Recent advances in antiviral therapy

Derek Kinchington

Abstract
In the early 1980s many institutions in Britain were seriously considering whether there was a need for specialist departments of virology. The arrival of HIV changed that perception and since then virology and antiviral chemotherapy have become two very active areas of biomedical research. Cloning and sequencing have provided tools to identify viral enzymes and have brought the day of the “designer drug” nearer to reality. At the other end of the spectrum of drug discovery, huge numbers of compounds for screening can now be generated by combinatorial chemistry. The impetus to find drugs effective against HIV has also stimulated research into novel treatments for other virus infections including herpesvirus, respiratory infections, and hepatitis B and C viruses. The need to understand the function of the immune system during HIV infection has brought virologists and immunologists together into new partnerships. The huge increase in activity in antiviral research is reflected in the frequency with which these drugs are now being licensed: in 1985 there were two licensed antiviral drugs for systemic use. Since then approximately 20 compounds have been licensed and more are being submitted to the regulatory authorities on a regular basis.

Keywords: antiviral agents; viruses; immunology

Drugs for treating HIV infection
In 1985/86 the nucleoside analogue 3’-azido-3’-deoxythymidine (AZT, or zidovudine, ZDV) was discovered as an inhibitor of the reverse transcriptase (RT) enzyme of HIV,1 the first of a line of antiviral agents used in the treatment of AIDS (table 1). This molecule blocks the formation of the RNA/DNA intermediate and prevents the double stranded proviral DNA from integrating into the host cell genome. The compound is given in an inactive form and is converted sequentially by cellular enzymes to the monophosphate, the diphosphate, and finally the bioactive triphosphate. Thus inactive zidovudine becomes the active zidovudine triphosphate on absorption, and is a substrate for reverse transcriptase. Incorporation of the phosphorylated form into the growing viral genome through 5’–3’ ester linkages between adjacent sugars blocks any further chain elongation.

The early success achieved with zidovudine in the management of HIV was short lived because of the development of drug resistance.4 Experience in the field of antimicrobials indicated that using a combination of drugs might overcome this problem. The only available drugs during the late 1980s were two other nucleotide reverse transcriptase inhibitors (NRTI) which also targeted HIV reverse transcriptase (HIV-RT): 2’,3’-dideoxycytidine (ddC) and 2’,3’-dideoxyinosine (ddI).5,6 In vitro combination studies gave surprising results: those viruses that became highly resistant to ZDV remained sensitive to both ddC and ddI.7 Furthermore, neither cross resistance nor interference between the drugs was an issue, and subsequent clinical experience showed that patients benefited when these two compounds were used in combination with ZDV.8 It was also found by in vitro studies that virus isolated from patients on long term ZDV monotherapy had become insensitive to ZDV, but regained sensitivity when these patients were switched to ddI monotherapy. Although the virus retained the ZDV resistance genotype, the mutation conferring resistance to ddI suppressed the ZDV resistance phenotype.7

A great improvement in nucleoside analogue combinations was the introduction of 3’-thiaribofuranosyl-βL-cytosine (3TC or Epivir) into the regimens. This compound is a more potent inhibitor of HIV than either ddI or ddC, but when used as monotherapy 3TC selects for resistant strains very rapidly. However, studies of isolates from patients on ZDV/3TC combination treatment showed that the mutation in the reverse transcriptase gene conferring 3TC resistance also greatly delayed the generation of resistance to ZDV.9

The use of cloned HIV-RT, in cell-free systems, as a target for anti-HIV drugs allowed the identification of a large number of non-nucleoside reverse transcriptase inhibitors (NNRTI) which selectively inhibited HIV-1 reverse transcriptase. These compounds were found to bind to reverse transcriptase in regions outside the nucleoside binding pocket. The first of these NNRTI were the TIBO derivatives, and the most potent compounds had a similar antiviral activity to ZDV.10 These compounds, although very potent, induced resistance after a few days of use and for a while interest was lost in their development. Their resurgence as useful drugs occurred once the principles of cross resistance between the various NRTI and the NNRTI were formulated at the molecular level. Two compounds, loviride and nevirapine, have been developed for clinical use11,12 and these two classes of drugs, in combination with NRTI and protease inhibitors, have shown clinical efficacy.13

