Expression of vascular endothelial growth factor in human oral squamous cell carcinoma: its association with tumour progression and p53 gene status

T Maeda, S Matsumura, H Hiranuma, A Jikko, S Furukawa, T Ishida, H Fuchihata

Abstract

Aims—To correlate vascular endothelial growth factor (VEGF) expression in oral squamous cell carcinoma with the clinico-pathological characteristics and prognosis; and to assess whether p53 gene status is associated with VEGF expression in human cancers.

Methods—Tumour specimens from 45 patients with oral squamous cell carcinomas were examined. Expression of VEGF was determined using an immunohistochemical method, and a tumour was considered positive when more than 5% of the neoplastic cells showed VEGF immunoreactivity. The p53 gene status was screened using a polymerase chain reaction–single strand conformation polymorphism analysis.

Results—VEGF positive staining was detected in 19 (42.2%) of the 45 cases. VEGF immunoreactivity did not correlate with the histological degree of tumour differentiation, clinical stages, or lymph node metastasis. The patients with VEGF positive tumours had a significantly worse prognosis than those with VEGF negative tumours. The five year overall survival rate of the VEGF negative patients was 76.5%, as compared with 48.8% for the VEGF positive patients. No significant association between VEGF expression and the p53 gene status of the tumours was found.

Conclusions—VEGF is a good prognostic indicator of the survival of patients with oral squamous cell carcinoma. The p53 gene status does not seem to be associated with VEGF expression in these cancers.

Keywords: vascular endothelial growth factor; p53; oral squamous cell carcinoma

Angiogenesis is an essential step in tumour growth and metastasis. It involves the formation of new blood vessels from pre-existing vessels by a multistep mechanism. Angiogenesis properties are correlated with tumour aggressiveness, and intratumour microvessel density has been found to be an independent prognostic factor. It is known that tumour cells can release diffusible angiogenic regulatory factors. Vascular endothelial growth factor (VEGF) is currently considered a leading candidate among the factors causing tumour angiogenesis. VEGF, which was first found to affect angiogenesis in malignant gliomas, stimulates the proliferation of endothelial cells and has been identified in many different tumours, particularly in hypoxic areas. The inhibition of VEGF signalling inhibits both tumour angiogenesis and the growth of experimental solid tumours.

It is thought that VEGF protein expression promotes tumour metastasis in human solid tumours. However, there is disagreement on the importance of VEGF in determining metastatic behaviour. With regard to the prognostic value of VEGF, some studies have shown that the expression of VEGF is an independent prognostic factor in patients with breast cancer. However, except in the case of breast cancer, few studies on the association between VEGF expression and cancer prognosis have been published.

Recent in vitro experimental studies have indicated that the p53 tumour suppressor gene is closely involved in the regulation of VEGF mRNA expression. These reports prompted us to investigate whether the VEGF expression pattern varies according to the p53 gene status in human cancers of patients.

We evaluated 45 cases of oral squamous cell carcinoma for the expression of VEGF by immunohistochemistry, and screened them for mutations of the p53 gene by polymerase chain reaction–single strand conformation polymorphism (PCR-SSCP) analysis. We investigated the correlations between the expression of VEGF protein and various clinicopathological factors including prognosis. In addition, we analysed the relation between VEGF expression and p53 gene status in these tumours.

Methods

PATIENTS AND SAMPLES

We examined 45 formalin fixed, paraffin embedded tissue samples of oral squamous cell carcinoma obtained by biopsy from 45 patients. The patients were enrolled at the Department of Radiology, Osaka University Dental Hospital, between October 1989 and February 1994. They were 28 males and 17 females, ranging in age from 32 to 87 years, with an average age of 63.7 years. All patients were staged according to the tumour–node–metastasis (TNM) classification of tumours (Union Internationale contre le Cancer (UICC), 1987). One patient had stage I, 18 had stage II, 13 had stage III, and 13 had stage IV tumours. All patients were fresh cases and...
had received no previous treatment. Of the 45 patients, 24 received radiotherapy alone and 21 received preoperative radiotherapy and then underwent surgery. Radiotherapy was provided by external beam megavoltage equipment, reaching a total dose of 50 to 70 Gy. For the preoperative radiotherapy, a total dose of 20 to 40 Gy was given. The histological degree of tumour differentiation was determined from haematoxylin and eosin stained sections by the hospital’s laboratory of clinical pathology.

