Hyperplastic polyps are common lesions and occur most frequently in the recto-sigmoid area.1 They are diagnosed on the basis of a characteristic serrated epithelial pattern, which results from micropapillary luminal infolding of columnar absorptive cells and mature goblet cells.2 Historically, their lack of dysplasia/carcinoma associated nuclear and architectural features has placed them in a benign and non-neoplastic category. However, more recent studies have demonstrated clonal relations between hyperplastic polyps and dysplastic epithelium in some mixed polyps, and a frequency of K-ras mutation similar to that of adenoma.2 In 1990, Longacre and Fenoglio-Preiser described mixed hyperplastic adenomatous polyps of the colorectum and coined the term “serrated adenoma”.3 The distinction between a serrated adenoma and a hyperplastic polyp is not always straightforward. Ajioka et al used the term “atypical hyperplastic polypl” for those polyps with some features of a serrated adenoma but where there was definite surface maturation.4 Other authors have been less precise in their use of the criteria of Longacre and Fenoglio-Preiser in differentiating hyperplastic polyps from serrated adenomas.5 Ban5 examined two colonic polyps that appeared to be “atypical” on light microscopy. Using the proliferative marker Ki-67 and CK20 as a marker of enterocyte maturity, he found that both polyps showed Ki-67 positive cells extending to the upper portion of some crypts and CK20 positive cells distributed in a scattered fashion. All of the “traditional” hyperplastic polyps showed Ki-67 confined to the lower half of the crypts and CK20 expression confined to the upper three quarters of the epithelium.

“More recent studies have demonstrated clonal relations between hyperplastic polyps and dysplastic epithelium in some mixed polyps, and a frequency of K-ras mutation similar to that of adenoma”

The aim of our study was to look at the expression of the same markers (Ki-67 and CK20) in a larger group of hyperplastic polyps (including a group with “atypical” features) to determine whether upper crypt Ki-67 staining and basal crypt CK20 staining correlated with these atypical features, as assessed by light microscopy.

METHODS
Fifty seven polyps of the hyperplastic type were retrieved from the files of the histopathology departments of Manchester Royal Infirmary and Stepping Hill Hospital, UK, and stained

Abbreviations: CK, cytokeratin; HNPCC, hereditary non-polyposis colonic carcinoma; MSI, microsatellite instability
with antibodies to Ki-67 and CK20, following detailed histological review of haematoxylin and eosin stained sections. For comparison, nine moderately dysplastic (non-serrated) adenomas and seven moderately differentiated colorectal carcinomas were stained with the same antibodies.

A standard immunohistochemical method was applied for the antibodies to Ki-67 (Dako, Ely, Cambridgeshire, UK; primary antibody, 1/100 dilution; 30 minutes' incubation at room temperature after heat mediated antigen retrieval) and CK20 (Dako; primary antibody, 1/100 dilution; 30 minutes' incubation at room temperature after heat mediated antigen retrieval). Detection was with a labelled streptavidin–biotin method (Dako).

The slides were assessed independently by two observers (CRH and RFTMcM) with a $\kappa$ value of 0.68. Consensus was reached after discussion. When assessing staining, Ki-67 positivity of more than very occasional cells within the upper half of crypts was considered abnormal and CK20 positivity in a scattered pattern seen within the lower crypt half was considered abnormal.

RESULTS

The 57 hyperplastic polyps (fig 1A), including 31 with atypical features (fig 1B), were from a total of 53 patients (28 men and 25 women), with an age range of 28–82 years (mean, 52.5), and were derived only from the left side of the colon. They had been fixed in 10% buffered formalin, routinely processed, and embedded in paraffin wax. Most of the polyps were received in a fragmented state. On histological measurement they ranged up to 9 mm in diameter but most were less than 5 mm. Coincidental conditions documented on the histology request cards included hereditary non-polyposis colorectal carcinoma (HNPCC) in three patients, previous colorectal carcinoma (one patient), synchronous but separate colorectal carcinoma (two patients), BRCA1 gene carrier (one patient), strong family history of colorectal carcinoma (one patient), synchronous hyperplastic polyps (five patients), and synchronous neoplastic poly (three patients). Non-steroidal anti-inflammatory drug use was not documented.

