Aims: Fetal hemoglobin (HbF) is an established serological indicator of cancer. However, its distribution in tumour tissues is rarely investigated. Therefore, HbF was studied immunohistochemically in different cancers characterised by high blood HbF concentrations.

Methods: Anti-HbF was immunoaffinity purified and used to study HbF immunohistochemically in the following cancers: germ cell tumour (GCT), trophoblastic disease (TD), lymphoma, myelodysplastic syndrome (MDS), multiple myeloma (MM), and ovarian adenocarcinoma (OA).

Results: In GCT a distinction was made between tumours substantially without HbF positive red blood cells (F-RBC) and those with F-RBC. Those without F-RBC were non-metastatic mature teratomas and dermoid cysts. Those containing F-RBC were mainly embryonal carcinomas and metastatic teratomas. HbF positive myeloid cells (F-MLC), HbF positive normoblasts (F-NBS), and F-RBC were common in the bone marrow and in the lymphoid tissues of lymphoma, MDS, and MM. In TD, normal and nucleated F-RBC were seen in the trophoblastic villi in one case with incomplete molar pregnancy (ICM) but not in other cases of ICM and complete molar pregnancy. However, F-RBC and F-MLC were seen in the decidua of both types of TD. Generally, F-cells were observed either within blood vessels or concentrated in certain areas of the neoplastic tissue.

Conclusions: HbF was evaluated as an inducible marker within different tumour tissue blood cells. The dual distribution of these cells—circulating in the blood or concentrated in areas of the neoplastic tissues—might reflect the two independent serological indicators of HbF: one in whole blood and the other in plasma of patients with cancer.

Materials and Methods

Subjects

The programme of this research, including studies on archival and stored clinical materials, was approved by the East London and the City Health Authority research ethics committee. Patients’ histological specimens were selected from the collection of the department of histopathology, Royal London Hospital, UK. They included the following diseases: GCT (26 cases), TD (14 cases), lymphoma (seven cases), MDS (seven cases), myeloma (seven cases), hairy cell leukaemia (10 cases), and OA (six cases). In parallel with the study of pathological tissues we tested normal tissues of the same organs.

Abbreviations: AFP, a fetoprotein; CM, complete molar pregnancy; EC, embryonal carcinoma; F-MLC, fetal haemoglobin positive myeloid cells; F-NBS, fetal haemoglobin positive normoblasts; F-RBC, fetal haemoglobin positive red blood cells; GCT, germ cell tumour; hCG, human chorionic gonadotrophin; HbF, fetal haemoglobin; HbFSRBC, fetal haemoglobin coated sheep red blood cells; ICM, incomplete molar pregnancy; MDS, myelodysplastic syndrome; MM, multiple myeloma; MMT, metastatic mature teratoma; NMT, non-metastatic mature teratoma; NSS, normal sheep serum; OA, ovarian adenocarcinoma; OD, optical density; PBS, phosphate buffered saline; RBC, red blood cells; TD, trophoblastic disease; YS, yolk sac
Affinity purification of anti-HbF antibody

The HbF antiserum was prepared in sheep and was rendered specific for HbF by absorbing it to HbA. The same antiserum has been used in the past in serological studies of HbF in patients with cancer. This antibody was further purified by absorbing it to HbF coated sheep red blood cells (HbFSRBC) and selectively eluting the anti-HbF. The HbFSRBC were prepared using glutaraldehyde as the coupling agent, as described previously. The affinity purification was performed by the following steps:

1. Extraction of the immunoglobulin by precipitation with 40% saturated ammonium sulfate.
2. Dialysis against phosphate buffered saline (PBS; pH 7.5).
3. Absorption with HbFSRBC for four hours at room temperature.
4. Disposal of the supernatant containing the unabsorbed protein, followed by washing the HbFSRBC with two volumes of PBS.
5. Elution of the anti-HbF from the HbFSRBC was performed batchwise by two steps.
   (a) Shaking with a 1:1 volume of 0.2M glycine/HCl (pH 2.7) for five minutes then saving the supernatant by spinning for five minutes at 1010 g at 4°C. The supernatant, designated eluate I, was immediately adjusted to pH 7.5 by the addition of 1M Na2HPO4.
   (b) The HbFSRBC from (a) were eluted again by shaking them with 1.3 volumes of 0.2M glycine/HCl (pH 2.7) for 15 minutes. The supernatant, designated eluate II, was saved and its pH adjusted as in (a).
6. Eluates I and II were dialysed against PBS then diluted 1:1 with glycerine for storage at 4°C.