The third development, which extended the repertoire of drugs for anti-HIV treatment, was the discovery that HIV encoded an aspartyl protease. The synthesis of inhibitors against
this enzyme quickly followed and saquinavir, the first of these protease inhibitors (PI), reached clinics in 1991. These are the most structurally complex of the compounds used in the treatment of HIV infections. Since then ritonavir and indinavir have also been licensed and nelfinavir and the new soft gel formulation, saquinavir-SGC, have undergone clinical trials and are licensed.

The function of HIV proteinase is to cleave the large HIV gag-pol precursor protein into all the structural proteins of the virion and the virus specific enzymes. These proteins are protease itself (through an auto-enzymatic step), reverse transcriptase (incorporating RNAse H), and integrase. The PI are polypeptide analogues of the natural substrates cleaved by HIV protease, and all these drugs have been optimised to bind with high affinity to the active site of the enzyme, thus blocking essential proteolytic steps needed for maturation of the virion. They are very potent.

Table 1 Some approved drugs for treatment of HIV and other virus infections

<table>
<thead>
<tr>
<th>Generic name</th>
<th>Abbreviated and Proprietary name</th>
<th>Principal activities</th>
<th>Description</th>
</tr>
</thead>
<tbody>
<tr>
<td>Nucleotide reverse transcriptase inhibitors (NRTI)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Didanosine</td>
<td>ddI Videx</td>
<td>HIV-1 and HIV-2</td>
<td>Purine NRTI used for the treatment of advanced HIV disease and in combination</td>
</tr>
<tr>
<td>Lamivudine</td>
<td>3TC Epivir</td>
<td>HIV-1, HIV-2, and HBV</td>
<td>Pyrimidine NRTI used in combination</td>
</tr>
<tr>
<td>Stavudine</td>
<td>D4T Zerit</td>
<td>HIV-1 and HIV-2</td>
<td>Pyrimidine NRTI, used for adults with advanced disease who are intolerant to other approved therapies or in combination</td>
</tr>
<tr>
<td>Zalcitabine</td>
<td>ddC Hivid</td>
<td>HIV-1 and HIV-2</td>
<td>Pyrimidine NRTI, used for adults with advanced disease who are intolerant to ZDV treatment in combination</td>
</tr>
<tr>
<td>Zidovudine</td>
<td>AZT, ZDV Retrovir</td>
<td>HIV-1 and HIV-2</td>
<td>Pyrimidine NRTI, used for the treatment of adults and children with HIV disease</td>
</tr>
<tr>
<td>Non-nucleotide reverse transcriptase inhibitors (NNRTI)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Delavirdine</td>
<td>U-90152S Rescriptor</td>
<td>HIV-1</td>
<td>Bis-heteroaryl-piperazine (BHAP) derivative, in phase III trials</td>
</tr>
<tr>
<td>Nevaripine</td>
<td>BI-RG 587 Viramune</td>
<td>HIV-1</td>
<td>Dipyrirdiazone NNRTI used in combination with NTRI</td>
</tr>
<tr>
<td>Proteinase inhibitors (PI)</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Indinavir</td>
<td>MK-639 Crixivan</td>
<td>HIV-1 and HIV-2</td>
<td>Hydroxamino-pentene amide derivative, used in combination with NRTI or as monotherapy</td>
</tr>
<tr>
<td>Nelfinavir</td>
<td>AG-1343 Viracept</td>
<td>HIV-1 and HIV-2</td>
<td>Non-peptide PI used, in combination with NRTI or as monotherapy</td>
</tr>
<tr>
<td>Nitonavir</td>
<td>ABT-538 Norvir</td>
<td>HIV-1 and HIV-2</td>
<td>C2 symmetry-based PI, used in combination with NRTI or as monotherapy</td>
</tr>
<tr>
<td>Saquinavir</td>
<td>R0 31-8959 Invirase</td>
<td>HIV-1 and HIV-2</td>
