Density of neoplastic lymphoid infiltrate, CD8+ T cells, and CD1a+ dendritic cells in mycosis fungoides

G Goteri, A Filosa, B Mannello, D Stramazzotti, S Rupoli, P Leoni, G Fabris

Background/Aims: CD8+ T cells and epidermal/dermal dendritic cells expressing CD1a are found among neoplastic CD4+ T cells in mycosis fungoides (MF) lesions. This study analysed the relation of CD8+ tumour infiltrating lymphocytes (TILs), CD1a+ epidermal Langerhan’s cells (LCs), and dermal dendritic cells (DDCs) to clinicopathological parameters in 46 MF cases.

Methods: Pretreatment diagnostic biopsy specimens of 46 MF cases were submitted to histological analysis and immunohistochemistry. Four histological grades were defined based on the density of the neoplastic infiltrate: grade 1 (mild superficial perivascular infiltrate), grade 2 (moderate superficial perivascular infiltrate with some tendency to confluence), grade 3 (pronounced superficial band-like infiltrate), and grade 4 (deep nodular infiltrate). Epidermotropism was scored as low, moderate, or high. Numbers of CD8+ T cells and of dermal and epidermal CD1a+ cells were scored as 1 (low), 2 (moderate), and 3 (high). Correlations between these parameters and clinical data (age, sex, clinical type of lesion, stage, response to treatment, and recurrence) were analysed by the χ² test.

Results: Numbers of TILs and DDCs were associated with subepidermal infiltrates, being lower in less dense infiltrates, whereas there was no association between epidermal CD1a+ cells and the analysed parameters. Complete remission in treated patients was related to subepidermal infiltrates but not to TILs, LCs, or DDCs.

Conclusions: These results support the notion that CD8+ cells and dermal CD1a+ cells are active against tumour cells. MF with low numbers of TILs could represent an early stage of the disease, before TILs are activated against tumour specific antigens.

Mycosis fungoides (MF) is the most common type of cutaneous T cell lymphoma, generally showing an indolent and prolonged clinical history with slow progression over the years from patches to more infiltrated plaques, tumour, and systemic involvement in later stages. Less frequently, it may exhibit a more aggressive behaviour from the beginning (“MF d’emblée”). MF is thought to arise from a background of chronic inflammation as a response to chronic antigenic stimulation, with a series of genetic mutations resulting in stepwise progression from eczematous patches, as in parapsoriasis, to plaques, tumours, and eventually haematoegenous dissemination. The molecular events underlying the different clinical courses have been reviewed by Hansen. The pathogenesis of MF is probably related to various factors of exogenous and/or endogenous origin.

"Mycosis fungoides is thought to arise from a background of chronic inflammation as a response to chronic antigenic stimulation"

Histologically, MF is composed of a dermal infiltrate of neoplastic T cells with cerebriform nuclei showing epidermotropism ranging from single lymphoid T cells linearly arranged at the epidermal basal layer to clusters of cells spreading over all layers and forming classic Pautrier’s abscesses. Neoplastic cells typically express the CD3+/CD4+ T helper phenotype; other T cell antigens, such as CD2, CD5, and CD7, can be hypopressed or lost in the advanced stages of the disease. Early MF, showing a mild T cell lymphoid infiltrate, is hard to differentiate histologically from superficial dermatosis; diagnosis can be supported by the demonstration of monoclonal rearrangement of T cell receptor (TCR) genes and/or T cell antigen loss. There is an ongoing debate on whether these difficult cases should be considered as early lymphoma capable of progression, frustrate lymphoma with no risk of evolution, or an altogether different entity unrelated to MF.

In addition to neoplastic CD4+ cells, reactive cells are also detected in typical MF lesions, such as T cells staining for CD8 (tumour infiltrating lymphocytes; TILs) and dendritic cells expressing CD1a antigen, both in the epidermis (Langerhan’s cells; LCs) and dermis (dermal dendritic cells; DDCs). The role of these cells in the pathogenesis and progression of MF is still unclear. A few studies have focused on the distribution of CD8+ and CD1a+ cells and their relation with clinical behavior in MF, with conflicting results.

The aim of our study was to find a correlation, if any, between the density of TILs, LCs, and DDCs and the density, distribution, and epidermotropism of the lymphoid neoplastic infiltrate, with special emphasis on the comparison between early and more advanced MF.

