Splenic lymphoma with circulating villous lymphocytes

F Imbing Jr, D Kumar, S Kumar, G Yuoh, F Gardner

Abstract
This report describes the occurrence of splenic lymphoma with villous lymphocytes (SLVL) in a 56 year old white female with a family history of chronic lymphocytic leukaemia. Other unusual features included a marked lymphocytosis with counts up to 224 x 10^9/l and marked clumping of lymphocytes in EDTA anticoagulated blood. The neoplastic cells were CD10⁺, CD20⁺, CD22⁺, CD25⁻, IgM +, λ +, κ −, CD5⁻ and CD10 −. The spleen had nodular infiltrates of B lymphocytes in the region of the white pulp with minimal red pulp involvement. Electron microscopy of peripheral blood lymphocytes revealed cells with polar cytoplasmic processes. This report underlines the need for detailed analysis, including morphology and immunophenotyping, for each patient with a small B cell lymphoproliferative disorder (J Clin Pathol 1995;48:584–587).

Keywords: Splenic lymphoma with villous lymphocytes, immunophenotype, familial, electron microscopy.

The disease now referred to as splenic lymphoma with villous lymphocytes (SLVL)¹ was previously described by Neiman et al² under the rubric malignant lymphoma simulating leukaemic reticuloendotheliosis. In their study of 10 cases, Neiman et al compared and contrasted this disorder with hairy cell leukaemia (HCL) and prolymphocytic leukaemia (PLL) and emphasised the importance of a histological examination of the spleen in making the distinction. The patient described here was initially thought to have B chronic lymphocytic leukaemia (CLL) based on her family history. A diagnosis of SLVL was later made by correlating the morphology and immunophenotype of peripheral blood lymphocytes and histology of the resected spleen.

Case report
A 56 year old white female presented with progressively increasing abdominal girth in November 1993. On physical examination, the spleen was massively enlarged, being palpable 13 cm below the left costal margin. There was no peripheral lymphadenopathy. Abdominal computed tomography scans confirmed massive splenomegaly with hypodense areas suggestive of leukaemic or lymphomatous infiltrate, or both. No mediastinal or abdominal lymphadenopathy was observed.

Examination of the peripheral blood revealed a haemoglobin concentration of 104 g/l; a platelet count of 121 x 10^11/l; a white blood cell count (WBC) of 69 x 10^9/l, with 98% lymphocytes. The lymphocytes were larger than small lymphocytes and had a moderate amount of cytoplasm. The nuclei were round, with few clefted and irregular forms and had a clumped chromatin pattern (fig 1). Small but distinct nucleoli were present in some of the cells. Irregularly distributed surface projections, which were sometimes concentrated at one or both poles of the cell, were seen in some cells. An unusual feature noticed in the smears was marked clumping of the lymphocytes (fig 1). The leukaemic cells were tartrate resistant acid phosphatase (TRAP) negative. Electron microscopic studies were carried out on buffy coated preparations of peripheral blood. Small villous
Splenic lymphoma with circulating villous lymphocytes

Figure 1  The panel on the left shows clumping of lymphoma cells in the peripheral blood smear (× 250). At high magnification (middle panel; × 1000), the neoplastic cells have a moderate amount of cytoplasm and a round to irregular nucleus with clumped chromatin. The electron micrograph shows the polar distribution of the villous processes (right panel; × 9000).

Figure 2  A portion of the spleen showing nodular involvement of the white pulp (left panel). At low magnification, exclusive involvement of the white pulp with effacement of its architecture can be seen (middle panel; × 100). At high magnification, the lymphoma cells have round to slightly irregular nuclei, moderately clumped chromatin and inconspicuous nucleoli (right panel; × 400).

cytoplasmic projections were seen arranged at one or both poles of many of the neoplastic cells (fig 1).

Flow cytometric immunophenotyping of gated lymphocytes gave the following results: CD45+, 98%; HLA-DR+, 95%; CD10+, 91%; CD20+, 94%; CD22+, 93%; CD10+, 0%; IgM+, 93%; IgG+, 1%; λ+, 92%; κ+, 0%; CD1+ 0%; CD2+, 7%; CD3+, 5%; CD4+, 3%; CD5+, 9%; CD7+, 5%; CD8+, 2%; CD11c+, 3%; CD13+, 3%; CD14+, 1%; CD15+, 4%; CD33+, 1%; and CD34+, 1%. A diagnosis of a B cell lymphoproliferative disorder with IgM, λ light chain restriction was made. It was noted that the neoplastic cells lacked CD11c, CD10 and CD5 antigens.