VEGF IMMUNOSTAINING

The avidine–biotin complex method was used to detect VEGF immunoreactivity. In all cases, 5 µm thick paraffin embedded sections were cut and mounted on silane coated glass slides. The sections were deparaffinised in xylene and rehydrated through a series of graded ethanol, and then incubated with 0.3% hydrogen peroxide in methanol for 30 minutes to inhibit endogenous peroxidase activity. The slides were placed in a 0.01 M citrate buffer (pH 6) and heated in a microwave oven at 500 W for two periods of five minutes each. The sections were then washed in phosphate buffered saline (PBS) and incubated in 3% normal swine serum for one hour to reduce non-specific antibody binding. As a primary antibody, A-20 (Santa Cruz Biotechnolog) was used. A-20 is a rabbit polyclonal antihuman VEGF antibody. The specimens were then incubated with a 1: 200 dilution of A-20 overnight at 4°C. After three washes with PBS, the sections were incubated with secondary biotinylated swine anti-rabbit antibody (Dako) at a dilution of 1: 500 for one hour. The sections were then exposed to avidine–biotin peroxidase complex (Vector Laboratories) for one hour. Finally, the reaction was made visible by incubation in 0.05 M Tris-HCl buffer (pH 7.4) containing 0.04% diaminobenzidine and 0.003% hydrogen peroxide with 0.1% nickel ammonium sulphate intensification. With each batch of staining runs, a positive control consisting of a tissue section from human normal kidney was used. In addition, negative controls were processed by omitting the primary antibody.

EVALUATION OF IMMUNOHISTOCHEMICAL RESULTS

The results of the immunohistochemical staining were evaluated by two observers without prior knowledge of the patients’ clinicopathological data. An individual tissue section was considered positive if unequivocal immunoreactivity was seen in more than 5% of the tumour cells, as previously described.11

DNA PREPARATION

The extraction of DNA from formalin fixed, paraffin embedded carcinoma tissues was performed by the proteinase K technique as previously described.19

PCR-SSCP ANALYSIS AND DIRECT SEQUENCING

We screened p53 mutations in the 45 tumours from exon 5 to 8 using a PCR-SSCP analysis. The PCR was performed in a 50 µl reaction mixture containing 100 ng genomic DNA, 20 pmol of each primer, 200 µM each of dATP, dTTP, and dGTP, 20 µM dCTP, 1 µCi [32P]dCTP (3000 Ci/mmol), 10 mM Tris-HCl (pH 8.3), 1.5 mM MgCl2, 50 mM KCl, and 2.5 units of Ampli-Taq polymerase (Perkin-Elmer). Thirty cycles of denaturation (94°C, 30 seconds), annealing (52–60°C, one minute), and extension (72°C, two minutes) were carried out in a DNA thermal cycler (Perkin-Elmer). We used synthesised oligomer primers for exons 5 to 8 of the p53 gene. The primer sequences were: exon 5: 5'-TACTTCCCTGCCCTCAACAA-3' and 5'-ATCGCTATCTGAGCAGCGCT -3'; exon 6: 5'-GATTGCTCTTAGGTCTGGCC-3' and 5'-CTGACAACCACCCTTAACCC-3'; exon 7: 5'-ACTGGCCTCATCTTGGGCCT -3' and 5'-TGTGCAGGGTGGCAAGTGGC-3'; exon 8: 5'-TAAATGGGACAGGTAGGACC-3' and 5'-TCCACCGCTTCTTGTCCTGC-3'. After the PCR, the reaction mixtures were diluted threefold with 0.1% sodium dodecyl sulphate (SDS) plus 10 mM EDTA, and mixed with the same volume of loading dye solution. Following denaturation at 95°C for five minutes, the diluted samples were applied to 5% neutral polyacrylamide gels with 5% glycerol, and elec-

![Figure 1](image1.png)

**Figure 1** Representative examples of VEGF immunostaining in oral squamous cell carcinoma. There is a strong cytoplasmic staining of the tumour cells. (**Magnification >100.**)

![Figure 2](image2.png)

**Figure 2** Typical examples of detection mutations by PCR-SSCP analysis in exons 5 and 8 of the p53 gene in oral squamous cell carcinoma (cases 21 and 37, respectively). Compared with the normal control (N), bands, abnormal band shifts can be seen in the tumours (T).
vascular endothelial growth factor and oral squamous cell carcinoma

We used as negative controls of band shifts. DNA samples from healthy human tissue were extracted from abnormal bands and directly sequenced using a dye terminator cycle sequencing FS kit (Applied Biosystems), according to the manufactures’ instructions. Normal DNA samples from healthy human tissue were used as negative controls of band shifts.

trophoresis was performed at a 40 W constant power for two hours at room temperature. The gel was dried on filter paper and exposed to x-ray film at −80°C. To confirm each mutations, DNAs were extracted from abnormal bands and directly sequenced using a dye terminator cycle sequencing FS kit (Applied Biosystems), according to the manufactures’ instructions. Normal DNA samples from healthy human tissue were used as negative controls of band shifts.