METHODS

We reviewed the records of 46 patients consecutively referred by dermatologists to the Institute of Anatomical Pathology for diagnosis and to the department of haematology (University of Ancona School of Medicine) for staging, treatment, and follow up from 1994 to 2001. Clinical data regarding age, sex, clinical type of lesion, stage, treatment, and follow up were obtained from the medical charts. Pretreatment diagnostic biopsy specimens were submitted to histological and immunophenotypical analysis. Skin samples had been received fresh and cut into two halves: one was fixed in formalin and

Abbreviations: CR, complete remission; DDC, dermal dendritic cell; LC, Langerhan’s cell; MF, mycosis fungoides; TCR, T cell receptor; TIL, tumour infiltrating lymphocyte
embedded in paraffin wax, and the other frozen in isopentane, cooled in liquid nitrogen, and stored at −80°C.

Haematoxylin and eosin stained sections from formalin fixed and paraffin wax embedded tissues were reviewed to evaluate the morphological features of the disease. The density of the neoplastic infiltrate was scored with a four point scale based on quantity and distribution as follows. Grade 1: mild, superficial, and perivascular subepidermid lymphoid infiltrate without tendency to confluence; grade 2: moderate, superficial, and perivascular infiltrate in the upper dermis showing a tendency to confluence; grade 3: band-like lymphoid infiltrate homogenously distributed below the dermal-epidermal junction; grade 4: heavy burden of neoplastic T cells, diffusely extending from the superficial to the reticular dermis or into the hypodermis.

Epidermotropism was scored with a three point scale as follows. Score 1: epidermis occasionally infiltrated by single haloed lymphoid cells at the basal layer or forming one Pautrier's abscess (defined as at least four atypical lymphocytes in a single epidermal vacuole); score 2: epidermis moderately infiltrated by single basal lymphoid cells or clusters forming less than three Pautrier's abscesses; score 3: epidermis widely infiltrated by neoplastic cells or showing more than three Pautrier's abscesses.

Immunohistochemical analysis was performed on both formalin fixed, paraffin wax embedded sections and frozen tissue with the antibodies listed in table 1. For immunohistochemical detection, we used the Dako Envision™ horseradish peroxidase kit with new fuchsin as chromogen for frozen sections and the Dako Envision™ alkaline phosphatase kit with anti-CD5 BL1a Monoclonal 1/25, Anti-CD1a 1/25, Anti-CD4 13B8.2 Monoclonal 1/25, Anti-CD8 8H10.5 Monoclonal 1/25, and Anti-CD1a 1/25. All antibodies were from Immunotech (Marseille, France).

Histological, immunophenotypical, and PCR results
Eleven patients showed a mild subepidermal lymphoid infiltrate made up of cerebriform atypical cells with a superficial and perivascular distribution, without tendency to confluence (fig 1A): these findings were subtle, and differential diagnosis of an early patch MF or with superficial dermatitis was supported by the demonstration of an aberrant T cell antigenic profile and/or a monoclonal TCR-γ gene rearrangement. A moderate lymphoid infiltrate in the upper dermis with a perivascular distribution showing tendency to confluence was seen in 15 cases; 16 cases showed a band-like lymphoid infiltrate homogenously distributed below the dermal–epidermal junction (fig 1D), and four cases presented a heavy burden of neoplastic cells extending deeply into the dermis or the hypodermis. Table 2 shows the distribution of all clinical and pathological parameters in the four lymphoid infiltrate grades. The subepidermal lymphoid infiltrate density was found to be associated with many of the clinical parameters. Patients with a grade 1–2 subepidermal lymphoid infiltrate were older than those with grade 3–4 (mean, 62 (SD, 10) and mean 54 (SD, 12), respectively; p = 0.0383) and showed patches in 24 of 26 cases, whereas eight of the 20 patients with grade 3–4 showed plaques or nodules (p = 0.008). Three of four of the patients with a grade 4 lymphoid infiltrate were stage ≥ IIA, whereas 37 of the 42 patients with less than grade 4 were in stage < IIA (p = 0.0015). None of the patients with a nodular pattern showed pronounced epidermotropism, whereas a variable degree of epidermotropism was seen in the other groups.