The efficiency of purification was estimated by comparing the ratios of antibody titre to protein concentration, before and after purification. The anti-HbF titre was measured by the graded haemagglutination system, and the protein concentration was inferred from the optical density (OD) at 280 μm.

As a control for the immunohistochemical staining we repeated purification steps 1–6 on normal, unimmunised sheep serum (NSS).

Immunohistochemistry of tumours

To confirm the specificity of our purified anti-HbF we checked it in parallel with the control unimmunised antibody (NSS) on normal placenta. In the blood vessels of the trophoblastic villi of the normal placenta, approximately half the non-nucleated RBC and all the nucleated RBC were positively stained for HbF (fig 1). The control NSS, checked in parallel with the anti-HbF antibody, showed no traces of staining after the application of the chromagen.

Germ cells tumours

Tumours from 26 patients were studied (table 2) immunohistochemically for HbF. In some of the tumours the immunohistochemical results of conventional tumour markers hCG (human chorionic gonadotrophin) and AFP (α fetoprotein) were available from the archive reports, and these were compared with our HbF results. In embryonal carcinoma (EC), including sections of testis and retroperitoneal lymph nodes, high numbers of HbF positive RBC (F-RBC) were seen in three of four patients. The F-RBC were congested in “blood lakes” surrounding tumour cells (fig 2A).

<table>
<thead>
<tr>
<th>Purification fraction</th>
<th>Anti-HbF titre (A)</th>
<th>OD280 (protein concentration; B)</th>
<th>Ratio of anti-HbF to protein (A/B)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Original anti-HbF serum</td>
<td>1000</td>
<td>35</td>
<td>28.5</td>
</tr>
<tr>
<td>Eluate I</td>
<td>86</td>
<td>2.0</td>
<td>43</td>
</tr>
<tr>
<td>Eluate II</td>
<td>160</td>
<td>0.70</td>
<td>228.5</td>
</tr>
<tr>
<td>Eluate II (control; NSS)</td>
<td>0</td>
<td>0.73</td>
<td>0</td>
</tr>
</tbody>
</table>

Purification product in eluate II = 228.5/28.5 (eightfold). The recovery of anti-HbF in eluate II = 160 × 100/1000 = 16%.

HbF, fetal haemoglobin; NSS, normal sheep serum.

Table 1: Affinity purification of sheep anti-HbF serum

Figure 1: Immunostaining (orange cytoplasmic staining) of nucleated and non-nucleated red blood cells in the microvilli of normal placenta. Arrow, unstained normal RBC.
In two cases of metastatic mature teratoma (MMT), similar high numbers of F-RBC were seen in tumours of the brain. These cells were also congested in “blood lakes” and large haemorrhagic spaces, and comprised 30% of all RBC (fig 2B). In the blood vessels of these sections, 5% of the RBC were F-RBC. In two other cases of MMT, including one lymph node, and one testis, no F-RBC were seen. Five cases of non-metastatic mature teratoma (NMT) of the testis were all negative for F-RBC. Similarly, in nine cases of ovarian teratoma represented by dermoid cysts, no F-RBC were observed. However, in one case of ovarian dermoid cyst, high numbers of congested F-RBC were seen, resembling the other positive cases. A similar positive picture of F-RBC was seen in the germinoma component of an ovarian mixed yolk sac (YS) and germinoma tumour. The YS component was also HbF negative in another case of mixed YS and MMT.