The polyps (31 in number) categorised as atypical on light microscopy showed increased mitotic figures, pseudostratification with gland crowding, hyperchromasia, and increased apoptosis (fig 1B). Occasional multinucleated cells were seen at crypt bases in five of the 57 cases (fig 1C). In all the polyps there was definite surface maturation and the atypical features were predominantly seen in the lower and middle crypt zones; therefore, they were not felt to represent serrated adenomas. Neither did the features represent a collision between adenomatous and hyperplastic areas.

Of the 31 hyperplastic polyps with atypical features determined by light microscopy, 11 showed the usual pattern of lower crypt Ki-67 staining (fig 2A,B) and upper crypt staining with anti-CK20 (fig 2C,D); two showed Ki-67 staining extending into the upper half of the crypts (fig 2E), but with normal surface staining with anti-CK20; 14 showed Ki-67 staining confined to the lower half of the crypts but scattered lower crypt staining for CK20 (fig 2F), whereas four showed both upper crypt Ki-67 staining and scattered CK20 lower crypt staining.

Of the 26 hyperplastic polyps that appeared normal by light microscopy there were 17 with usual staining for both Ki-67 and CK20, one with upper crypt Ki-67 staining but normal surface CK20 staining, seven with Ki-67 confined to the lower half of crypts but with scattered lower crypt CK20 staining, and one with both upper crypt Ki-67 staining and scattered CK20 lower crypt staining. Table 1 summarises the results. When Fisher's exact test was applied, there was a significant difference ($p = 0.036$) between typical and atypical hyperplastic polyps with CK20 immunoreactivity but no significant difference ($p = 0.19$) for Ki-67 immunostaining.

DISCUSSION

Colonic polyps containing simultaneous but separate areas of hyperplastic and adenomatous glands are known to exist, but in 1984 Urbanski et al reported a different type of polyp with
the combined morphological features of hyperplastic and adenomatous epithelium. This was designated a “mixed hyperplastic adenomatous polyp”. Descriptions of this phenomenon were scarce until Longacre and Fenoglio-Preiser’s landmark paper on the subject was published in 1990. In this, they commented on 110 mixed hyperplastic/adenomatous polyps (exhibiting the architectural but not the typical bland cytological features of a hyperplastic polyp) and proposed the replacement term “serrated adenoma”.

According to Longacre and Fenoglio-Preiser, serrated adenomas were distinguished by several features, namely: the presence of a serrated epithelial surface in addition to goblet cell immaturity, upper crypt mitoses, prominence of nucleoli, absence of a thickened collagen table, and a nuclear to cytoplasmic ratio lying between that of hyperplastic polyps and traditional adenomas.

“Our examination of 31 hyperplastic polyps with atypical features on light microscopy revealed a difference in CK20 staining compared with that seen in the group of traditional looking hyperplastic polyps.”

To overcome the difficulty of not always being able to distinguish serrated adenomas from hyperplastic polyps, Ajioka et al referred to a group of polyps as atypical hyperplastic polyps. These were histologically characterised by “branched serrated glands with loss of goblet cells and the

Figure 2 (A) Low power view of typical hyperplastic polyp showing lower crypt pattern of nuclear staining (Ki-67 immunostaining). (B) Higher power view demonstrating basal location (Ki-67 immunostaining). (C) Low power view of typical polyp showing upper crypt staining (cytokeratin 20 (CK20) immunostaining). (D) Higher power view showing upper crypt staining, with membranous and cytoplasmic staining (CK20 immunostaining). (E) Atypical hyperplastic polyp demonstrating upper crypt Ki-67 nuclear staining in serrated epithelium (Ki-67 immunostaining). (F) Lower crypt staining in atypical hyperplastic polyp (muscularis mucosae to right) with typical mosaic pattern (CK20 immunostaining).
Ki-67 and CK20 expression in colorectal hyperplastic polyyps

Table 1 Comparison of Ki-67 and cytokeratin 20 (CK20) staining in atypical and typical hyperplastic polyyps

<table>
<thead>
<tr>
<th>Staining pattern</th>
<th>Typical on light microscopy (n=26)</th>
<th>Atypical on light microscopy (n=31)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Usual pattern (Ki-67 positivity confined to lower crypt, CK positivity confined to upper crypt)</td>
<td>17</td>
<td>11</td>
</tr>
<tr>
<td>Upper crypt zone Ki-67 positivity, usual pattern CK20 positivity</td>
<td>1</td>
<td>2</td>
</tr>
<tr>
<td>Usual pattern Ki-67 positivity with basal CK20 positivity</td>
<td>7</td>
<td>14</td>
</tr>
<tr>
<td>Upper crypt zone Ki-67 positivity and basal CK20 positivity</td>
<td>1</td>
<td>4</td>
</tr>
</tbody>
</table>

p=0.036 for CK20 differences (Fisher’s exact test; p value for the same or stronger association); p=0.19 for Ki-67 differences (Fisher’s exact test; p value for the same or stronger association).

