Infection by *Rhodococcus equi* in a patient with AIDS: histological appearance mimicking Whipple’s disease and *Mycobacterium avium-intracellulare* infection

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**Abstract**

*Rhodococcus equi* pneumonia with systemic dissemination is being reported increasingly in immunocompromised patients. This is the first case report of disseminated *R equi* infection with biopsy documented involvement of the large intestine. The patient was a 46 year old male with AIDS who was diagnosed with cavitating pneumonia involving the left lower lobe. *R equi* was isolated in culture from the blood and lung biopsies. Subsequently, the patient developed anaemia, diarrhoea, and occult blood in the stool. Colonoscopy revealed several colonic polyps. Histological examination of the colon biopsies showed extensive submucosal histiocytic infiltration with numerous Gram positive coccobacilli and PAS positive material in the histiocytes. Electron microscopy showed variably shaped intracellular organisms which were morphologically consistent with *R equi* in the specimen. Disseminated *R equi* infection may involve the lower gastrointestinal tract and produce inflammatory polyps with foamy macrophages which histologically resemble those seen in Whipple’s disease and *Mycobacterium avium-intracellulare* infection.

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**Keywords:** *Rhodococcus equi*, AIDS, colonic disease

**Case report**

A 46 year old HIV positive white male was admitted to the Medical College of Ohio hospital in late October 1996, complaining of fever, night sweats, dry cough, and weight loss. Previously, the patient underwent a bronchoscopy in May 1996 in response to a mass in the left lower lobe discovered on routine chest x ray. A culture at the time of bronchoscopy grew *R equi*, which was sensitive to penicillin, erythromycin, trimethoprim-sulphamethoxazole (TMP-SMX), rifampicin, and vancomycin. It was resistant to clindamycin. His clinical condition improved moderately but not fully, after taking TMP-SMX, rifampicin, and clarithromycin daily for several months.

Biopsies from a subsequent bronchoscopy in September 1996 showed a severe active, chronic pneumonitis with *Gram positive coccobacilli* within foamy histiocytes. *R equi* grew from cultures of these transbronchial biopsies as well as from bronchial washings. Three blood cultures drawn at the same time also grew *R equi*. All of these bacterial isolates from both the lung and blood cultures were sensitive to the same antibiotics as in May 1996, except that all isolates were now uniformly resistant to penicillin.

Because the lung lesion had not completely resolved and the patient was still having cough and fever despite taking the antibiotics mentioned, it was agreed to resect the affected lobe. A presurgical evaluation showed mild anaemia and occult blood present in the stool. The patient also complained of chronic diarrhoea and night sweats. The patient was admitted to the Medical College of Ohio in May 1996 in response to a mass in the left lower lobe discovered on routine chest x ray. A culture at the time of bronchoscopy grew *R equi*, which was sensitive to penicillin, erythromycin, trimethoprim-sulphamethoxazole (TMP-SMX), rifampicin, and vancomycin. It was resistant to clindamycin. His clinical condition improved moderately but not fully, after taking TMP-SMX, rifampicin, and clarithromycin daily for several months.

While in the hospital, the patient slowly began losing his short term memory. Magnetic resonance imaging of the brain showed several enhancing lesions in both hemispheres. In addition to his current three-antibiotic regi-
men, pyrimethamine was added to cover for possible infection by *Toxoplasma gondii*. In December 1996, computed tomography (CT) of the brain was performed because of the onset of seizure activity. This scan showed several additional ring enhancing lesions. Vancomycin and clindamycin were added for additional *R equi* and *Toxoplasma gondii* coverage, respectively. At a subsequent hospital admission in February 1997, multiple biopsies were obtained from several of the brain lesions to determine their cause. These samples showed an inflammatory microangiopathy on routine haematoxylin and eosin stains. A Gram stain revealed scant Gram positive rod shaped microorganisms consistent with *R equi*. No *Toxoplasma gondii* organisms were seen. A repeat CT scan of the brain showed that the lesions were resolving after approximately one month of vancomycin. Subsequently, all antibiotics were discontinued except for vancomycin and rifampicin. After two months of this treatment, his neurological symptoms had resolved and another CT scan of the brain showed that nearly all the brain lesions had cleared. There was also no recurrence of pneumonia.