<td>Hydroxyethyl amine derivative used in combination with NRTI</td>
</tr>
<tr>
<td>Soft gel formulation: Saquinavir-SGC</td>
<td>Fortovase</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Some approved drugs for other viruses</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Acyclovir</td>
<td>ACV Zovirax</td>
<td>HSV-1, HSV-2, VZV, EBV and CMV</td>
<td>Purine nucleoside analogue, used in the treatment of mucosal, cutaneous and systemic HSV-1 and HSV-2; also used for the prophylaxis of HSV infections (genital herpes), VZV and CMV infections</td>
</tr>
<tr>
<td>Famiclovir</td>
<td>FCV Famvir</td>
<td>HSV-1, HSV-2, VZV, EBV and HBV</td>
<td>Acyclic guanine nucleoside (oral prodrug of penciclovir); in phase III clinical trials for HSV infections</td>
</tr>
<tr>
<td>Foscarnet</td>
<td>PFA Foscavir</td>
<td>HSV-1, HSV-2, CMV, VZV, EBV, HHV-6, HHV and HBV</td>
<td>Organic analogue of inorganic pyrophosphate, used primarily in the treatment of CMV disease</td>
</tr>
<tr>
<td>Ganciclovir</td>
<td>GCV Cymevene/Cytovene</td>
<td>HSV-1, HSV-2, CMV, HHV-6, VZV, EBV and HHV</td>
<td>Acyclic purine nucleoside used in the treatment and prophylaxis of CMV disease including CMV retinitis</td>
</tr>
<tr>
<td>Idoxuridine</td>
<td>5-IdUdr Herpid, Stoxil, Iduridin Virudox, Idoxene, Kercide</td>
<td>HSV-1, HSV-2 and VZV</td>
<td>Iodinated analogue of thymidine, used in the topical treatment of keratoconjunctivitis caused by HSV and cutaneous herpes zoster</td>
</tr>
<tr>
<td>Ribavirin</td>
<td>ICN-1229 Virazid, Virazide, Virazole, Vilona</td>
<td>RSV, MV, HAV, HBV, HCV, influenza A and B, Lassa viruses, Hantaan and Junin virus</td>
<td>Nucleoside analogue used primarily in the treatment of RSV in infants; useful wide spectrum antiviral in life threatening situations</td>
</tr>
<tr>
<td>Valaciclovir</td>
<td>VACV Valtrex</td>
<td>HSV-1, HSV-2, VZV, EBV and CMV</td>
<td>Acyclic guanine nucleoside (oral prodrug of acyclovir)</td>
</tr>
<tr>
<td>Zanamir</td>
<td>Relenza</td>
<td>influenza A and B viruses</td>
<td>Sialic acid analogue, neuraminidase inhibitor</td>
</tr>
</tbody>
</table>
inhibitors of HIV and can have a have a significa-
cant impact on HIV replication for several
months when used as a monotherapy. How-
ever, with time treatment failure still occurs in
most patients, again caused by the selection of
mutant strains of HIV and the subsequent
development of resistance.

**CHANGING THE PARADIGM**

In 1995 two reports were published which
reversed what was once the conventional view
that HIV was present in small quantities in
infected individuals. This change in our
understanding of the kinetics of HIV replica-
cation followed exploration of the potent anti-
iral effect of the protease inhibitors, and the
new PCR technology which enabled very sens-
itive changes in HIV replication to be
measured as the viral load (HIV RNA copy
number/ml of blood). Studies measuring the
drop in viral load in patients treated with
ritonavir and indinavir monotherapies showed
that $10^9$–$10^{10}$ virus particles are produced and
cleared daily in peripheral blood. Further, at
the end of the acute phase of infection each
individual achieved a steady viral load concen-
tration which ranged from $10^5$–$10^6$ copies/ml
of blood. This was in contrast to an earlier
model of HIV infection, which proposed a
relatively low steady state of HIV replication,
based on less sensitive measurements of viral
concentration.