All MF cases showed neoplastic cells with the typical CD4+ phenotype, with loss of other T cell antigens in 45 cases: CD2 was lost in 44 cases, CD5 in 25, and CD7 in 39. On polymerase chain reaction analysis, 39 cases showed one or two monoclonal bands and seven showed a polyclonal pattern. The subepidermal lymphoid infiltrate density was significantly associated with the number of CD8+ T cells (p = 0.004; fig 1B–E); the density of CD8+ T cells was lower in cases with a grade 1 subepidermal lymphoid infiltrate than in those with grade > 1 (p = 0.001; fig 2).

The dermal (p = 0.03), but not the epidermal (p = 0.42), CD1a positivity score was significantly associated with the density of the lymphoid infiltrate (fig 1C–F); the density of the dermal CD1a+ dendritic cells was lower in grades 1–2 than in grades 3–4 (p = 0.0037; fig 3).

Among the studied parameters, only age, clinical type of lesion, stage of disease, and density of subepidermal infiltrate

### Table 1 Antibodies used in our study for the phenotypic analysis of T cells and the detection of dendritic cells

<table>
<thead>
<tr>
<th>Antibody</th>
<th>Clone</th>
<th>Type</th>
<th>Dilution</th>
</tr>
</thead>
<tbody>
<tr>
<td>Anti-CD3</td>
<td>UCHT-1</td>
<td>Polyclonal</td>
<td>1/50</td>
</tr>
<tr>
<td>Anti-CD4</td>
<td>12B8.2</td>
<td>Monoclonal</td>
<td>1/25</td>
</tr>
<tr>
<td>Anti-CD8</td>
<td>B9.2</td>
<td>Monoclonal</td>
<td>1/25</td>
</tr>
<tr>
<td>Anti-CD5</td>
<td>B11a</td>
<td>Monoclonal</td>
<td>1/25</td>
</tr>
<tr>
<td>Anti-CD2</td>
<td>6FI0.3</td>
<td>Monoclonal</td>
<td>1/25</td>
</tr>
<tr>
<td>Anti-CD7</td>
<td>8H10.5</td>
<td>Monoclonal</td>
<td>1/25</td>
</tr>
<tr>
<td>Anti-CD1a</td>
<td>C10</td>
<td>Monoclonal</td>
<td>Prediluted</td>
</tr>
</tbody>
</table>

All antibodies were from Immunotech (Marseille, France).
were significantly associated with response to treatment. Patients who achieved CR were older than non-responders (mean, 60 years (SD, 10) v 40 (8); p = 0.0014). Nineteen of 20 patients showing patches responded to treatment, whereas five of eight patients with plaques or nodules did not respond (p = 0.0008). Nineteen of 22 stage < IIA patients responded to treatment, whereas three of six patients at stage > IIA did not (p = 0.05). Moreover, CR was achieved more frequently in cases with subepidermal lymphoid infiltrate grades 1–2 than 3–4 (15 of 16 and seven of 12, respectively; p = 0.024). In contrast, CD8 (p = 0.43), epidermal CD1a (p = 0.42), and dermal CD1a (p = 0.12) positivity scores were not associated with a higher CR rate.

Clinical stage ≥ IIA (p = 0.006) and male sex (p = 0.0096) were found to be significantly associated with disease recurrence, but not age (p = 0.24), dermal infiltrate (p = 0.53), CD8 positivity (p = 0.41), epidermal CD1a positivity (p = 0.22), or dermal CD1a positivity (p = 0.59).

**DISCUSSION**

We found that patients with band-like and nodular subepidermal lymphoid infiltrates were younger, were more likely to have clinical plaques/nodules and be stage ≥ IIA, and were less likely to achieve CR than those with a less dense subepidermal lymphoid infiltrate. TIL and DDC numbers were found to be related only to the subepidermal infiltrate, being lower in less dense neoplastic infiltrates.

At an early stage, MF is very similar immunologically and morphologically to T cell cutaneous reactions; for this reason, it may be difficult to differentiate MF from aspecific superficial dermatosis. These similarities support the hypothesis that MF originates from a neoplastic transformation of T helper cells involved in a cutaneous immunological response. A model based on in vitro experiments has been proposed in which T helper CD4+ cells undergo retroviral infection as a result of the interaction with epidermal LCs, which take the retroviruses up from the infected keratinocytes nearby and transmit them to T cells through antigen presentation on major histocompatibility complex class II molecules. This would result in malignant transformation of the T cells, which become atypical and cerebriform. Inhibitory control is exerted by the cytotoxic CD8+ cells, the so called TILs: they are activated by the expression of tumour specific antigens on MF cells, which have the ability to escape the immune control of Fas ligand.

