A latex slide agglutination test for rapid detection of antimyeloperoxidase antibody

K H Ko, S S Lee, J W M Lawton

Abstract

Aim—To develop and test a new latex slide agglutination test (MPO-LSAT) to detect antimyeloperoxidase (anti-MPO) antibody in serum.

Methods—Latex bead coating was adjusted to give maximum sensitivity by attending to latex size, MPO to latex ratio for coupling, ratio of diluted serum to MPO-latex, reaction time and temperature for coupling, and reaction time for agglutination. Inhibition studies were performed using MPO, proteinase 3, bactericidal/permeability increasing protein, and lactoferrin.

Results—There was very good correlation between this test and the conventional anti-MPO enzyme linked immunosorbent assay (ELISA): 81% of sera positive in the ELISA were positive by MPO-LSAT. MPO-LSAT results correlated better with IgM anti-MPO than with IgG anti-MPO.

Conclusions—MPO-LSAT is a simple diagnostic test that is potentially useful in the clinical laboratory as a rapid screening tool for vasculitic diseases.

Keywords: latex agglutination; antineutrophil cytoplasmic antibody; anti-MPO antibody

Antineutrophil cytoplasmic antibody (ANCA) comprises a group of autoantibodies first reported by Davies et al in 1982.1 Its association with systemic vasculitis and the classification into C-ANCA and P-ANCA forms have become established.2 Up to now, indirect immunofluorescence has remained the standard method for ANCA detection, using cyto-centrifuged, alcohol fixed human neutrophils as substrate. Myeloperoxidase (MPO), one major target antigen of ANCA found in some vasculitic diseases, such as necrotising and crescentic glomerulonephritis and microscopic vasculitic diseases, such as necrotising and crescentic glomerulonephritis and microscopic vasculitic diseases, such as necrotising and crescentic glomerulonephritis and microscopic vasculitic diseases, such as necrotising and crescentic glomerulonephritis and microscopic vasculitic diseases, such as necrotising and crescentic glomerulonephritis and microscopic vasculitic diseases, such as necrotising and crescentic glomerulonephritis and microscopic vasculitic diseases, such as necrotising and crescentic glomerulonephritis and microscopic vasculitic diseases, such as necrotising and crescentic glomerulonephritis and microscopic vasculitic diseases, such as necrotising and crescentic glomerulonephritis and microscopic vasculitic diseases, such as necrotising and crescentic glomerulonephritis and microscopic vasculitic diseases, 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at 25°C. After washing three times in glycine buffer (1% bovine serum albumin in glycine buffered saline, pH 8, with 0.1% Tween 20), coated latex was suspended in glycine buffer at a concentration of 0.5%. MPO coated latex was stored at 4°C until use.

Serum anti-MPO was detected by passive slide agglutination. Briefly, 5 µl of MPO–latex were added to 10 µl diluted serum on the dark background glass slide. After rotating the glass slide for two minutes by hand, the agglutination was read by eye. The strength of agglutination was graded as −, +/−, +, ++, or ++++. The titre of serum was defined as the dilution which gave a + result. All sera were tested neat and at 1/4 dilution to exclude “prozone effect.” Positive sera were then further titrated by twofold serial dilution to determine the titre.

Results

The development of the new anti-MPO latex agglutination test required optimisation of the assay conditions. The single most important factor was establishment of the optimum MPO to latex concentration ratio. Owing to potential batch variation, titration of MPO was necessary for each new batch. For this batch of MPO, a combination of 100 µl MPO (0.5 mg/ml) and 50 µl latex (10%) gave the best results. The stability of MPO–latex during storage was monitored by sequential testing of aliquots of standard anti-MPO positive serum, stored at −70°C. MPO–latex was found to be stable at 4°C, without any change of titre of reference serum, for at least six months (data not shown).

In terms of positivity and negativity, correlation of the MPO-LSAT with the conventional ELISA was excellent. However, absolute values correlated only weakly in the IgM assay (fig 1). MPO-LSAT did not react with normal human sera, anti-MPO negative P-ANCA sera, and C-ANCA positive sera except for one sample (table 1). The latter belonged to a C-ANCA positive patient and tested strongly positive for anti-MPO IgM (binding activity > 100%). In P-ANCA positive sera, 35 of 43 anti-MPO IgG positive samples were detected by MPO-LSAT. Agglutination negative samples had low MPO binding in the IgG ELISA (binding activity < 6%) and no significant activity in the IgM anti-MPO ELISA. Thus, using MPO IgG ELISA results as the reference, the sensitivity of MPO-LSAT was 81%.

It was confirmed that MPO–latex was specifically agglutinated by anti-MPO, but not by anti-PR3, anti-bactericidal/permeability increasing protein (BPI) or anti-lactoferrin. This agglutination ability of anti-MPO could be inhibited by MPO but not by PR3, BPI, or lactoferrin.

A sample strongly positive in MPO-LSAT was further evaluated by serial dilution. MPO–latex did not agglutinate in the neat serum, owing to the prozone effect. In view of this phenomenon, a 1/4 dilution of test serum was routinely included in MPO-LSAT.

Finally, MPO–latex was agglutinated better by IgM anti-MPO than by IgG. MPO-LSAT titre (r = 0.51, p < 0.001) correlated with IgM anti-MPO binding in ELISA but less so with IgG anti-MPO (r = 0.12, p = 0.45) (fig 1).

Discussion

Patients with anti-MPO associated vasculitis often present with severe illness, such as acute renal failure. Access to early and reliable testing is crucial in guiding subsequent clinical management. This is particularly important in Hong Kong where anti-MPO is the commonest autoantibody found in patients with small vessel vasculitis. The development of the latex
test therefore contributes towards a more efficient and effective system in the diagnosis of anti-MPO positive vasculitis.

The new MPO-LSAT is a simple procedure, the performance of which is comparable with the conventional assay by ELISA. It is a rapid diagnostic tool, the results of which can be generated within minutes. Technically, the test is simple to perform and results are easy to read. The procedure’s speed and simplicity are particularly important when an urgent result is needed in a clinical laboratory.

To exclude confounding results in the presence of other autoantibodies, we tested 20 antinuclear antibody positive sera and 20 rheumatoid factor positive sera. All were negative by IgG anti-MPO ELISA. Two rheumatoid factor positive sera proved to be positive in the MPO-LSAT and the IgM anti-MPO ELISA. Thus rheumatoid factor may occasionally give rise to false positive results.

MPO-LSAT has a very high specificity and correlates positively with anti-MPO ELISA, especially with the ELISA for IgM. IgM anti-MPO antibody had been reported to be associated with clinical manifestation such as pulmonary haemorrhage1 but it is not commonly assayed in clinical laboratories. MPO-LSAT may prove to be a useful laboratory diagnostic tool in IgM anti-MPO associated disease. The stability of the test reagent and reproducibility of results add value to the simple assay system. Its preferential reaction with IgM anti-MPO would need to be further investigated to determine its clinical relevance.

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