Abnormal regulation of the oestrogen receptor in benign breast lesions

B S Shoker, C Jarvis, R B Clarke, E Anderson, C Munro, M P A Davies, D R Sibson, J P Sloane

Abstract

Background—In normal breast tissue the oestrogen receptor (ER) and the proliferation associated antigen Ki67 are negatively associated, indicating that ER+ cells are non-dividing, or that the receptor is downregulated as cells enter cycle. This relation is completely or partially lost in many ER+ breast cancers and in in situ proliferations associated with an increased cancer risk, where coexpression of the two markers is often found.

Aims—To determine whether similar changes can be identified in other risk associated breast lesions.

Patients/Methods—Paraffin wax blocks from 12 cases of lactational change, 21 apocrine metaplasias, 22 duct ectasias, 20 sclerosing adenosis, 20 fibroadenomas, 19 phyllodes tumours, 20 radial scars, 21 papillomas (15 solitary and six multiple), 15 gynaecomastias, and nine postmortem male breast tissues were retrieved. Immunohistochemistry was used to determine the expression of ER and dual labelling immunofluorescence was used to detect cells expressing both ER and Ki67.

Results—Increased numbers of ER+ cells were seen in sclerosing adenosis, radial scars, papillomas, fibroadenomas, and phyllodes tumours but not in apocrine cysts (where no ER+ cells were detected) or duct ectasia (where normal numbers were found). As in the normal breast, the proportion of ER+ cells increased with age in all lesions with the exception of fibroadenomas. Coexpression of ER and Ki67 was found in an increased proportion of cells of all risk associated lesions studied. ER+ cells were less likely to be dividing than ER− cells in all cases, although this was significant only for sclerosing adenosis. The data on sclerosing adenosis, radial scars, papillomas, and fibroadenomas are comparable with those reported previously in hyperplasia of usual type, whereas those in duct ectasia are similar to those of the normal breast. The findings in all lesions, however, differed from those in ductal carcinoma in situ, where proportions of ER+ and ER+/Ki67+ cells are higher and the relation between ER+ cell numbers and age is lost. Thus, the nature and degree of dysregulation of ER in benign breast lesions is broadly in accordance with the degree of risk of developing breast cancer with which they are associated. In gynaecomastia, the proportions of ER+ and ER+/Ki67+ cells were comparable with those seen in benign female breast lesions, but changes with age were not observed. However, the changes in gynaecomastia were similar to those seen in normal male breast.

Conclusion—These findings are in keeping with the contention that the dissociation of ER and Ki67 expression is a very early change in the pathway to many breast cancers. However, this change might only have preneoplastic importance in the hormonal milieu of the female breast.

Keywords: oestrogen receptor; proliferation; benign breast; precancerous breast

Oestrogen is thought to be important in the pathogenesis of breast cancer, probably through the stimulation of epithelial proliferation via the oestrogen receptor (ER).1 It is associated with most of the epidemiological risk factors such as the ages at menarche, first child, and menopause, and the use of oral contraception or hormone replacement therapy.2 In the normal breast, ER+ cells are in the minority, but the proportion increases somewhat with age. There is a negative association between the expression of ER and the proliferation marker Ki67, indicating that either ER+ cells are non-dividing or that the receptor is downregulated as cells enter cycle. This important relation breaks down in many ER+ cancers, where the receptor is often detected on proliferating cells.3 We have recently studied ER expression in the in situ proliferations hyperplasia of usual type (HUT), atypical ductal hyperplasia (ADH), ductal carcinoma in situ (DCIS), and lobular in situ neoplasia (LIN) because there is strong evidence that they are precursors of breast cancer,4,5 and because they form a histological spectrum in which the molecular changes in the development of cancer can be studied. We showed that the proportion of ER+ cells is increased above normal postmenopausal values in all of these proliferations and that in all except HUT the normal increase with age is lost.6 Furthermore, the proportion of ER+/Ki67+ cells was increased in all lesions and the negative association between ER and Ki67 was lost in all except HUT.7 Thus, increases in ER+/Ki67+ cells and the breakdown of normal age regulatory mechanisms of ER+ cell numbers might represent early precancerous changes.