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**Table 2  Clinicopathological features in 46 patients with mycosis fungoides divided according to the subepidermal infiltrate grades**

<table>
<thead>
<tr>
<th>Subepidermal lymphoid infiltrate</th>
<th>Grade 1 (n=11)</th>
<th>Grade 2 (n=15)</th>
<th>Grade 3 (n=16)</th>
<th>Grade 4 (n=4)</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>M/F</strong></td>
<td>7/4</td>
<td>4/11</td>
<td>8/8</td>
<td>3/1</td>
</tr>
<tr>
<td><strong>Mean age (range)</strong></td>
<td>60.18 (45–73)</td>
<td>63.13 (45–76)</td>
<td>56.5 (37–72)</td>
<td>45.25 (28–61)</td>
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<tr>
<td><strong>Clinical patch</strong></td>
<td>11</td>
<td>13</td>
<td>12</td>
<td>0</td>
</tr>
<tr>
<td><strong>Clinical plaque/nodule</strong></td>
<td>0</td>
<td>2</td>
<td>4</td>
<td>4</td>
</tr>
<tr>
<td><strong>Stage</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt; IIA</td>
<td>10</td>
<td>13</td>
<td>14</td>
<td>1</td>
</tr>
<tr>
<td>≥ IIA</td>
<td>1</td>
<td>2</td>
<td>2</td>
<td>3</td>
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<tr>
<td><strong>Epidermotropism</strong></td>
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<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Grade 1</td>
<td>4</td>
<td>2</td>
<td>3</td>
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<td>Grade 2</td>
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<td>1</td>
</tr>
<tr>
<td>Grade 3</td>
<td>3</td>
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<td>5</td>
<td>0</td>
</tr>
<tr>
<td><strong>Loss of T cell antigens</strong></td>
<td>11/11</td>
<td>12/13</td>
<td>16/16</td>
<td>4/4</td>
</tr>
<tr>
<td><strong>T cell monoclonality</strong></td>
<td>8/11</td>
<td>14/15</td>
<td>13/16</td>
<td>4/4</td>
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<td><strong>Epididermal CD1a+ cells</strong></td>
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<td>Score 1</td>
<td>3</td>
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<td>Score 3</td>
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<td><strong>Treatment</strong></td>
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<tr>
<td>PUVA</td>
<td>1</td>
<td>2</td>
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<td>0</td>
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<tr>
<td>PUVA+IFNα</td>
<td>4</td>
<td>9</td>
<td>8</td>
<td>1</td>
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<tr>
<td>CHOP</td>
<td>0</td>
<td>0</td>
<td>1</td>
<td>1</td>
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<tr>
<td>RT</td>
<td>0</td>
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<td>0</td>
<td>1</td>
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<tr>
<td>None</td>
<td>6</td>
<td>4</td>
<td>7</td>
<td>1</td>
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<tr>
<td><strong>Response to treatment</strong></td>
<td></td>
<td></td>
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<td></td>
</tr>
<tr>
<td>CR</td>
<td>5</td>
<td>10</td>
<td>6</td>
<td>1</td>
</tr>
<tr>
<td>PR</td>
<td>0</td>
<td>1</td>
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<td>1</td>
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<tr>
<td>SD</td>
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<td>0</td>
<td>0</td>
<td>1</td>
</tr>
<tr>
<td><strong>Relapse</strong></td>
<td>2</td>
<td>2</td>
<td>3</td>
<td>0</td>
</tr>
<tr>
<td><strong>Status at last follow up</strong></td>
<td></td>
<td></td>
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<td></td>
</tr>
<tr>
<td>AW</td>
<td>5</td>
<td>10</td>
<td>4</td>
<td>0</td>
</tr>
<tr>
<td>AD</td>
<td>6</td>
<td>5</td>
<td>12</td>
<td>2</td>
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<tr>
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<tr>
<td>DNOD</td>
<td>0</td>
<td>0</td>
<td>0</td>
<td>1</td>
</tr>
<tr>
<td><strong>Mean (range) follow up time in months</strong></td>
<td>24.4 (2–79)</td>
<td>30.2 (7–70)</td>
<td>26.4 (2–84)</td>
<td>31.2 (7–76)</td>
</tr>
</tbody>
</table>

The follow up time was calculated from the date of diagnosis. Treatment: PUVA, psoralen + ultra violet A rays; IFNα, interferonα2b; CHOP, cyclophosphamide, doxorubicin, vincristin, and prednisone; RT, radiotherapy. Response to treatment: CR, complete remission; PR, partial remission; SD, stable disease. Status at last follow up: AW, alive and well; AD, alive with disease; DOD, died of disease; DNOD, died but not of disease.
expression, causing TIL apoptosis through the Fas pathway or inducing the expression of molecules such as killing inhibitors on TILs. The proportion of TILs in MF tends to reduce with the increase in the lymphoid infiltrate.