The original assumption made about anti-
HIV treatment was that drugs could inhibit the
production of new virus but could not quickly
delete virus already present in the blood
stream. Treatment of infected individuals with
protease inhibitors showed very dramatically
that every two days the level of plasma HIV was
reduced by half and that within 14 days viral
load could be reduced by 2–3 logs. The
conclusion drawn from these studies was that
chronically infected cells, which are a small
proportion of the total number of peripheral
blood mononuclear cells, are not capable of
replenishing daily plasma HIV levels. For plasma
HIV levels to reach those observed in
untreated individuals many new cells of the
immune system need to be infected constantly.
It was estimated that about 5% of the daily
production of CD4+ cells, the primary host
cell for HIV replication, are killed by HIV.
Further studies showed that a turnover of
10–100 million CD4+ cells per day was usual
in infected people not undergoing chemo-
therapy. Studies on CD4+ kinetics with the
protease inhibitors showed that the rise in
CD4+ counts doubled every 15 days and cor-
related with a fall in viral load. Further, the
data showed that even in severely immuno-
compromised people a significant increase in
CD4+ count occurred.

More recently it has been proposed that the
large increase in CD4+ found in peripheral
blood following effective antiretroviral treat-
ment is not primarily the result of T cell prolif-
eration but is caused by a redistribution of the
cells already present. During active viral
replication masses of T and B cells are trapped
in the lymph nodes by the immune response
and can remain in there for a long time.
Following chemotherapy, it is the release of
these cells into the blood that causes a rise in T
cells rather than a high level of proliferation.
These studies on HIV dynamics have indicated
that that lymph nodes of the secondary
lymphoid system are the major reservoir
of HIV and not the circulating PBMC. It has been
proposed that long memory T cells constitute a
major compartment in which HIV survives. It
is on the latently infected long lived compart-
ments within the body that antiretroviral treat-
ment must have its impact if HIV is to be
eliminated.

This new understanding of HIV kinetics has
several consequences for HIV treatment. Virus
replication, as measured by the HIV RNA lev-
els, is clearly related to clinical progression.
Thus driving down the viral load as far as pos-
sible has become a goal of antiretroviral
treatment. Further, it is considered that the
rapid development of drug resistance results
from the very high replication rate of HIV and
not from an abnormally high mutation rate.
Clinical trial data show that long term suppres-
sion of HIV replication can only be achieved by
the use of several drugs in combination, and
this approach is now used as first line
treatment. However, the efficacy of the combi-
nation is dependent on the cross resistance
profiles of each drug and not necessarily on the
potency of the drug. The use of one PI in com-
bination with two NRTI is now considered to
be one of the most potent anti-HIV regimens to
date (HAART, highly active antiretroviral
treatment). Consensus opinion is that the most
potent combination antiretroviral treatment
should begin as soon as possible and certainly
once the virus load begins to increase.