However, little is known about these relations in other benign breast lesions, some of
which have also been shown to be associated with an increased risk of developing breast cancer. Several studies have shown that there is an increased relative risk of developing breast cancer associated with sclerosing adenosis,\(^\text{10}\) intraduct papilloma,\(^\text{7,11}\) radial scar,\(^\text{12-14}\) and even fibroadenoma.\(^\text{6,15,16}\) Other changes such as apocrine cysts, duct ectasia, and gynaecomastia are not thought to be associated with an increased risk of cancer. The purpose of our study was to determine whether any of these lesions contained excessive numbers of ER+ cells or showed a breakdown in the normal relations between ER expression, age, and cell proliferation and, if so, whether the presence and degree of the abnormalities related to cancer risk.

Materials and methods

PATIENTS

Paraffin wax blocks and slides of 165 cases spanning a 10 year period were retrieved from the files of the department of pathology at the Royal Liverpool University. They comprised 12 showing lactational change, 21 apocrine metaplasias, 22 duct ectasias, 20 sclerosing adenosis, 20 fibroadenomas, 19 phylloides tumours, 20 radial scars, 21 papillomas (15 solitary and six multiple), 15 gynaecomastias, and nine postmortem male breast tissues. Two pathologists (BSS, JPS) made all the diagnoses following the pathology guidelines of the NHS breast screening programme.\(^\text{17}\) Some of the blocks studied contained more than one lesion. Immunohistochemistry and dual labelling immunofluorescence were performed.

IMMUNOHISTOCHEMISTRY

All the tissue samples were fixed in 10% formalin followed by four hours fixation in methacarn and then routinely processed to para wax. The ER was detected with a mouse monoclonal anti-ER antibody (clone 1D5; Dako Ltd, Ely, Cambridge, UK). Immunohistochemistry was performed using a standard streptavidin–biotin method with previous microwave antigen retrieval. Negative controls, in which the primary antibody was omitted, and three positive controls of ER+ breast carcinoma of varying staining intensities were included in each batch of immunohistochemistry. The method was identical to that used for the routine assessment of ER status in which our laboratory performs well in the UK external quality assessment scheme. To maximise the consistency of scoring, only nuclei showing moderate or strong staining were regarded as positive. The percentage of ER+ cells for each lesion was calculated.

DUAL LABELLING IMMUNOFLUORESCENCE

The method used was similar to that described previously.\(^\text{7}\) Briefly, proliferation and ER receptor expression were visualised using antibodies raised against the Ki67 antigen, which is expressed in the late G1, S, G2, and M phases of the cell cycle (Ki67p; Novocastra, Newcastle upon Tyne, UK), and against the human ER (clone 1D5). Dual fluorescent label immunohistochemistry was carried out after microwave antigen retrieval, and primary antibody binding was detected with a fluorescein labelled (green) secondary antibody in the case of Ki67 and a Cy3 labelled (red) antibody for ER. Cell nuclei were then counterstained with 4',6-diamidino-2-phenylindole, which fluoresces blue. Control slides were included in each analysis by performing the same procedures and substituting non-immune serum for primary antibodies and secondary antibodies individually.

Quantification of the fluorochromes was performed by either scoring the entire lesion or between 1000 and 4000 cells across several representative fields (chosen using a 4',6-diamidino-2-phenylindole filter), depending on the size of the lesion. Each field was examined under a high powered lens for the red (Cy3), green (fluorescein), and blue (4',6-diamidino-2-phenylindole) fluorochromes using the appropriate filters in succession to assess the presence or absence of double labelled cells. A triple band filter in which all three fluorochromes could be seen simultaneously was used for confirmation of dual staining. In addition, because many high powered fields contained few if any proliferating cells, the entire number or a minimum of 100 Ki67+ cells were assessed for coexpression of ER to give a better overall indication of dual positivity within a lesion.