**Statistical analysis**

The relation between VEGF expression and the clinicopathological characteristics or p53 gene status was examined by the χ² test. The Kaplan–Meier survival method was used for univariate analysis and the log rank test was used to assess differences between the two groups (VEGF positive and negative). Statistical significance was defined as p < 0.05. The StatView software (Abacus Concepts) was used for statistical analysis.

**Results**

**VEGF Immunostaining**

The tumour specimens obtained from the 45 patients with oral squamous cell carcinoma were analysed immunohistologically for the expression of VEGF. VEGF immunoreactivity was mainly localised to the cytoplasm of the tumour cells (fig 1). Nineteen carcinomas (42.2%) were VEGF positive and 26 (57.8%) were negative.

The prognosis of the 45 patients was examined. We found it was worse in the patients with VEGF positive tumours than those with VEGF negative tumours. The overall survival rates of the VEGF negative and positive patients were significantly different (p = 0.02). The five year overall survival rate of the VEGF negative patients was 76.5%, compared with 48.8% for the VEGF positive patients (fig 3).

**Discussion**

Neovascularisation nourishes a growing tumour and also allows tumour cells to be in contact with the vascular bed of the host. Tumour angiogenesis is regulated by factors released either by the tumour cells themselves or by macrophages attracted into the tumour proper. Much work has been done in recent years to identify angiogenic factors. VEGF is the only angiogenic peptide known to act specifically on endothelial cells and is therefore considered to be the leading candidate of the

---

**Table 1** Relation between vascular endothelial growth factor (VEGF) expression and clinicopathological characteristics

<table>
<thead>
<tr>
<th>Variable</th>
<th>VEGF Negative</th>
<th>VEGF Positive</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Differentiation</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Good</td>
<td>10</td>
<td>4</td>
<td>NS</td>
</tr>
<tr>
<td>Moderate</td>
<td>13</td>
<td>12</td>
<td></td>
</tr>
<tr>
<td>Poor</td>
<td>3</td>
<td>3</td>
<td></td>
</tr>
<tr>
<td>Lymph node</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>metastasis</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Negative</td>
<td>15</td>
<td>10</td>
<td>NS</td>
</tr>
<tr>
<td>Positive</td>
<td>11</td>
<td>9</td>
<td></td>
</tr>
<tr>
<td>Clinical stage</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>I</td>
<td>1</td>
<td>0</td>
<td>NS</td>
</tr>
<tr>
<td>II</td>
<td>12</td>
<td>6</td>
<td></td>
</tr>
<tr>
<td>III</td>
<td>6</td>
<td>7</td>
<td></td>
</tr>
<tr>
<td>IV</td>
<td>7</td>
<td>6</td>
<td></td>
</tr>
<tr>
<td>Primary treatment</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>RT</td>
<td>13</td>
<td>11</td>
<td>NS</td>
</tr>
<tr>
<td>RT + surgery</td>
<td>13</td>
<td>8</td>
<td></td>
</tr>
</tbody>
</table>

RT, radiotherapy.

---

**Table 2** Vascular endothelial growth factor (VEGF) expression according to p53 gene status

<table>
<thead>
<tr>
<th>p53 Status</th>
<th>VEGF expression</th>
<th>Wild-type</th>
<th>Mutant type</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Negative</td>
<td>18</td>
<td>8</td>
<td>NS</td>
<td></td>
</tr>
<tr>
<td>Positive</td>
<td>9</td>
<td>10</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

p53 GENE MUTATIONS

We identified p53 mutations in 18 (40.0%) of the 45 tumour specimens. We found seven, two, four, and six mutational alterations in exons 5, 6, 7, and 8, respectively. One specimen had mutations in both exon 7 and 8. Typical examples of the PCR-SSCP of exons 5 and 8 are shown in fig 2.

RELATION BETWEEN VEGF EXPRESSION AND CLINICOPATHOLOGICAL CHARACTERISTICS

The correlations between VEGF expression and various clinicopathological characteristics are shown in table 1. As for the primary treatment method, no bias was found between the VEGF positive and VEGF negative groups. There were no significant associations between VEGF expression and the histological degree of tumour differentiation or clinical stages.

Cervical lymph node metastasis was observed in 20 of the 45 patients at diagnosis. Eleven of the 26 VEGF negative patients (42.3%) developed lymph node metastasis, compared with nine of the 19 VEGF positive patients (47.4%). No significant correlation was found between VEGF expression and the incidence of lymph node metastasis.

The prognosis of the 45 patients was examined. We found it was worse in the patients with VEGF positive tumours than those with VEGF negative tumours. The overall survival rates of the VEGF negative and positive patients were significantly different (p = 0.02). The five year overall survival rate of the VEGF negative patients was 76.5%, compared with 48.8% for the VEGF positive patients (fig 3).