**Take home messages**

- The normal pattern of lower crypt Ki-67 and upper crypt cytokeratin 20 (CK20) staining was seen in 28 of the 57 hyperplastic polyyps and, in general, this corresponded with standard light microscopic appearances.
- Twenty one of the 57 polyyps showed lower crypt mosaic CK20 staining, which in general corresponded with basal abnormalities on light microscopy, although seven specimens had normal appearances.
- Two smaller subsets emerged, one showing upper crypt Ki-67 staining in the presence of normal CK20 expression (three cases) and another in which a combination of lower crypt CK20 and upper crypt Ki-67 was seen (five cases).
- This last pattern was similar to that of neoplastic polyyps and raises the possibility that a subgroup of hyperplastic polyyps exists that may be a variant with malignant potential.
- Further studies with markers of mismatch repair genes and Kras mutations are needed to help clarify this issue.

**REFERENCES**


**Authors’ affiliations**

A Davenport, G Bigley, R F T McMahon, Department of Histopathology, Manchester Royal Infirmary, Oxford Road, Manchester M13 9WL, UK
R J Hale, C R Hunt, Department of Histopathology, Stepping Hill Hospital, Poplar Grove, Stockport SK2 7JE, UK

**Authors’ affiliations**

A Davenport, G Bigley, R F T McMahon, Department of Histopathology, Manchester Royal Infirmary, Oxford Road, Manchester M13 9WL, UK
R J Hale, C R Hunt, Department of Histopathology, Stepping Hill Hospital, Poplar Grove, Stockport SK2 7JE, UK
Children with aplastic anaemia and parvovirus infection are linked

Children with severe aplastic anaemia should be tested for B19 parvovirus infection, according to a small study that has shown that the two are associated. The study of 30 children with severe aplastic anaemia and healthy matched controls detected active or recent infection with B19 parvovirus with specific viral DNA or antibody, or both, in six anaemic children.

Two of the six had had previous erythema infectiosum—an infection associated with parvovirus B19. The four others had asymptomatic B19 parvovirus infection. Two children died and four recovered with combination treatment with horse antilymphocyte globulin, cyclosporin, and intravenous immunoglobulin.

The children, 18 boys and 12 girls, median age 6.8 years (range 1–14 years) were admitted with severe aplastic anaemia between April 1995 and December 1996. Healthy controls were selected, matched for age, community, and time of presentation. Parvovirus B19 DNA was detected by nested PCR of lysed serum samples and antibody against parvovirus B19 by ELISA.

Human parvovirus B19 is already linked with many other conditions. The commonest and most serious complication is severe aplastic anaemia in sickle cell anaemia and hereditary sphaerocytosis. Asymptomatic B19 parvovirus infection has been recorded with childhood severe aplastic anaemia only in one previous case.

How the virus causes severe aplastic anaemia is not clear. It may act directly, targeting cell lines in the bone marrow involved in blood formation through its attachment to P substance or by immunological means through interference with regulation of the phagocytic system by interferon γ.

Arrows of Disease in Childhood 2002;87:436–437.
Expression of Ki-67 and cytokeratin 20 in hyperplastic polyps of the colorectum

A Davenport, R J Hale, C R Hunt, G Bigley and R F T McMahon

*J Clin Pathol* 2003 56: 200-204
doi: 10.1136/jcp.56.3.200

Updated information and services can be found at:
http://jcp.bmj.com/content/56/3/200

These include:

References
This article cites 15 articles, 6 of which you can access for free at:
http://jcp.bmj.com/content/56/3/200#BIBL

Email alerting service
Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

Topic Collections
Articles on similar topics can be found in the following collections

- Clinical diagnostic tests (805)
- Cancer: small intestine (32)
- Immunology (including allergy) (1664)

Notes

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/