EVALUATION OF DATA

The data were analysed by the Pearson product moment correlation coefficient and the non-parametric Mann Whitney and Kruskal Wallis tests using SPSS software for Windows (release 6.1).

Results

ER EXPRESSION AND ITS RELATION TO PATIENT AGE

All the female breast lesions studied, except cysts showing apocrine metaplasia, contained ER+ epithelial cells. In most of these lesions the percentage of ER+ cells correlated positively with the patient’s age; however, the results were significant only for papillomas and phylloides tumours, with fibroadenomas showing no age correlation (table 1). Lactating breast and the breast lesions associated with no increased risk of developing carcinoma—duct ectasia and apocrine metaplasia—contained few or no ER+ cells, even when they occurred in postmenopausal women. In comparison with the breast lesions associated with no increased risk, the breast lesions associated with a low risk of developing cancer (fibroadenoma, sclerosing adenosis, papillomas, and radial scars) contained increased numbers of ER+ cells, and this was significant (Mann Whitney, highest \(p = 0.001\) for fibroadenomas). The low risk breast lesions also contained a higher mean percentage of ER+ cells than age matched (most cases age matched to within 2 years) normal breast tissue (Mann Whitney, highest \(p = 0.04\) for sclerosing adenosis). However, there was pronounced heterogeneity in ER staining within similar lesions. The range of ER+ cells in all the lesions

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was wide (mean range, 68%) and for fibroadenomas it was between 1% and 85%, whereas for papillomas the range was 1–95%.

### ER AND Ki67 DUAL EXPRESSION

Ki67+ cells were also present within all the lesions studied, ranging from 1.5% of cells in sclerosing adenosis to 4.4% in fibroadenomas (table 2). There was less variability in the expression of Ki67 than ER within similar and between different lesions, although for fibroadenomas it did vary between 1.6% and 12%. However, only papillomas had a significantly higher mean percentage of Ki67+ cells than age matched normal breast tissue (Mann Whitney, *p* = 0.001). The proportion of dual positive cells is given as the percentage of all cells counted (table 2, column 7) and as the percentage of Ki67+ that were also ER+ (table 2, column 10). The former gives an indication of the overall proportion of dual labelled cells but is dependent on the total numbers of ER+ and Ki67+ cells. The latter is not dependent on overall cell numbers and consequently is more likely to reveal an abnormal relation between the two markers. Cells dually expressing ER and Ki67 were not seen in apocrine metaplasia (fig 1A) or duct ectasia (fig 1B). In contrast, the lesions associated with a low cancer risk contained significantly greater numbers of dual expressing cells (fig 1C), and in all except sclerosing adenosis the percentage of ER+/Ki67+ cells was significantly increased compared with normal premenopausal and postmenopausal lobules (table 2), and in comparison with age matched normal breast tissue (Mann Whitney, highest *p* = 0.05 for radial scars).

### OBSERVED/EXPECTED RATIOS

Because the lesions have different numbers of ER+ and Ki67+ cells, the percentages of double labelled cells that would be expected if the two variables were independent have been calculated. This was calculated for each lesion by multiplying the percentage of ER+ and Ki67+ cells and then dividing by 100. The observed values were lower than the expected values for all the lesions, although this was significant only in sclerosing adenosis (*p* = 0.0028) (table 2). Calculations for duct ectasia and apocrine metaplasia were not possible because of the absence of dual labelled cells.