**Trophoblastic disease**

Table 3 summarises the results for trophoblastic disease. Sections from nine patients with complete molar pregnancy (CM) and five patients with incomplete molar pregnancy (ICM) were analysed. No differences between the two types of TD were noted, except in one case of ICM where normal and nucleated F-RBC were seen in chorionic villi blood vessels, similar to normal placenta. In all other cases, no F-RBC were seen in the chorion. In most cases, there were young or mature forms of fetal haemoglobin positive myeloid cells (F-MLC) or fetal haemoglobin positive normoblasts (F-NBS), clustered or dispersed in inflammatory parts of the decidua (fig 2C, D) and the F-MLC were usually associated with F-RBC. In five cases, unusually high numbers of F-RBC were also seen in haemorrhagic spaces of the decidua, as was also seen in the GCTs (fig 2A).

**Lymphoma**

Lymph nodes from six patients were examined and in three of these samples 20% of the RBC in the haemorrhagic regions were F-RBC (fig 2E). Bone marrow samples were examined in four patients and all of these samples had small clusters (up to four cells), mainly of F-MLC, at different stages (fig 2F), occasionally with some F-NBS and F-RBC. The lymphocytes were all negative for HbF. In one spleen sample examined we also saw such clusters of F-cells with no HbF staining of the lymphocytes.

**Myelodysplastic syndromes**

In five of the seven cases we saw many large clusters of F-RBC, together with F-NBS, in the bone marrow (fig 3A).

---

Figure 2  (A) Immunostained (orange) fetal haemoglobin positive red blood cells (F-RBC) in “blood lakes” surrounding tumour cells in testicular embryonal carcinoma tissue. (B) Immunostained (pale to strong orange) F-RBC in large haemorrhagic space of embryonal carcinoma in brain. (C) Immunostaining (orange) of clustered fetal haemoglobin positive myeloid cells (F-MLC) and F-RBC in the decidua of incomplete molar pregnancy (ICM). (D) Immunostaining (orange) in one dispersed F-MLC in the decidua of ICM. (E) Immunostained (pale to strong orange) F-RBC in the haemorrhagic region of a lymph node from a patient with non-Hodgkin lymphoma. Arrow, unstained normal RBC. (F) Immunostained (orange) F-MLC in the bone marrow of a patient with non-Hodgkin lymphoma.
Table 2  Distribution of F-RBC in germ cell tumours

<table>
<thead>
<tr>
<th>No</th>
<th>Sex</th>
<th>Organ</th>
<th>Tumour</th>
<th>F-RBC distribution</th>
<th>Comment</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Male</td>
<td>Testis</td>
<td>EC</td>
<td>Congested (30%) F-RBC in blood lakes surrounding the tumour cells (fig 2A)</td>
<td>hCG and AFP negative</td>
</tr>
<tr>
<td>2</td>
<td>Male</td>
<td>Testis</td>
<td>EC</td>
<td>Positive as in 1. Also 2–5% F-RBC in blood vessels</td>
<td>No F-RBC in the counterpart normal tests</td>
</tr>
<tr>
<td>3</td>
<td>Male</td>
<td>Retroperitoneal lymph node</td>
<td>EC</td>
<td>Positive as in 2</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>Male</td>
<td>Testis</td>
<td>EC</td>
<td>None</td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>Male</td>
<td>Brain</td>
<td>MMT</td>
<td>Congested (30%) F-RBC in blood lakes and haemorrhagic zones (fig 2B) in blood vessels 5%</td>
<td>hCG and AFP positive</td>
</tr>
<tr>
<td>6</td>
<td>Female</td>
<td>Brain</td>
<td>MMT</td>
<td>Positive as in 5</td>
<td>hCG positive FAP negative</td>
</tr>
<tr>
<td>7</td>
<td>Male</td>
<td>Retroperitoneal lymph node</td>
<td>Tests</td>
<td>None</td>
<td></td>
</tr>
<tr>
<td>8</td>
<td>Male</td>
<td>Tests</td>
<td>MMT</td>
<td>None</td>
<td></td>
</tr>
<tr>
<td>9</td>
<td>Female</td>
<td>Ovary</td>
<td>Mixed germinoma and YS</td>
<td>In germinoma positive as in 1. In YS none</td>
<td>AFP positive in YS, hCG negative</td>
</tr>
<tr>
<td>10–14</td>
<td>Male</td>
<td>Testis</td>
<td>NMT</td>
<td>None</td>
<td></td>
</tr>
<tr>
<td>15</td>
<td>Male</td>
<td>Brain</td>
<td>Mixed seminoma and mature teratoma</td>
<td>None</td>
<td></td>
</tr>
<tr>
<td>16</td>
<td>Female</td>
<td>Lymph node</td>
<td>Mixed YS and mature teratoma</td>
<td>None</td>
<td>AFP positive in YS</td>
</tr>
<tr>
<td>17</td>
<td>Female</td>
<td>Ovary</td>
<td>Dermoid cyst</td>
<td>Positive as in 2</td>
<td></td>
</tr>
<tr>
<td>18–26</td>
<td>Female</td>
<td>Ovary</td>
<td>Dermoid cyst</td>
<td>None</td>
<td></td>
</tr>
</tbody>
</table>