Serum protein electrophoresis showed normal levels of IgG, IgA, and IgM. No M spike was present. Direct and indirect antoglobulin tests were negative.

The patient was started on chlorambucil, prednisolone and allopurinol. Her WBC counts varied between 163–224 × 10⁹/l with 95–99% lymphocytes. The patient became severely anaemic (haemoglobin concentration, 59 g/l) and thrombocytopenic (platelets, 43 × 10⁹/l). As the spleen continued to increase in size, a course of radiation was given without any response. Splenectomy was performed in February 1994.

The spleen was 2010 g in weight. Serial sections revealed multiple white nodules up to 0.3 cm in diameter, involving the whole spleen (fig 2). On histological examination, there was effacement of the white pulp by nodular aggregates of neoplastic lymphoid cells which coalesced in some areas. There was minimal involvement of the red pulp. The neoplastic lymphocytes were small to medium in size, with scanty cytoplasm and had round to slightly irregular nuclei, containing moderately coarse chromatin. Nucleoli were inconspicuous (fig 2). The mitotic count was three per high power field (× 400). The neoplastic cells did not show a marginal or a mantle zone pattern of arrangement. These cells stained positively for the CD20 antigen using the L-26 antibody. No staining was observed with the Leu-22 antibody.

Post-operative WBC counts fluctuated between 72 × 10⁹ and 161 × 10⁹/l with 84–90%
lymphocytes. The patient was placed on cyclophosphamide, vincristine, prednisolone, and doxorubicin therapy. At the last follow-up in May 1994, after two courses, her WBC count was 46 x 10^9/l with 95% lymphocytes.

**FAMILY HISTORY**

The patient's father had been diagnosed with CLL in October 1984 at 71 years of age when a routine work up revealed a total WBC count of 21.1 x 10^9/l with 81% lymphocytes. His haemoglobin concentration at diagnosis was 151 g/l and platelet count was 213 x 10^9/l. The bone marrow aspirate had 67% lymphocytes and the core biopsy revealed nodular aggregates of small lymphocytes. He did not receive any therapy for CLL before his death in May 1986 from an unrelated cause.

Information on the patient's brother was obtained from the necropsy report. He had been diagnosed with CLL in 1988 at the age of 45 years. The total leucocyte count in July 1990 was 94 x 10^9/l with 95% lymphocytes. The morphology of the leukaemic cells was described to be typical of CLL. Additionally, immunophenotypic studies carried out using flow cytometry revealed the following phenotype: HLA-DR+, 91%; k+, 91%; CD5, 100%; λ+, 9%; and CD2+, 6%. The patient underwent multiple courses of chemotherapy to which he was refractory. At his last admission in February 1993 he was leucopenic (WBC count 2.2 x 10^9/l) and thrombocytopenic (platelet count 15 x 10^9/l). Almost all of the leucocytes in the peripheral blood were large abnormal lymphocytes. He died soon thereafter of disseminated Aspergillus and Pseudomonas infections. Pertinent findings at necropsy were splenomegaly (weight 2000 g) with no mass lesions, and mediastinal and para-aortic lymphadenopathy. Leukaemic infiltrates were present in bone marrow, spleen and lymph nodes. The neoplastic cells were described as large and atypical.

**Discussion**

SLVL is a B cell lymphoproliferative disorder that usually presents with marked splenomegaly and moderate lymphocytosis. As CLL, PLL, HCL, and the leukaemic phases of mantle cell lymphoma (MCL) and small cleaved cell lymphoma (SCCL) can have similar clinicopathological features, a diagnosis of this disorder can only be made following detailed immunophenotypic and morphological, including electron microscopy, studies. B-CLL is the most common of the familial leukemias and therefore CLL was the first diagnostic consideration in our patient. The degree of peripheral lymphocytosis observed was also in keeping with this diagnosis, as in SLVL lymphocytosis is usually not very high. In a study of a large group of patients with SLVL the highest absolute lymphocyte count observed was 35 x 10^9/l. The lymphocytes in CLL, however, are small with a high nuclear to cytoplasmic ratio and clumped nuclear chromatin. Immunophenotypically, B-CLL cells are CD5+ in more than 90% of cases. The leukaemic cells in de novo PLL are usually CD5+ and patients usually present with massive splenomegaly. In our case, however, the morphology of the leukaemic cells was unlike that seen in PLL. Ultrastructural studies later demonstrated short polar villous projections, confirming a diagnosis of SLVL.

A close relationship has been suggested between HCL and SLVL, and TRAP positivity has been reported in SLVL. However, absence of CD11c antigen, nodular white pulp involvement of the spleen, and absence of circumferential villous projections on the leukemic cells excluded a diagnosis of HCL and variant HCL.

