Distribution of ferritin, transferrin and lactoferrin in breast carcinoma tissue

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SUMMARY An immunoperoxidase staining technique was used for detecting three major iron binding proteins (ferritin, transferrin and lactoferrin) in 40 breast carcinoma cases and six benign breast proliferative lesions.

Ferritin staining was detected mainly in connectival stroma and in histiocytes surrounding neoplastic cells. Few and faint ferritin positivities were also detected in neoplastic cells of 20 carcinoma cases. Transferrin was found inconsistently in myoepithelial cells surrounding normal ducts, or around neoplastic ducts of ductal in situ carcinoma. In eight carcinoma cases, transferrin staining was also positive in neoplastic cells. Lactoferrin was detected only in normal breast epithelial cells and in benign breast proliferative lesions.

These immunohistochemical findings may suggest that raised serum ferritin concentrations in breast carcinoma patients might be attributed to stromal reaction rather than to tumor synthesis. Transferrin staining of neoplastic cells in these carcinoma cases appears to be very intriguing, particularly since transferrin is considered an obligate requirement for growing cells, and transferrin receptors have been demonstrated only in dividing cells. On the basis of the immunohistochemical data, lactoferrin might be used as a pointer to benign lesions.

Ferritin, lactoferrin and transferrin are the three major iron-binding proteins in man. Until recently the clinical interest for these proteins was mainly confined to their role in iron transport and/or storage.

Nevertheless, increased serum ferritin concentrations have been found in a wide variety of malignant states including Hodgkin’s disease, leukaemia, carcinoma of breast, pancreas, stomach and colon-rectum.6 8 10 12 19

Metabolic disorders in mononuclear phagocytic cells, lymphocytes or macrophages synthesis, tissue damage and tumor synthesis have been suggested as an explanation for this increase in ferritin serum concentrations in malignancy.2 6 8-10 12 15 17 21 22

On the other hand, raised transferrin serum concentrations and transferrin receptors have been demonstrated in breast carcinoma.3 5 11 16

Lactoferrin, like transferrin, is considered to play an important role in iron transport in human breast and other glandular tissues.13 14 20

In a previous study,13 transferrin was found, by an immunoperoxidase method, mainly in periductular myoepithelial mammary cells, whereas ferritin was detected in periacinar, and lactoferrin in acinar cells.

In view of an increased interest in oncology for these iron-binding proteins, we have investigated ferritin, transferrin and lactoferrin distribution in breast carcinoma by an immunoperoxidase method.

Material and methods

Forty cases of breast carcinoma (31 invasive ductal, three in situ ductal, three lobular invasive and three mucoid carcinoma cases) and six benign neoplastic lesions (three fibroadenoma and three fibrocystic disease cases) were included in this study.

Tissue specimens were frozen in isopentane, cooled with liquid nitrogen or fixed in Bouin’s and in 10% buffered formalin, and then processed routinely. Both procedures gave similar results, although the staining generally appeared more intensive with Bouin’s fixative.

Ferritin, lactoferrin and transferrin (Dako antisera) identification was carried out by immunoperoxidase, PAP method, according to Taylor18 with
Fig. 1  Ferritin in invasive ductal carcinoma. Prevalent distribution of ferritin staining in connectival stroma PAP–Haematoxylin × 100.

Fig. 2  Ferritin in invasive ductal carcinoma. Few histiocytes surrounding neoplastic cells show an intensive ferritin staining PAP–Haematoxylin × 250.

Fig. 3  Ferritin in invasive ductal carcinoma. Faint ferritin staining in some neoplastic cells. PAP–Haematoxylin × 250.

Fig. 4  Transferrin in intraductal carcinoma. Spindle-shaped cells, around neoplastic ducts or in connectival stroma display intensive staining. PAP–Haematoxylin × 250.

Fig. 5  Transferrin in a “clinging” carcinoma. Transferrin positive cells are mainly arranged at the periphery of neoplastic lesion. PAP–Haematoxylin × 250.
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Fig. 6 Transferrin in invasive ductal carcinoma. Several neoplastic cells show a variable transferrin staining PAP-Haematoxylin × 100.

Fig. 7 Lactoferrin in a lobular unit. Secretory lactoferrin positive material may be observed within the wide ductal lumen. PAP-Haematoxylin × 100.

Fig. 8 Lactoferrin in adenosis area. Focal distribution of lactoferrin staining. PAP-Haematoxylin × 250.

minimal modifications as previously reported.

The sections were sequentially incubated for 20–30 min at room temperature with each of following reagents: (a) methanolic hydrogen peroxide; (b) normal swine serum (Dako); (c) rabbit antihuman ferritin, transferrin or lactoferrin antiserum; (d) swine antirabbit IgG serum (Dako); (e) PAP (Dako).