AFT, a fetoprotein; EC, embryonal carcinoma; F-RBC, fetal haemoglobin positive red blood cells; hCG, human chorionic gonadotrophin; MMT, metastatic mature teratoma; NMT, non-metastatic mature teratoma; TD, trophoblastic disease; YS, yolk sac.

Table 3  The distribution of fetal haemoglobin positive cells in trophoblastic diseases

<table>
<thead>
<tr>
<th>No</th>
<th>Disease</th>
<th>In inflammatory decidua</th>
<th>In haemorrhagic decidua</th>
<th>In chorion</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>CM</td>
<td>Randomly dispersed single F-RBC and F-MLC</td>
<td>(–)</td>
<td>None</td>
</tr>
<tr>
<td>2</td>
<td>CM</td>
<td>Clustered and dispersed F-MLC and F-RBC (fig 2C)</td>
<td>2–3% F-RBC</td>
<td>None</td>
</tr>
<tr>
<td>3</td>
<td>CM</td>
<td>Randomly dispersed single F-RBC and F-MLC</td>
<td>(–)</td>
<td>None</td>
</tr>
<tr>
<td>4</td>
<td>CM</td>
<td>Large clusters of up to 20 F-MLC and F-RBC</td>
<td>8% F-RBC</td>
<td>None</td>
</tr>
<tr>
<td>5</td>
<td>CM</td>
<td>None</td>
<td>0.3% F-RBC</td>
<td>None</td>
</tr>
<tr>
<td>6</td>
<td>CM</td>
<td>None</td>
<td>1% F-RBC</td>
<td>None</td>
</tr>
<tr>
<td>7</td>
<td>CM</td>
<td>Randomly dispersed single F-RBC and F-MLC</td>
<td>(–)</td>
<td>None</td>
</tr>
<tr>
<td>8</td>
<td>CM</td>
<td>None</td>
<td>(–)</td>
<td>None</td>
</tr>
<tr>
<td>9</td>
<td>CM</td>
<td>None</td>
<td>(–)</td>
<td>None</td>
</tr>
<tr>
<td>10</td>
<td>ICM</td>
<td>Clusters of F-MLC mainly with only few F-RBC</td>
<td>(–)</td>
<td>None</td>
</tr>
<tr>
<td>11</td>
<td>ICM</td>
<td>Clusters of F-MLC mainly with only few F-RBC</td>
<td>(–)</td>
<td>None</td>
</tr>
<tr>
<td>12</td>
<td>ICM</td>
<td>Clusters of F-MLC mainly with only few F-RBC</td>
<td>2% F-RBC</td>
<td>None</td>
</tr>
<tr>
<td>13</td>
<td>ICM</td>
<td>Randomly dispersed single F-MLC (fig 2D)</td>
<td>2% F-RBC</td>
<td>None</td>
</tr>
<tr>
<td>14</td>
<td>ICM</td>
<td>Decidua not present</td>
<td>F-RBC mature and nucleated</td>
<td></td>
</tr>
</tbody>
</table>

(–), No haemorrhagic decidua present; CM, complete molar pregnancy; F-MLC, fetal haemoglobin positive myeloid cells; F-RBC, fetal haemoglobin positive red blood cells; ICM, incomplete molar pregnancy.