**Pathological findings**

All three biopsies taken from the caecum, ascending colon, and transverse colon showed massive infiltration of histiocytes within the submucosa (fig 1). These histiocytes had abundant eosinophilic cytoplasm which stained strongly with periodic acid Schiff (PAS) stain. A Gram stain revealed many intracytoplasmic Gram positive coccobacilli. A silver stain for fungal organisms was negative, and an acid-fast stain was unremarkable in that only a few organisms stained faintly positive. Electron microscopy studies revealed oval shaped bacte-

![Figure 1](https://group.bmj.com/group.bmj.com/resource/1342-5238-19-4-69-g001f1a)

**Figure 1** Endoscopic colon biopsy showing extensive infiltration of foamy histiocytes with abundant eosinophilic PAS positive cytoplasm. Left, H&E stain low power; middle, H&E stain high power; right, PAS stain.

![Figure 2](https://group.bmj.com/group.bmj.com/resource/1342-5238-19-4-69-g002f2)

**Figure 2** Transmission electron microscopy of colon biopsy (right) and lung biopsy (left), showing intracytoplasmic coccobacilli with thick cell walls and lipid vacuoles (×59 325).
ria with a thick cell wall and multiple lipid vacuoles, very similar to the morphology of mycobacteria (fig 2). There were no aggregates of membranous material to suggest that the organisms were Whipple’s bacilli.

The left lower lobe of the lung measured $13.0 \times 11.0 \times 6.0$ cm and weighed 630 g. The pleural surface had many fine adhesions and a $5.0 \times 4.5$ cm area of subpleural haemorrhage was also present. The entire lobe was consolidated by a soft, creamy, yellowish-tan pasty material. Architecturally, the lobe resembled a liver with macronodular cirrhosis. Microscopically, there was obliteration of the normal lung parenchyma with dense infiltration of histiocytes. The outline of alveoli was observed only occasionally, and when they were seen, they were nearly filled with histiocytes. Similar to those in the intestinal wall, these histiocytes had abundant, eosinophilic, globular cytoplasm (fig 3). A Gram stain showed the presence of Gram-positive coccobacilli within the cytoplasm of the histiocytes. The organisms did not stain definitively with an acid-fast stain, and a silver stain for fungal organisms was negative. These histiocytes were likewise strongly PAS positive. Electron microscopy of a portion of the lung tissue showed many intracytoplasmic coccobacilli with thick cell walls and intracytoplasmic lipid vacuoles (fig 2). No Michaelis-Gutmann bodies were identified to suggest malakoplakia. As mentioned cultures from this tissue grew *R equi*.

**Discussion**

Human infection with *R equi* is now increasingly being reported in immunocompromised patients as a result of HIV infection, haematological malignancies, and immunosuppression for transplantation. Rare cases have also been reported in patients with functioning immune systems. *R equi* is usually transmitted by inhalation and commonly produces a cavitating pneumonia. So far as we know, this is the first report describing a case of disseminated *R equi* infection with histologically documented involvement of the gastrointestinal tract.

When there is intense infiltration of foamy eosinophilic histiocytes within the wall of the intestine, Whipple’s disease and infection by *Mycobacterium avium-intracellulare* are two main infections which should be considered in the differential diagnosis. With this case, we show that the inflammatory response elicited by *R equi* in the intestine produces a near identical histological picture to *Mycobacterium avium-intracellulare* infection and Whipple’s disease. Culture of the colon biopsies were not ordered. However, the morphological appearance of the organisms by light and electron microscopy was identical to those seen in lung biopsies which grew *R equi*. Furthermore, the documentation of systemic spread was confirmed by positive blood cultures. Indeed, an acid-fast stain, which would be expected to be strongly positive in *Mycobacterium avium-intracellulare* infection, was weakly positive in focal areas only, consistent with *R equi*. Although the histiocytes were strongly PAS positive, electron microscopy failed to reveal the characteristic *Tropheryma whippelli* bacilli seen in Whipple’s disease. In addition, Whipple’s disease involving the gastrointestinal tract more often affects the small rather than the large intestine. Also, Gram staining showed numerous intracytoplasmic Gram-positive coccobacilli whereas *T whippelli* is Gram negative.

In conclusion, *R equi* infection should be included in the differential diagnosis along with Whipple’s disease and *Mycobacterium avium*.

![Figure 3](http://jcp.bmj.com/)
intracellulare whenever foamy eosinophilic histiocytes are seen in colon biopsies from an immunocompromised patient.

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