If treatment can reduce HIV replication to
ever very low levels there is hope that the compart-
ments harbouring infected cells may be
cleared by the immune system itself. Even
though some patients achieve large increases
in CD4+ counts with antiretroviral treatment,
their ability to mount an immune challenge is
still suboptimal. A further approach to HIV
chemotherapy now receiving much attention is
to combine antiretroviral treatment with an
immunomodulator. The most studied treat-
ment is with interleukin 2 (IL-2). Individuals
treated with IL-2 have sustained a significant
improvement in their CD4+ counts compared
with people who have received antiretroviral
treatment alone. In some cases CD4+ counts
have risen by 400/µl. There are also data sug-
gest that individuals who start treatment
with a higher CD4+ count have the best
response. Thus, theoretically, immunomodu-
lators should be used sooner rather than later
in the infection. IL-2 itself is toxic and requires
infusion over a period of time. It also enhances
HIV replication, but this is controlled by con-
comitant antiretroviral treatment. Recently
several other biological response modifiers,
cytokines, and small molecular weight mol-
ecules have been investigated in HIV infec-
tions.
Drugs for herpesvirus infections
Acyclovir was discovered in the 1974 and was the first effective antiviral drug to be used extensively. It is a guanine analogue with a truncated sugar and is a member of the class of acyclic nucleotide inhibitors. This compound is widely used for the treatment of herpesvirus-1 (HSV-1) and HSV-2 infections. It is the safest of all the nucleoside analogues as it requires the virus to activate the compound. The first phosphorylation step is carried out by the virus enzyme, thymidine kinase, a process which is about 200 times faster than the host cell analogue. The pharmacological result of this selectivity is that ACV is essentially absent in uninfected cells. The second and third phosphorylation steps are brought about by cellular enzymes. ACV-triphosphate competes with the normal substrate, deoxyguanosine triphosphate, for the herpesvirus DNA polymerase, and incorporation of ACV-triphosphate into the growing virus DNA causes chain termination. Advances have been made recently in synthesising valaciclovir (VCV) which is the prodrug of ACV. VCV has a half life of about 12 hours and is consequently given less often, which improves compliance. Famciclovir (FCV), a prodrug of penciclovir (PCV), has also been licensed for the treatment of varicella-zoster virus and genit-al herpes. It has a similar mode of action to acyclovir, but PCV-triphosphate has a half life about 20 times longer than ACV-triphosphate, although being a significantly weaker inhibitor of herpesvirus DNA polymerase.

The development of resistance to ACV is not an important issue in the treatment of herpesvirus infections in non-immunocompromised hosts because acyclovir resistant strains are less fit and do not outgrow the wild type. In immunocompromised patients, however, they may be fit and do not outgrow the wild type. In immunocompromised patients is cytomegalovirus (CMV), and another nucleoside analogue, ganciclovir (GCV), was developed to treat CMV reactivation in HIV infected patients and those undergoing transplantation. The viral target is the CMV DNA polymerase. Biochemical studies have shown that GCV, like ACV, is phosphorylated by a viral protein, in this case the UL97 viral gene product. The amino acid sequence of this UL97 indicates that it is primarily a protein kinase, but it also possesses the ability to phosphorylate nucleosides. More recently HPMPC (cidofovir), a nucleoside monophosphate, has been developed to treat CMV retinitis in AIDS patients. Again the target is the CMV DNA polymerase. The drug causes severe renal toxicity, but because it is stable for many days it can be delivered directly to the eye intermittently, thus avoiding systemic overexposure. Molecular cloning of the herpesviruses has also revealed a conserved sequence coding for a protease which is an essential gene function for their replication. Although this enzyme is well characterised and high throughput assays systems have been developed, there has been no major breakthrough in the discovery of potent inhibitors to date.

Influenza infections
Many other viral infections cause significant human disease, but until recently antiviral intervention has not achieved much success. Influenza epidemics are common and account for many fatalities, particularly in infants and elderly people. Amantadine and rimatadine are compounds which were found to block an ion channel established by influenza A in order to allow the virion to fuse with the cell membrane and enter the cell. However, drug resistance occurs within a few rounds of replication, which proved to be a serious limitation to their use. The most recent advance in this field is the development of influenza neuraminidase inhibitors. Neuraminidase is a glycoprotein found in the viral envelope, and involved in the cell to cell spread of the virus. Neuraminidase is active at a late stage in influenza replication and removes cellular receptors which bind the other major influenza surface enzyme glycoprotein, haemagglutinin, a process essential for virus–cell fusion. Thus interference with neuraminidase activity inhibits the escape of the influenza virus from the infected cell. Crystallographic studies have shown that the active site of neuraminidase is highly conserved in all strains of influenza A and B, thus supporting its role as a target for chemotherapy. A number of potent inhibitors of both influenza A and B neuraminidase have been synthesised: zanamivir (4-guanidino-2,4-dideoxy-2,3-dehydro-N-acetylneuraminic acid) and GS4104, which is a neuraminidase inhibitor prodrug, are the first to have significant activity in vivo. Zanamivir is given by aerosol as it is cleared rapidly from the plasma when given systematically. Recent trials have shown that, when taken prophylactically or within 26 hours after infection, zanamivir is effective in reducing symptoms. Resistance does occur but the neuraminidase of the viral mutants is unstable and such viruses do not replicate as well as the wild type.