Multiple myeloma
In the bone marrow of four of seven patients we saw many large clusters containing a mixed population of F-MLC, F-NBS, and F-RBC. At the periphery of the bone marrow, there were only F-RBC (fig 3B). In all the samples examined the plasma cells were negative for HbF.

Hairy cell leukaemia
Bone marrow samples from 10 patients were examined and F-RBC were seen in only one. In one spleen sample examined, 15% of the cells in the blood vessels were F-RBC and in its pulp there were clusters of F-RBC together with F-NBS (fig 3C).

Ovary adenocarcinoma
Ovarian tumours from six patients were tested. When the usual dilution of the 1/20 anti-HbF antibody was used no positive results were seen, but when the antibody was less dilute (1/5 dilution), F-RBC were detected in two of the six patients. These F-RBC were seen in the blood vessels (1–2%) and dispersed in regions of loose connective or fat tissue.

Controls
In parallel with each pathological organ (testis, ovary, brain, lymph nodes, spleen, and bone marrow), at least three samples of normal organs were examined by the same immunochemical techniques for F-cells. All these controls were HbF negative.

DISCUSSION
Our affinity purified anti-HbF antibody did not detect HbF in the tumour cells of GCTs. This is in contradiction to previous work demonstrating immunostaining of HbF in tumour cells. With regard to our previous preliminary work, we were able to prove that the positive staining of tumour cells was non-specific, because affinity purification of the same anti-HbF antibody obliterated that staining. We found that the HbF in our GCT samples was present in RBC (F-RBC) rather than tumour cells. Although the number of specimens was small, it seems that in GCT F-RBC are indicators of aggressive rather than less aggressive tumours (table 2). Most cases of EC and two MMT cases (table 2) contained F-RBC. Like MMT, EC was previously reported as a metastatic disease in approximately 66% of patients. Similarly, in dysgerminoma (table 2), previously reported as a metastatic disease, there were many F-RBC. In contrast,
in the less aggressive disease NMT (table 2), and in dermoid cyst (table 2), known to be a benign tumour, there were essentially no F-RBC. The extravascular congestion of F-RBC is probably the source of plasma HbF in patients with GCT. The large blood vessels seen in all of these tumours do not contain F-RBC or have fewer numbers of F-RBC than is seen in the extravascular regions of HbF positive GCTs. HbF, although not produced by the tumour cells, might be a potential tumour indicator, complementary to the conventional markers of GCT, such as hCG and AFP. For instance, in table 2 we see that in one case (number 1) with a high number of F-RBC neither hCG nor AFP was detected, whereas in another case (number 9), F-RBC were congested in the germinoma compartment but AFP was detected only in the YS compartment.

"The extravascular congestion of fetal haemoglobin positive red blood cells is probably the source of plasma fetal haemoglobin in patients with germ cell tumours"

In TD (table 3) the extravascular F-cells are also probably the source of the plasma HbF found in this disease. In TD, we found in addition to F-RBC, young and mature F-MLC in clusters or single cells in the inflammatory decidua (fig 2C, D), which were only a minority of the total MLC population of that zone. The presence of F-MLC in TD is interesting, and should be investigated further to reveal the relevance and specificity of these cells in these tumours. However, we were unable to evaluate the difference between CM and ICM. The only case with F-RBC in the chorionic villi may be related to an advanced stage of ICM, although more cases would need to be studied to prove this assumption.

