Enzyme linked immunosorbent assay for detecting adenoviruses in stool specimens: comparison with electron microscopy and isolation

A L Martin, G Kudesia

Abstract
A commercial enzyme linked immunosorbent assay (ELISA) for detection of adenoviruses in stool samples was compared with the use of electron microscopy and isolation in Graham 293 cells. Although specific, the ELISA was less sensitive than both electron microscopy and isolation. The ELISA had an overall sensitivity of 78% and a specificity of 100%. The sensitivity was related to the amount of virus particles present in the stool sample, increasing to 90% with about 10⁵ viral particles/ml of stool. The ELISA was easy to perform, requiring no instrumentation, and is a useful first line test for detection of adenoviruses in stool samples, especially in laboratories without access to an electron microscope.

Wider use of ELISAs should help in evaluating the role of adenoviruses in viral gastroenteritis.

Adenovirus are often isolated from the stools of children with or without gastroenteritis. Although several outbreaks of gastroenteritis due to adenoviruses have been described, their role in sporadic cases of gastroenteritis remains uncertain. Serotypes 40 and 41 have recently been strongly linked with gastroenteritis, although some workers suggest that other serotypes may also cause gastroenteritis. The diagnosis of adenoviral gastroenteritis is limited to electron microscopical examination as serotypes 40 and 41 are not easily grown in cell culture, hence the name fastidious adenoviruses. We have evaluated a group specific commercial adenovirus ELISA NovoBiolabs (formerly Boots Celltech) and compared it with electron microscopy and isolation in tissue culture.

Methods
One hundred and thirteen stool samples were examined by the new aden ELISA method. Forty five stools were from 37 children, all of whom had diarrhoea, there were two or more serial specimens from six patients, and all the stools were positive for adenovirus by electron microscopical examination. Sixty eight stools were included as controls, of which 18 were negative controls taken from children without diarrhoea, 50 stools were from children with diarrhoea, 40 stools were negative by electron microscopical examination, and 10 had other viral pathogens. To test for cross reaction with staphylococcal protein A in the enzyme immunoassay a 10⁻¹ to 10⁻⁴ dilution of lyophilised Cowan strain Staphylococcus aureus (Sigma) was made in a negative stool sample and 100 μl of each dilution treated as sample.

Electron Microscopy
Electron microscopical examination was performed after 10% PBS extraction of stool; this extract was then concentrated by ultracentrifugation at 200000 x g for 60 minutes. The pellet was suspended in 1% Bactracin and negatively stained with phosphotungstic acid at pH7 and then examined at a magnification of 57000 in a Philips 301 electron microscope. The quantity of virus present was scored on a scale of occasional to 4+.

ELISA
Boots Celltech IDEIA adenovirus enzyme immunoassay was performed according to the manufacturer’s instructions. This is a solid phase sandwich type assay. Breakaway microwells are coated with a murine monoclonal antibody against the group reactive hexon antigen of adenovirus. Diluted faecal sample (100 μl) was then added to each well with 100 μl of enzyme conjugate. After 60 minutes of incubation at room temperature the sample well was washed with deionised water to remove unbound specimen and excess enzyme labelled antibodies. Enzyme substrate (urea peroxyde) and chromogen (tetramethyl benzidine) were then added to the wells and incubated for 10 minutes at room temperature: any bound enzyme conjugate in the wells converts the colourless substrate to a blue colour, signifying a positive reaction. The plates were read at 450 nm after stopping the reaction with 1N sulphuric acid after 10 minutes. Positive and negative controls were included in each run.

Cell Culture
All stool extracts were cultured in Graham 293 cell line, which supports the growth of faecal adenoviruses. The cells were grown in minimal essential medium (MEM) with 10%, fetal calf serum (FCS), and maintained in MEM with 1%, FCS. The inoculum was adsorbed at 37°C for one hour, after which the maintenance medium was added. The tubes were examined daily. The presence of adenovirus was confirmed by the appearance of typical cytopathic effects, electron microscopical examination of the tissue culture fluid, or by ELISA.
Use of ELISA to detect adenoviruses in stool specimens

Table 1 Correlation of adenovirus EIA with electron microscopy

<table>
<thead>
<tr>
<th>EM score</th>
<th>No of stools</th>
<th>EIA+</th>
<th>EIA−</th>
<th>Sensitivity (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>4+</td>
<td>2</td>
<td>2</td>
<td>0</td>
<td>100</td>
</tr>
<tr>
<td>3+</td>
<td>6</td>
<td>6</td>
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<td>91</td>
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<td>1+</td>
<td>9</td>
<td>8</td>
<td>1</td>
<td>89</td>
</tr>
<tr>
<td>Occ</td>
<td>17</td>
<td>9+</td>
<td>8</td>
<td>53</td>
</tr>
</tbody>
</table>

*Six weak positive
†Occasional = about 10⁶ particles/ml
1+ = about 10⁷ particles/ml
2+ = about 10⁸ particles/ml
3+ = about 10⁹ particles/ml
4+ = about 10¹⁰ particles/ml

Discussion

The adenovirus serotypes 40 and 41 have recently been associated with viral gastroenteritis. As these viruses do not grow in cell culture used routinely in most diagnostic laboratories infection with them is going to be missed unless electron microscopy is available. An ELISA has the advantage of not requiring expensive instrumentation and being rapid and easy to perform. In our hands the new IDEIA had a specificity of 100%. Although the manufacturers state that the presence of protein A may give false positive reactions, we found no experimental evidence of this after artificially adding S aureus to a negative stool sample. Furthermore, staphylococcal enterocolitis in children is extremely rare, and therefore the likelihood of staphylococcal protein A being present in a clinical sample is very low.

The overall sensitivity of 78% was disappointing, but it increased with the number of viral particles/ml of stool, reaching 100% with about 10⁶−10⁷ viral particles/ml of stool sample. In our group of patients there was a direct relation between the number of virus particles being shed in the stool and the clinical importance—that is, the severity of illness and association with other enteric pathogens. It may be argued, therefore, that the ELISA will be useful in detecting clinically important infections.

Type specific ELISA (serotypes 40 and 41) are not widely available in this country but have been valuable in assessing the role of these viruses in gastroenteritis. Adenoviruses have been reported in 13% and 8% of all stool samples from children with diarrhoea from Sweden⁵ and Glasgow⁶ respectively, and may make up to 22% of all stools positive by electron microscopy (G Kudesia, unpublished observations). There is therefore a need for a screening test for adenovirus gastroenteritis, especially in laboratories without access to an electron microscope.

The group specific EIA would be an appropriate screening test, but a confirmation step for 40 and 41 by neutralisation would be useful as that would enable the role of the higher adenovirus serotypes to strengthen in viral gastroenteritis, and at the same time the role of other adenoviruses in viral gastroenteritis could be evaluated further.

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