The observed/expected (O/E) ratio gives an indication of whether the two markers are positively or negatively associated with each other and the strength of the association. In the former, values of >1 would be expected and in the latter, <1. Figure 2 is a box plot of the O/E values. The lesions associated with no increased risk of breast carcinoma have median values of 0, whereas all the low risk lesions have intermediate values between 0 and 1. All cases

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**Table 1** Relation between proportion of the oestrogen receptor (ER) positive cells and age in breast lesions associated with no or a low increased risk of subsequently developing breast cancer

<table>
<thead>
<tr>
<th>Age</th>
<th>Breast lesions</th>
<th>Mean % of ER+ cells SD</th>
<th>No. cases</th>
<th>Mean % of ER+ cells SD</th>
<th>No. cases</th>
<th>Mean % of ER+ cells SD</th>
<th>No. cases</th>
<th>Pearson correlation coefficient</th>
<th>p Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>&lt;46</td>
<td><em>Normal female</em></td>
<td>12 (11) 7</td>
<td>31 (27) 20</td>
<td>53 (34) 22</td>
<td>0.2803</td>
<td>0.006</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td><strong>Lactating breast</strong></td>
<td>12 (3) 5</td>
<td>11 (15) 9</td>
<td>11 (15) 9</td>
<td>0.4103</td>
<td>0.058</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Duct ectasia</td>
<td>4 (5) 12</td>
<td>8 (35) 28</td>
<td>8 (72) 16</td>
<td>0.4003</td>
<td>0.080</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Radial Scar</td>
<td>2 (15) 0</td>
<td>11 (45) 24</td>
<td>7 (47) 21</td>
<td>0.3644</td>
<td>0.114</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Papilloma (all)</td>
<td>6 (17) 5</td>
<td>5 (31) 18</td>
<td>10 (60) 25</td>
<td>0.5692</td>
<td>0.007</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Papilloma (solitary)</td>
<td>4 (12) 10</td>
<td>4 (35) 18</td>
<td>7 (57) 28</td>
<td>0.5642</td>
<td>0.028</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Papilloma (multiple)</td>
<td>2 (27) 25</td>
<td>1 (15) –</td>
<td>3 (67) 18</td>
<td>0.6435</td>
<td>0.168</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Fibroadenoma</td>
<td>12 (37) 12</td>
<td>3 (15) 9</td>
<td>5 (27) 27</td>
<td>–0.1064</td>
<td>0.655</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Phyllodes tumour</td>
<td>4 (40) 26</td>
<td>1 (50) –</td>
<td>10 (73) 19</td>
<td>0.5438</td>
<td>0.851</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Male (all)</td>
<td>10 (61) 22</td>
<td>3 (78) 13</td>
<td>11 (56) 24</td>
<td>–0.0405</td>
<td>0.851</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Normal male</td>
<td>0 (–) –</td>
<td>2 (85) 7</td>
<td>7 (55) 27</td>
<td>–0.1852</td>
<td>0.633</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Gynaecomastia</td>
<td>10 (61) 22</td>
<td>1 (65) –</td>
<td>4 (57) 23</td>
<td>–0.0457</td>
<td>0.871</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

*Data from Shoker et al.*

**Table 2** Coexpression of the oestrogen receptor (ER) and a proliferation marker (Ki67) in breast lesions associated with no or a low increased risk of subsequently developing breast cancer

