How should stainable iron in bone marrow films be assessed?

D A Hughes, S E Stuart-Smith, B J Bain

Iron deficiency is relatively common in the community as a result of both physiological and pathological losses. Usually, iron status is assessed initially by the use of surrogate markers, such as ferritin or combinations of more specialised haematinic markers—for example, serum transferrin receptor and the total iron binding capacity. However, although a low ferritin concentration is indicative of iron deficiency, the diagnosis may be missed when ferritin is raised in the context of inflammation. Similarly, anaemia of chronic disease may complicate the interpretation of haematinic markers when iron deficiency coexists. Therefore, the gold standard for the diagnosis of iron deficiency has been considered to be the assessment of a bone marrow aspirate stained with Perls’ stain. Historical papers comment on appropriate grading of iron stores on bone marrow aspirate preparations stained in this manner, but give no guidance as to how extensive an examination should be performed to distinguish between no visible iron (grade 0) or very slight stainable iron (grade 1) and normal or increased iron stores. Indeed, one study shows that up to 50% of specimens that were said to be deficient in iron did not correlate with the clinical iron status of the patient. This suggests that either the specimens or their examination were inadequate to identify the low amounts of iron present. It has been suggested that a minimum of two bone marrow films should be examined, but the number of particles that should be examined has not been assessed, and it seems likely that this would be of greater relevance. We aimed to answer the question of how many particles should be examined so that a confident assessment of the presence or absence of iron stores can be made.

METHODS

Aims: To identify how many particles should be examined to enable a confident assessment of the presence or absence of iron stores and the quantity of iron in a bone marrow aspirate to be made.

Methods: One hundred and ninety consecutive bone marrow aspirate samples were stained with Perls’ stain and the iron content of 10 consecutive particles was recorded. The first particle found to be positive and the particle that was most positive were also noted.

Results: A minimum of seven particles must be examined to establish the absence of stainable iron. A minimum of nine particles must be reviewed to see the maximum iron stores in 100% of samples and therefore make a valid judgment of whether iron stores are reduced, normal, or increased. By these criteria, 46% of the samples tested here could not be optimally assessed for absence of iron or maximum iron stores.

Conclusions: The sensitivity of examination of bone marrow aspirates for iron stores can be optimised by increasing the number of particles reviewed to seven or more. This may require the staining of additional slides.

Iron deficiency is relatively common in the community as a result of both physiological and pathological losses. Usually, iron status is assessed initially by the use of surrogate markers, such as ferritin or combinations of more specialised haematinic markers—for example, serum transferrin receptor and the total iron binding capacity. However, although a low ferritin concentration is indicative of iron deficiency, the diagnosis may be missed when ferritin is raised in the context of inflammation. Similarly, anaemia of chronic disease may complicate the interpretation of haematinic markers when iron deficiency coexists. Therefore, the gold standard for the diagnosis of iron deficiency has been considered to be the assessment of a bone marrow aspirate stained with Perls’ stain. Historical papers comment on appropriate grading of iron stores on bone marrow aspirate preparations stained in this manner, but give no guidance as to how extensive an examination should be performed to distinguish between no visible iron (grade 0) or very slight stainable iron (grade 1) and normal or increased iron stores. Indeed, one study shows that up to 50% of specimens that were said to be deficient in iron did not correlate with the clinical iron status of the patient. This suggests that either the specimens or their examination were inadequate to identify the low amounts of iron present. It has been suggested that a minimum of two bone marrow films should be examined, but the number of particles that should be examined has not been assessed, and it seems likely that this would be of greater relevance. We aimed to answer the question of how many particles should be examined so that a confident assessment of the presence or absence of iron stores can be made.

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particle noted to be positive and the particle with the highest iron content were noted.

Iron stores were graded according to the method of Rath and Finch6 (table 1).

RESULTS

Evaluation of bone marrow aspirate specimens for suitability for assessment of iron status

Of 190 consecutive bone marrow aspirate specimens, 87 (45.7%) contained 10 or more particles, 43 (22.6%) had no particles, and 60 (31.5%) had between one and 10 particles.