After each incubation, the slides were washed with Tris-saline and placed in Tris-buffer for 20 min. The sites of antibody binding are shown by developing the peroxidase reaction with diaminobenzidine according to Weir.

The slides were then washed with water, counterstained with haematoxylin, dehydrated and mounted in Entellan.

The specificity of the immunohistochemical method was confirmed by the negative results obtained after omission and replacement of the antisera, and by neutralisation studies using antisera adsorbed with human ferritin and lactoferrin (Calbiochem-Behring).

Results

The results are summarised in the Table. In areas of normal and dysplastic tissue as well as in fibroadenomas, no significant immunoperoxidase staining was observed in epithelial cells. Only few macrophages in connectival stroma showed faint cytoplasmic staining. In 20 cases of carcinoma (50%) a weak ferritin staining in few neoplastic cells was usually observed.

In connectival stroma, particularly in areas sur-
**Distribution of ferritin, transferrin and lactoferrin in benign and malignant breast lesions.**

<table>
<thead>
<tr>
<th>Lesions</th>
<th>Ferritin</th>
<th>Transferrin</th>
<th>Lactoferrin</th>
</tr>
</thead>
<tbody>
<tr>
<td>Breast normal tissue</td>
<td>E −</td>
<td>−</td>
<td>+++</td>
</tr>
<tr>
<td></td>
<td>S +</td>
<td>++*</td>
<td>−</td>
</tr>
<tr>
<td>Fibroadenoma</td>
<td>E −</td>
<td>−</td>
<td>−</td>
</tr>
<tr>
<td></td>
<td>S +</td>
<td>++*</td>
<td>−</td>
</tr>
<tr>
<td>Mammary dysplasia</td>
<td>E −</td>
<td>−</td>
<td>++</td>
</tr>
<tr>
<td></td>
<td>S +</td>
<td>++*</td>
<td>−</td>
</tr>
<tr>
<td>Carcinoma</td>
<td>E +(50%)</td>
<td>+++ (20%)</td>
<td>−</td>
</tr>
<tr>
<td></td>
<td>S +++</td>
<td>+</td>
<td>−†</td>
</tr>
</tbody>
</table>

E = epithelial component, S = stromal component, *Myoepithelial cells and histiocytes, †Only lactoferrin positive-granulocytes. +++ = strongly positive, ++ = moderately positive, + = weakly positive.

It has been demonstrated that ferritin staining was mostly observed in connectival stroma and macrophagic cells surrounding neoplastic tissue. Ferritin staining was seldom observed in neoplastic cells and only in half of carcinoma cases. Other workers, on the contrary, have suggested that hyperferritinemia might be due to tumour synthesis. Furthermore, few immunohistochemical data have been reported in the literature: ferritin has been demonstrated in embryonal carcinoma cells by immunofluorescence and in carcinoma-in-situ of the testes by immunoperoxidase.

It has long been known that ferritins are a family of isomeric proteins containing two subunits, termed H and L, in different proportions. H subunit is found predominantly in the so-called acidic isoferritins, which are detectable in the heart, tumours, and HeLa cells. Basic isoferritins, on the other hand, contain prevalently L subunits and are usually detectable in the liver and spleen tissues. The antiserum employed by us (Dako) has been raised in rabbit with human liver and spleen ferritins as immunising antigens.

Consequently, this antiserum shows a lesser reactivity for the more acidic tumour isoferritins. The faint ferritin staining in neoplastic cells, as detected in the present study, might be attributed to this lower reactivity.

An unexplained, prevailing transferrin localisation in myoepithelial cells has been reported. In the present study, this peculiar localisation appeared to be inconstant and non-specific, since transferrin staining was also detected in stromal histiocytes. In addition, the presence of transferrin positivities in neoplastic cells of eight carcinoma is very intriguing. Transferrin has been identified as an obligate growing factor in serum-free conditions. Transferrin receptors have also been considered to be a potential marker for identifying cells undergoing divisional activity and requiring the incorporation of additional iron. Therefore, it is tempting to speculate that transferrin staining may occur only in carcinoma and in neoplastic cells with high mitotic activity. Further studies are required to clarify these possible relations.

In the case of lactoferrin, our findings in normal breast epithelium appear to be in agreement with several papers which have provided immunohistochemical evidence for the presence of this protein in glandular tissue of different organs. The lack of lactoferrin staining in all of 40 breast carcinoma suggests that this protein might be considered as a
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potential marker of benign lesions.

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References


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