<table>
<thead>
<tr>
<th>Lesions</th>
<th>Mean age (SD)</th>
<th>No. of cases</th>
<th>Mean no. of cells counted (SD)</th>
<th>Mean % ER+ positive cells (SD)</th>
<th>Mean % Ki67+ positive cells (SD)</th>
<th>Mean % dual positive cells (SD)</th>
<th>Mean % dual positive cells expected (SD)</th>
<th>Mean observed/expected ratio</th>
<th>Mean % of total Ki67+ that are ER+ (SD)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Normal (premenopausal)</em></td>
<td>42 (10)</td>
<td>14</td>
<td>1475 (885)</td>
<td>6.8 (5.4)</td>
<td>2.6 (3.1)</td>
<td>0.01 (0.02)</td>
<td>0.17 (0.37)</td>
<td>0.05 (0.14)</td>
<td>0.44 (1.44)</td>
</tr>
<tr>
<td><em>Normal (postmenopausal)</em></td>
<td>65 (7.5)</td>
<td>10</td>
<td>1099 (328)</td>
<td>42 (23)</td>
<td>0.34 (0.33)</td>
<td>0.04 (0.05)</td>
<td>0.11 (0.12)</td>
<td>0.39 (0.52)</td>
<td>11 (20)</td>
</tr>
<tr>
<td>Lactation</td>
<td>29 (5.8)</td>
<td>10</td>
<td>1623 (353)</td>
<td>1.33 (1.15)</td>
<td>2.5 (2.6)</td>
<td>0.01 (0.03)</td>
<td>0.03 (0.03)</td>
<td>0.02 (0.52)</td>
<td>1.03 (1.79)</td>
</tr>
<tr>
<td>Apocrine metaplasia</td>
<td>57 (8.9)</td>
<td>13</td>
<td>954 (829)</td>
<td>0.0 (0.0)</td>
<td>1.10 (1.06)</td>
<td>0.0 (0.0)</td>
<td>0.0 (0.0)</td>
<td>0.0 (0.0)</td>
<td>0.0 (0.0)</td>
</tr>
<tr>
<td>Duct ectasia</td>
<td>51 (15)</td>
<td>10</td>
<td>915 (261)</td>
<td>6.5 (9.0)</td>
<td>1.57 (1.32)</td>
<td>0.0 (0.0)</td>
<td>0.12 (0.27)</td>
<td>0.0 (0.0)</td>
<td>0.0 (0.0)</td>
</tr>
<tr>
<td>Sclerosing adenosis</td>
<td>54 (12)</td>
<td>13</td>
<td>1353 (858)</td>
<td>0.49 (24)</td>
<td>1.01 (1.55)</td>
<td>0.18 (0.52)</td>
<td>0.37 (0.56)</td>
<td>0.22 (0.31)</td>
<td>12 (21)</td>
</tr>
<tr>
<td>Radial scar</td>
<td>56 (6.5)</td>
<td>13</td>
<td>2417 (946)</td>
<td>0.29 (18)</td>
<td>1.57 (1.14)</td>
<td>0.29 (0.34)</td>
<td>0.63 (0.50)</td>
<td>0.52 (0.27)</td>
<td>18 (15)</td>
</tr>
<tr>
<td>Papilloma</td>
<td>53 (12)</td>
<td>11</td>
<td>1795 (437)</td>
<td>50 (18)</td>
<td>3.1 (2.5)</td>
<td>0.79 (0.83)</td>
<td>1.60 (1.57)</td>
<td>0.54 (0.32)</td>
<td>21 (16)</td>
</tr>
<tr>
<td>Fibroadenoma</td>
<td>37 (11)</td>
<td>10</td>
<td>1375 (202)</td>
<td>45 (23)</td>
<td>4.4 (3.7)</td>
<td>1.03 (1.25)</td>
<td>1.92 (1.71)</td>
<td>0.46 (0.33)</td>
<td>25 (22)</td>
</tr>
<tr>
<td>Phyllodes tumour</td>
<td>93 (14)</td>
<td>10</td>
<td>1245 (157)</td>
<td>68 (16)</td>
<td>4.1 (4.0)</td>
<td>1.50 (1.54)</td>
<td>2.50 (2.16)</td>
<td>0.44 (0.27)</td>
<td>36 (18)</td>
</tr>
<tr>
<td>Normal male</td>
<td>45 (9)</td>
<td>12</td>
<td>654 (36)</td>
<td>63 (29)</td>
<td>1.39 (0.51)</td>
<td>0.11 (0.18)</td>
<td>0.18 (0.24)</td>
<td>0.57 (0.56)</td>
<td>38 (32)</td>
</tr>
<tr>
<td>Gynaecomastia</td>
<td>33 (16)</td>
<td>11</td>
<td>1510 (272)</td>
<td>63 (23)</td>
<td>1.7 (1.5)</td>
<td>0.56 (0.74)</td>
<td>0.96 (0.87)</td>
<td>0.47 (0.32)</td>
<td>33 (24)</td>
</tr>
</tbody>
</table>

*Data from Shoker et al.*

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in which the observed and expected values were 0 (for example, all cases of apocrine metaplasia) were excluded from the ratio analyses.