Splenomegaly is common in SCCL, and the spleen shows a miliary pattern of involvement on gross examination because of expansion of the white pulp, not unlike that seen in our case. Cytologically, however, SCCL shows a monotonous proliferation of lymphocytes with highly irregular nuclei (cleaved cells). Immunophenotypically, SCCL cells are usually positive for the CD10 antigen. MCL would be difficult to exclude on a purely morphological basis. The cytology of the neoplastic cells in MCL and SLVL is very similar. However, as with SCCL, splenic involvement in MCL is usually evident only when the lymphoma is disseminated and the patient has generalized lymphadenopathy. The peripheral blood counts, although not usually high, can reach 269 x 10^9 in MCL. Also, the neoplastic cells in MCL are usually CD5+. Demonstration of translocations involving 11q13 in MCL and approximately 20% of SLVL cases suggests these two disorders may be closely related, and we believe that the morphological features of the neoplastic cells supports this relationship.

We observed marked clumping of lymphoma cells in the peripheral blood smears prepared from EDTA anticoagulated blood. Such a phenomenon has been described in only one previous report, which incidentally described a patient with SLVL. These authors stated that this finding had never been described before, in either leukaemias or lymphomas involving the peripheral blood. The cause of this clumping remains uncertain but has been attributed to the presence of EDTA.

We believe that the most interesting aspect of this case is the occurrence of SLVL in a patient with family history of a B cell lymphoproliferative disorder. It is conceivable that the patient's brother and father also suffered from SLVL, particularly as we were able to find one previous report describing occurrence of SLVL in two sisters. However, with the limited material available for study on the relatives, this possibility is speculative. Absence of splenomegaly in the father does not exclude a diagnosis of SLVL. In the study by Melo et al only 18 of the 22 patients with SLVL had splenomegaly, and in another study by Oscier et al only 23 of 31 patients presented with an enlarged spleen. In addition, Bassan et al have reported a monoclonal B cell lymphocytosis.
without organomegaly as an early form of SLVL.

In case of the patient’s brother, presence of splenomegaly, refractoriness to chemotherapy and a possible transformation to large cell lymphoma would be consistent with a diagnosis of SLVL. Demonstration of CD5 antigen co-expression on B cells is not diagnostic of B-CLL. In a recent study of 100 SLVL cases, Matutes et al.13 found CD5 expression in 19% of the cases. Therefore, in the absence of detailed immunophenotypic and ultrastructural studies, we cannot exclude the possibility of SLVL in the patient’s relatives.

We conclude that a detailed work up of small B-cell lymphoproliferative disorders can help accurately classify these disorders and these distinctions may have important therapeutic implications.


Lipoprotein composition and serum Lp(a) lipoprotein in hypobetalipoproteinaemia

M Crook, R Swaminathan

Abstract
A family with hypobetalipoproteinaemia was studied to examine the Lp(a) lipoprotein, lipoprotein cholesterol, and triglyceride composition of the serum lipids. Lp(a) lipoprotein was measured by immunoassay. Serum lipoproteins were separated by ultracentrifugation. Cholesterol and triglycerides were measured using standard enzymatic assays. Serum apolipoprotein B was low and Lp(a) undetectable in the index patient and in her father and son. Separation of the lipoproteins by ultracentrifugation showed a low cholesterol content of serum low density lipoprotein in the affected family members and also a low triglyceride content of high density lipoprotein particles in two affected members. It is concluded that serum lipoprotein cholesterol is altered in hypobetalipoproteinaemia, and family members of index cases have undetectable serum Lp(a) lipoprotein concentrations.

Keywords: Hypobetalipoproteinaemia, Lp(a) lipoprotein, lipoprotein composition.

Hypobetalipoproteinaemia is a rare autosomal dominant condition characterised by low serum cholesterol concentrations. The exact biochemical defect is not known although truncated forms of apolipoprotein B (ApoB) have been described.1 Heterozygotes tend to be asymptomatic, whereas homozygous individuals may present with neurological sequelae similar to individuals with abetalipoproteinaemia. Lp(a) lipoprotein is considered to be a cardiovascular risk factor, yet how it exerts this action is unclear; possibly it acts in part by counteracting the fibrinolytic system.4 There are few published data upon Lp(a) lipo-
Splenic lymphoma with circulating villous lymphocytes.

F Imbing, Jr, D Kumar, S Kumar, G Yuoh and F Gardner

doi: 10.1136/jcp.48.6.584

Updated information and services can be found at:
http://jcp.bmj.com/content/48/6/584

Email alerting service
Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

Notes

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/