---

**Figure 3** Kaplan–Meier curves for the overall survival of the patients according to the expression of vascular endothelial growth factor (VEGF). The two curves are significantly different.
factors playing a key role in tumour angiogenesis. Raised VEGF levels are frequently observed in tumour cells located near necrotic regions within the tumour, and it has been suggested that hypoxia may induce VEGF expression. In addition, Kim et al reported that tumour growth was suppressed by anti-VEGF treatment in vivo. VEGF expression has been found in a wide variety of tumour cells, and is detected mainly in the cytoplasm of the cells. In breast carcinomas and gastric carcinomas, approximately half the tumour cells are VEGF positive. In the present study, 19 (42.2%) of 45 oral squamous cell carcinomas showed cytoplasmic VEGF protein expression. There was no significant association between VEGF expression and the histological degree of tumour differentiation or the clinical stage. The mechanism of metastasis is thought to be a complex of multiple steps including the degradation of extracellular matrix, the intravasation of tumour cells, and the attachment to distant organs. The role of VEGF in tumour angiogenesis remains unclear. An in vivo experimental study with VEGF transfected ovarian cells showed that VEGF did not make a tumour metastatic, whereas Claffey et al showed that VEGF transfection increased experimental metastasis of human melanoma cells. In a clinical study, Maeda et al reported a significant correlation between VEGF expression and lymph node metastasis or liver metastasis in patients with gastric carcinoma, whereas no significant correlation was detected between VEGF expression and lymph node metastasis in lung carcinoma. In our present study, the expression of VEGF did not correlate with lymph node metastasis. VEGF may not influence lymphatic tumour metastasis. We observed that VEGF expression in oral squamous cell carcinoma had prognostic significance. The prognosis of the patients with VEGF positive tumours was significantly worse than that of the patients with VEGF negative tumours. This is consistent with previous reports indicating that VEGF expression is an independent prognostic factor in patients with breast carcinoma, lung carcinoma, or gastric carcinoma. These data suggest that the presence of VEGF expression may be a good prognostic indicator, along with other clinicopathological factors, in patients with oral squamous cell carcinoma. Little is known about the regulation of VEGF expression. Hypoxia is thought to be one factor which promotes the expression of VEGF. There may well be other mechanisms of regulation for this important angiogenic protein. The tumour suppressor gene p53 encodes a nuclear phosphoprotein that is associated with cell cycle regulation, cellular differentiation, the suppression of abnormal cell proliferation, and tumour development. Since mutations of the p53 gene have been found in various types of human cancer, p53 has been considered a marker of both malignancy and tumour progression. The progression from low grade astrocytoma to the highly malignant glioblastoma involves the clonal expansion of p53 mutant cells. During the progression of these tumours towards high grade malignancy, an increase of neovascularisation has also been observed. This finding suggests a possible correlation between p53 mutations and enhanced production of VEGF. Recent in vitro studies have indicated that the p53 gene is closely involved in the regulation of VEGF. Kieser et al showed that the induction of VEGF mRNA and protein by activated protein kinase C was strongly synergistic with mutant but not wild-type p53 gene. However, Mukhopadhyay et al reported that wild-type p53 down-regulated the endogenous VEGF mRNA level, whereas mutant forms of p53 had no effect. These observations prompted us to investigate whether there is a significant correlation between the VEGF expression and p53 gene status in human solid tumours. To our knowledge, this is the first such study in human patients. We screened the oral squamous cell carcinoma specimens for p53 gene status using a PCR-SSCP analysis instead of an immunohistochemical examination, because the immunohistochemical staining of p53 protein is not always dependent on p53 gene mutations. Of the 18 tumours with p53 gene mutations, 10 (55.6%) showed VEGF positive staining, whereas of the 27 tumours without p53 gene mutations, nine (33.3%) showed positive staining. Contrary to our expectation, we found no significant correlation between p53 gene status and VEGF expression. However, there was a trend towards a positive correlation between p53 mutations and VEGF expression. Further investigation is necessary to determine the effect of the p53 gene on VEGF expression. We believe that important information about the genetic regulation of angiogenesis in tumours will be acquired by more detailed in vitro and in vivo studies. In conclusion, our present results indicate that the expression of VEGF may be a good prognostic indicator in patients with oral squamous cell carcinoma. The examination of VEGF expression before treatment may be useful in assessing the biological behaviour of such tumours and thereby suggesting the most effective treatment.

This study was partly supported by a grant-in-aid from the Ministry of Education, Culture and Science of Japan (No 07557367).
Vascular endothelial growth factor and oral squamous cell carcinoma

T Maeda, S Matsumura, H Hiranuma, A Jikko, S Furukawa, T Ishida and H Fuchihata

J Clin Pathol 1998 51: 771-775
doi: 10.1136/jcp.51.10.771

Updated information and services can be found at:
http://jcp.bmj.com/content/51/10/771

These include:

Email alerting service
Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

Notes

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/