"With prolonged stimulation, subclones might appear that no longer require antigen dependent stimulation to proliferate, but show a tendency to infiltrate and disseminate."

Our data on the density of LCs, DDCs, and TILs are consistent with the above immunological model. Our series is mostly made up of early cases, some showing a mild infiltrate. However, we saw a wide spectrum of disease, with the increase of the lymphoid infiltrate being accompanied by a higher clinical stage and a worse response to treatment. Interestingly, a large proportion of patients with early disease and a mild lymphoid infiltrate showed a lower number of TILs compared with all the other patients. In daily pathology practice, patients like these represent a diagnostic problem between suspicious early MF or indeterminate lymphoid infiltrate. The diagnostic role of clonality and immunophenotypical analysis is still debated because aberrant phenotype and monoclonality have sometimes been described in bona fide reactive dermatoses. The diagnosis should be based on morphological features, such as the presence of lymphocytes with extremely convoluted, medium to large nuclei, single or clustered in the epidermis, and arranged in small sheets in the dermis. All our patients with low numbers of TILs showed the typical histological features of MF and were monoclonal with an aberrant phenotype. A possible explanation for low TIL density in these patients might be the absence of activation as a result of low

Figure 1 Type of subepidermal lymphoid infiltrate and density of CD8+ and CD1a+ cells in mycosis fungoides. (A,D) Microphotographs taken from two cases with different densities of neoplastic infiltrate and grades of epidermotropism: (A) mild lymphoid infiltrate, superficial, and with mild exocytosis and no tendency to confluence (infiltrate density and epidermotropism grades 1); (D) band-like subepidermal lymphoid infiltrate associated with more than three Pautrier’s microabscesses, one included in this field (infiltrate density and epidermotropism grades 3). (B,E) Immunostaining with anti-CD8 monoclonal antibody in two cases on frozen sections: (B) there are few and isolated dermal reactive lymphocytes in a case with dermal infiltrate grade 1 (CD8 positivity score 1); (E) CD8+ cells are numerous and form small groups in a case with dermal infiltrate grade 2 (CD8 positivity score 3). (C,F) Immunostaining with anti-CD1a monoclonal antibody in two cases on paraffin wax embedded sections: (C) there are few epidermal Langerhan’s cells and dermal dendritic cells, with no tendency to form small groups in a case with dermal infiltrate grade 1 (epidermal and dermal CD1a positivity score 1); (F) there are many Langerhan’s cells in all the epidermal layers with a tendency to cluster (epidermal CD1a positivity score 2), whereas the dermal dendritic cells are particularly numerous and form large aggregates (dermal CD1a positivity score 3); the case shows a dermal infiltrate grade 3.

Figure 2 Distribution of CD8 positivity scores in the four groups with different grades of subepidermal lymphoid infiltrate.

Figure 3 Distribution of dermal CD1a positivity scores in the four groups with different grades of subepidermal lymphoid infiltrate.
tumour specific antigen expression in MF cells. Thus, these cases could represent a very early step in MF; the next step could be the activation of TILs after exposure to a great number of tumour specific antigens. There is controversy on the soundness of treating these patients with early disease in the same way as those with more advanced MF. Large studies with long term follow up of patients with different proportions of CD8+ cells like those in our series are needed to clarify whether such patients exhibit a different clinical behaviour. Moreover, we found that cases with a denser lymphoid infiltrate rarely showed low TIL density, a feature explaining the improved survival rate described in advanced MF that was not associated with epidermotropism. LCs could be equally active in all disease phases. Their density difference in epidermal LC numbers between groups with different infiltrate grades show the same distribution, whereas numbers of TILs and DDCs tend to increase with the accumulation of neoplastic cells, suggesting that they are active against tumorous cells.

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