Hepatitis virus infections
Chronic viral hepatitis is known to infect several hundred million people worldwide and causes severe liver disease. The association of hepatitis B and C viruses with blood transfusion, intravenous drug use, and sexual exposure has encouraged the pharmaceutical industry to expand their drug discovery programmes in this area.

The acute disease seen in hepatitis B virus (HBV) infection ranges in severity from asymptomatic to fatal fulminant hepatitis. The chronic condition also varies from benign conditions to chronic active hepatitis and liver cancer. The World Health Organisation estimates that of the 350 million carriers of HBV, 65 million will die of chronic liver disease. Studies have determined that HBV has a virally encoded reverse transcriptase which transcribes viral RNA into DNA, within the core particles, during the late stages of the replication cycle. Thus many of the reverse transcriptase inhibitors which showed activity against HIV were evaluated against HBV.

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Unfortunately most of the compounds which were active in vitro were found to be either inactive or too toxic in vivo. However, a few drugs are under development. The antiviral prodrug, famciclovir, is in phase II clinical studies and suppresses the virus load in chronic HBV infection. Originally it was thought that famciclovir required activation by the herpes specific thymidine kinase but it is now clear that cellular kinases can carry out the first phosphorylation step. Treatment with lamivudine, which is in phase III development, resulted in significant histological improvement in most patients as well as a statistically significant reduction in HBV e antigen (HBeAg) seroconversion. These compounds do not clear the infection and a “rebound effect” is observed in HBV replication when treatment is withdrawn. Both famciclovir and lamivudine induce mutations in the HBV RNA polymerase, and although the HBV mutation rate is similar to that of the retroviral pol gene the emergence of resistance takes much longer than for HIV. The virulence and replication capacity of these drug resistance strains has yet to be assessed in relation to the pathogenesis of HBV.

The nucleoside analogue, BMS-200475, a cyclopentyl guanosine analogue, is in phase II/IIa development. In the WHV (woodchuck hepatitis virus) model there are data indicating that this compound may be one of the most potent HBV inhibitors under development; phase II studies indicate that total daily doses as low as 5 mg may be effective. The acyclic nucleoside analogues adefovir dipivoxil and lobucavir are also in phase II development; both induce approximately a 2 log drop in HBV DNA over several months of treatment.

Hepatitis C virus (HCV) was first identified in 1989 as the non-A non-B transfusion associated virus by a combination of cDNA cloning and expression techniques. Subsequent DNA sequence analysis showed it to be a member of the flavivirus group. Worldwide prevalence is estimated to be 300 million chronically infected carriers. The severe consequences of long term infection and the limitations of post-translational processing of the HCV NS3 protein. This viral enzyme is needed for post-translational processing of the non-structural region of the HCV polyprotein. Even though high throughput screens—using cell-free systems—are in place, no potent inhibitors have been reported. However, modest antiviral activity against the isolated protease has been achieved with substrate based peptide mimetics. Polyprotein inhibitors selected from bacteriophage libraries have also shown some activity.

The other key target is the HCV helicase which is located at the C terminal end of the NS3 protein. This is essential for RNA replication and presents various potential sites for small molecule inhibitor binding. These include the binding sites for ATP, the single stranded polynucleotide, and the double stranded polynucleotide, all of which may exist in multiple conformations. A molecule which traps the enzyme in either an open or a closed conformation would also block its activity. Treatment for HCV infections is with interferon alfa, but only about one third of patients respond and the symptoms return following cessation of treatment. Ribavirin has been used for the treatment of HCV and although there seems to be no effect on the level of circulating virus there may be a reduction in the markers of active hepatitis.

I thank Dr John Williamson for his comments on this review.

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