1</td>
</tr>
<tr>
<td>Antibodies</td>
<td>Anti-A (pool)</td>
<td>Anti-H (CER) (1)</td>
<td>Anti-I (ABG) (2)</td>
<td>Anti-I (PEA)</td>
</tr>
<tr>
<td>Dilution</td>
<td>1/24</td>
<td>1/21</td>
<td>1/10500</td>
<td>1/100</td>
</tr>
<tr>
<td>Red blood cells</td>
<td>A</td>
<td>O</td>
<td>O</td>
<td>O</td>
</tr>
<tr>
<td>Reference inhibitory activities</td>
<td>A substance from pig and horse stomach</td>
<td>O saliva pool O and A</td>
<td>O sera pool</td>
<td>O and A sera pool</td>
</tr>
<tr>
<td>Remarks</td>
<td>Treatment of plasma by 2 mercapto-ethanol 0.2M</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

(1; B subject; 2 Auto-immune haemolytic anaemia)
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Table 2  

<table>
<thead>
<tr>
<th></th>
<th>Healthy subjects</th>
<th>Subjects with GIT carcinoma</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>n</td>
<td>Mean value</td>
</tr>
<tr>
<td>Secretor</td>
<td>31</td>
<td>168</td>
</tr>
<tr>
<td>Non-secretor</td>
<td>9</td>
<td>76</td>
</tr>
<tr>
<td>Total</td>
<td>40</td>
<td>111.5</td>
</tr>
</tbody>
</table>

n = number of subjects tested

Results

A PLASMA ANTIGEN

A plasma activity in group A (A₁ and A₂) subjects with cancer of the gastrointestinal tract appeared significantly higher than that in healthy subjects of the same group (Fig. 1) (p < 0.001).

The average values were 70 and 140 AIU for the healthy and cancer subjects respectively.

H PLASMA ANTIGEN

H plasma activity in group O subjects with gastrointestinal tract neoplasms was significantly higher than that in healthy group O subjects (p < 0.01). On the other hand, H plasma activity in A₁ subjects was identical in the two populations. In the healthy subjects, A and H activities were dependent on Lewis and secretor phenotypes, while the abnormal A and H activities in cancer subjects were independent of these systems (Table 2).

Fig. 2  A plasma activity is higher in patients with a gastrointestinal tract carcinoma than in the normal population (— normal subjects (64); —— subjects with G I T carcinoma (22)).

Fig. 3  Comparative study of I plasma antigen in normal subjects (— 39) and subjects with G I T carcinoma (—— 38).

Fig. 4  Comparative study of I plasma antigen in normal subjects (— 33) and subjects with G I T carcinoma (—— 34).
have described an excess of ABH substances in the plasma of cancer subjects (epithelium, ovary, stomach, pancreas). In these exceptional observations, the amount of A or H substance was particularly high, almost the same as the quantity of A or H substance in the saliva. The increases of A and H plasma activity shown in this study are much less than those described in these rare cases. The substances and blood group enzymes have been shown in tumour cells on culture. In the HT 29 cellular line from a cancer of the colon, Cartron (1978, personal communication) demonstrated A, H, Le^a, and T glycosyltransferase activities. In the BOT 2 line (Anglin et al., 1977), originating from a breast cancer, glycoproteins with blood group activity were present in the culture medium. In the HeLA line, cells in culture possess H specificity (carried by the glycoproteins, notably CEA) and can be converted in vitro in A or B specificity (Pann and Kuhns, 1972; Mousseron-Canet et al., 1975; Bali et al., 1976). These results take into account the persisting activity of the synthesis of blood group substances of the tumour cells on the one hand and the marked polymorphism of the different cells on the other.

Elsewhere, many authors (Nairn et al., 1962; Davidson et al., 1969; Davidson and Ni, 1969; Davidson et al., 1971; Lill et al., 1976; Lin et al., 1977) have shown that there is more often a weakened A, B, or H activity in gastric adenocarcinoma and a high I activity; but, in each tumour studied possessed a particular profile with regard to ABH activities.

There is no correlation between the levels of A, H, and I in the plasma of a cancer subject. These results obtained in plasma ‘far from the tumour’ take into account the polymorphism already described for tumour tissues and tumour cells on culture, as far as the haemagglutination inhibition has allowed the ‘inhibitor activities’ to be shown, without regard to their biochemical support.

This heterogeneity forms a barrier to the dosage of A, H, I, and i plasma activities for screening gastrointestinal tract neoplasms, apart from a few exceptional cases.

As far as anti-T is concerned, we found almost the same data as Springer et al. (1977), but this is probably not useful in the detection of carcinoma. The normal anti-B and anti-T antibodies might result predominantly from continuous antigenic stimulation by intestinal flora; the difference between anti-T and anti-B shows that the immunological function is normal and that the anti-T decrease could be related to absorption on tumour cells, on the membranes of which the T antigen is well developed. Springer et al. (1977) have shown the same phenomenon in breast cancers and have tried
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to stimulate the production of anti-T by inducing active immunotherapy.

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References


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