**MALE BREAST TISSUE**
The percentage of ER+ cells in normal postmortem male breast tissue and gynaecomastia specimens was similar, although gynaecomastia specimens did have a significantly greater percentage of Ki67+ cells (p = 0.02) and a greater percentage of dual expressing cells (p = 0.06) (fig 1D). O/E ratios, however, were similar for both. Male breast tissue (normal and gynaecomastia) has similarities to the low risk lesions in the female—a relatively high percentage of ER+ and dual expressing cells with an intermediate median value for the O/E ratio (table 2).

**Discussion**
There is overwhelming evidence that the effects of oestrogen are important in the development of human breast cancer. We have shown previously that the percentage of ER+ cells is increased within in situ proliferations (HUT, ADH, DCIS, and LIN) of the breast and that the proportion of these cells in different lesions correlates with the risk attributed to them in prospective studies.8 Furthermore, a fundamental change appears to occur between the HUT and ADH stages. The proportion of ER+ cells in the former, as in the normal breast, increases with age, whereas in the latter, in common with ER+ DCIS, no age association is seen, the percentage of ER+ cells being high at all ages.8 This suggests that the regulation of the expression of ER or the control of ER+ cells numbers escapes the normal age related regulatory mechanisms at the ADH stage.

In our present study we have undertaken similar investigations of other breast lesions, some of which have been shown to be associated with an increased relative risk of developing breast cancer comparable with that associated with HUT.5 7 10–16 These low risk lesions had increased proportions of ER+ cells, whereas those not associated with increased...
risk did not. An age related increase in the proportion of ER+ cells was seen in most lesions, but it was less pronounced than that seen in normal breast tissue or in HUT. However, the age related variation was significant in papillomas and phyllodes tumours only, and was not demonstrable in fibroadenomas. All the benign breast lesions showed a pronounced variation for the markers assessed. This heterogeneity might be related to the relative lack of consensus in the literature that they are associated with an increased risk in comparison with HUT, where the consensus is strong. Perhaps only subsets of the lesions studied are really associated with an increased cancer risk.

It is likely that the increase in ER+ cells with age is associated with declining plasma oestrogen concentrations. Furthermore, it is conceivable that the major architectural disturbances associated with the lesions studied in our investigation and the pronounced differences in their stromal components could modify local oestrogen concentrations and consequently the number of ER+ cells. Indeed, differences in the expression of other receptors within fibroadenomas and phyllodes tumours have already been described. The difference between fibroadenomas and phyllodes tumours in our study is perplexing, but stromal differences might be important. It is also worth noting that phyllodes tumours have not been found to be associated with an increased risk of developing breast cancer although, to the authors' knowledge, they have not been studied extensively in this context. The fact that certain subsets of fibroadenomas might be associated with increased risk has been established relatively recently.

In normal premenopausal lobules there is a negative association between steroid receptor expression and cell proliferation, as judged by the lack of dual immunostaining for ER and the proliferation marker Ki67. However, this normal relation is lost in many invasive ER+ breast cancers, in which a variable proportion of ER+ cells is also Ki67+. A similar phenomenon is seen within in situ carcinomas and atypical hyperplasias, but in HUT the negative association is at least partly retained. Although the negative association between ER and Ki67 was lost in some of these lesions, coexpression was not seen with greater frequency than would be expected by chance if the two markers were expressed independently of each other. Although the presence of ER+/Ki67+ in the male breast is more difficult to explain because there is no good evidence that gynaecomastia is precancerous. However, it is not known whether the breast epithelial cells in this condition lack the genetic instability of proliferative lesions in women, or whether the molecular changes in the female breast lead to malignancy only in the appropriate hormonal milieu. Certainly, the sex hormonal differences between men and women mean that it is not possible to extrapolate the function of ER from one gender to the other, or its role in breast carcinogenesis.

We are especially grateful to the Cancer and Polio Research Fund, which provided a grant to BSS, and for the support and facilities provided by Clatterbridge Cancer Research Trust.

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