Technical methods

A simple method for recording thin-layer acrylamide gel electropherograms

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Thin-layer acrylamide gel electrophoretic technique, based on the method described by Nandi and Lewis, has been used in our laboratory for the electrophoresis of blood and malaria parasite enzymes. It was found that preservation of the enzyme electropherograms in acid solution is cumbersome, space-occupying, and unstable. Alternative methods for permanent recording of the electropherograms are direct scanning of the gels in a densitometer and photographic techniques. These techniques require extra equipment and skill. In this paper we describe a simple procedure for recording electropherograms of enzyme patterns in their original size and colour.

Method

Electrophoresis is carried out by using parasite enzyme extracts or haemolysates on 5·5 g% acrylamide gel plates (2 × 120 × 165 mm). After electrophoresis, the gel is carefully detached from the glass plate under ice-cold water using a blunt, thin spatula. The gel is stained with an appropriate MTT-linked enzyme substrate staining solution. The stained gel is washed with phosphate-buffered saline (pH 7·4; 0·15 M) to stop the enzymic action and staining. The washed gel slab is then placed on a Perspex sheet or paper-thin plastic sheet which has been placed on the base of a metallic tray. Excess buffer is gently drained off by tilting the base plate. The gel is covered with a strip of Whatman filter paper No. 1, which is slightly larger than the size of the gel. The filter paper is well flattened out on the surface of the gel by the middle finger, removing air bubbles at the same time. The gel sandwich thus prepared is dried at room temperature or at 37°C overnight. The dried gel is well adhered to the filter paper, forming a glossy-type of coloured electropherogram which has been filed and stored more than six months without deterioration. The electropherogram retains the original size and colour of the enzyme patterns (Figure).

We express our thanks to Ko Sein Min for his able technical assistance.

References


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Received for publication 12 February 1980
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doi: 10.1136/jcp.33.7.699