Figure 1 shows the distribution of the number of particles in each specimen. This distribution is polarised, with most preparations found to be completely aparticulate or to have an adequate number of particles. For the initial assessment, only samples with 10 or more particles were reviewed.

Minimum numbers of aspirate particles required to establish a diagnosis of iron deficiency

Eighty seven bone marrow aspirates with 10 or more particles were stained by Perls’ method for iron and the iron content of each consecutive particle was assessed. The first particle to show an iron grading of 1 or more was recorded. Figure 2 shows the cumulative distribution of samples becoming positive with each consecutive particle. The distribution plateaus at seven particles, when 90% of the sample shows at least one particle positive for iron. No further samples became positive even when 10 particles were reviewed. This indicates that seven particles must be assessed before a confident report of zero iron stores can be made.

Of our original 190 specimens, 43 had no particles (22.6%) and 12 (6.3%) had one to six particles without stainable iron and, therefore, using these new criteria should be considered not assessable.

Minimum number of aspirate particles required to establish the maximum iron status of a sample

The 87 bone marrow aspirate samples with 10 or more particles were then assessed to determine the particle exhibiting the maximum iron staining (fig 3). This distribution plateaus at nine particles, with no further increase in the grading of iron stores achievable with the 10th particle. The mean (SD) variation in iron content between particles in individual patients was 1.33 (0.9) units. Most (98.7%) samples had reached their maximum positivity by the seventh sample. Of the original 190 samples, 33 (17.3%) had visible iron but had only one to six particles and could not therefore be optimally graded for maximum positivity.

DISCUSSION

Our study shows that for the optimum assessment of a bone marrow aspirate for the absence of iron or for maximum iron stores a minimum of seven particles is required. Any sample that has fewer than seven particles and that contains no particles with stainable iron cannot be confidently said to have absent iron stores. This may explain why other studies have reported discrepancies between the bone marrow aspirate iron and the clinical status of the patient. A total of 46% of the samples in our study would have been considered not optimally assessable: 22.6% were aparticulate, 6.3% had only one to six particles and were negative, and 17.3% had only one to six particles but were positive. We were surprised that nearly a quarter of the samples were aparticulate. We suspect that this is an increasing phenomenon related to the demands for samples to be available for immunophenotyping, cytogenetic and molecular genetic analysis, and other specialised tests. We would always advocate the use of a small syringe to obtain a sample for the preparation of films before other diagnostic samples are taken; 0.2–0.3 ml is generally sufficient for this purpose. If it is clear that particles are not numerous, excess blood should be drained off before spreading further films. If the initial slide stained does not contain storage iron, further slides...
should be selected to give a minimum of seven fragments. Although the clinical importance of discriminating between grades 5 and 6 iron storage may be limited, it is essential to be able to assess marrow specimens for absent iron stores when investigating anaemia. An attempt should be made to optimise the number of particles to seven; if necessary, more than one slide should be stained and reviewed. If this cannot be achieved and no iron is seen on the existing particles, we suggest that the sample should be reported as lacking stainable iron, but the assessment should be stated to be unreliable. However, if a small initial sample was obtained for film preparation, it is likely that a definitive report could be issued on a single film much more often.

**Take home messages**

- When examining bone marrow aspirates for iron stores, a minimum of seven particles must be examined to establish the absence of stainable iron.
- At least nine particles must be reviewed to see the maximum iron stores in 100% of samples and therefore make a valid judgment of whether iron stores are reduced, normal, or increased.
- To achieve this, additional slides may need to be stained.
- If less than seven particles are present and no iron is seen, the sample should be reported as lacking stainable iron, but the assessment should be stated to be unreliable.

“We were surprised that nearly a quarter of the samples were aparticulate”

If fewer than seven particles are available for assessment in a thus far negative bone marrow, the examination of bone marrow trephine biopsy sections for haemosiderin might be helpful. This will be the subject of a further study.

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