Very high numbers of extravascular F-RBC were also found in the lymph nodes of lymphoma, in accord with the high concentration of plasma HbF in patients with lymphoma. The presence of such extravascular F-RBC in lymphoma and in GCT has not been reported previously. No evidence was found for their regeneration within the tumour because there were no precursors of F-cells. Further studies should be conducted to evaluate the role of these F-RBC in tumour development. However, they may have been attracted from the circulation by the tumour cells, which would have decreased the concentration of HbF in the whole blood, eventually leading to increased plasma HbF. This assumption is supported by the results of the concomitant measurement of whole blood and plasma HbF concentrations—in general, they were inversely related. 7 12 The detection of such changes between the concentrations of whole blood and plasma HbF might be exploited for diagnostic purposes. However, to verify such a proposed dynamic, a follow up study for HbF in whole blood versus plasma must be performed. Our immunohistochemical findings of F-cells in two spleen samples, one from lymphoma, the other from hairy cell leukaemia, are very similar to previous findings. In both of these studies there was evidence for fetal haemoipoiesis in the spleen of lymphoproliferative and myeloproliferative disease, in addition to the fetal spleen. Fetal haemoipoiesis does not occur in the spleen of healthy adults, so that the spleen is probably the source of the raised whole blood HbF concentrations previously found in patients with hairy cell leukaemia. In MDS, the dense clusters of F-cells may also be the source of extremely high plasma HbF found in such patients. The large clusters of F-cells containing haemoipoietic cells (NSB or young MLC) found in the spleen and bone marrow of the patients with haematological malignancies are apparently germinal centres. However, they are not the source of malignancy because the malignancy arose from lymphoid or plasma cells. Thus, in those cases (as in germ cell tumours), HbF is not a direct oncofetal antigen produced by the tumour cells, but an inducible antigen. The mechanism of this induction in adults is not yet clear, and is the subject of a current investigation. For instance, it was shown in vitro that some tumour cell lines, including GCT (teratocarcinoma), induced HbF production in precursor erythroid cells. Later on it was found that particular growth factors, including c-kit and interleukin 3, stimulated HbF production in such erythroid cells. One source (among others) of c-kit is lymphoma cells. The presence of F-MLC in the bone marrow of haematological malignancies and in the decidua of TD was surprising because myelocytes do not normally produce haemoglobin. However, this is not the first finding of HbF synthesis in transformed myelocytic cells, because the...
expression of HbF (γ and α globins) genes has been reported in myelocytic leukaemic cells. In other studies, myelocytic leukaemic cells grown in culture differentiate spontaneously or by chemical induction, or by erythroid cells producing HbF.

The concentration of HbF in F-RBC from OA was lower than that found in the other tumours. It is probable that such poorly detectable F-cells also exist in other tumours. To investigate this question we need to increase the specific activity of our affinity purified anti-HbF antibody.

“The presence of fetal haemoglobin positive myelocytes in the bone marrow of haematological malignancies and in the decidua of trophoblastic disease was surprising because myelocytes do not normally produce haemoglobin”

In conclusion, we have provided immunohistochemical evidence of the source of HbF in both plasma and whole blood, which can be used as an indicator of cancer, and have proved the independence of these two sources.

This adds to our understanding of the importance of serologically high concentrations of HbF in plasma or in whole blood and raises intriguing possibilities of this mechanism of tumour inducible gene expression as a model of other paraneoplastic phenomena.

ACKNOWLEDGEMENTS

We are grateful to C. Nichols and A. Brown from the unit of experimental pathology at the Royal London Hospital, for their excellent methodological and technical assistance. This work was partially supported by a travel grant to Dr M. Wolk from the Leo Foundation.

Authors’ affiliations

M. Wolk, Department of Morbid Anatomy and Histopathology, The Royal London Hospital, Central Laboratory, London E1 1BB, UK

J. E. Martin, Pathology Group, Institute of Cell and Molecular Sciences, The Royal London Hospital, London E1 1BB, UK

R. Constantini, Department of Pathology, Shaare-Zedek Medical Centre, Jerusalem 91301, Israel

REFERENCES


Blood cells with fetal haemoglobin (F-cells) detected by immunohistochemistry as indicators of solid tumours
M Wolk, J E Martin and R Constantin

J Clin Pathol 2004 57: 740-745
doi: 10.1136/jcp.2003.013938

Updated information and services can be found at:
http://jcp.bmj.com/content/57/7/740

These include:

References
This article cites 24 articles, 8 of which you can access for free at:
http://jcp.bmj.com/content/57/7/740#BIBL

Email alerting service
Receive free email alerts when new articles cite this article. Sign up in the box at the top right corner of the online article.

Topic Collections
Articles on similar topics can be found in the following collections
Immunology (including allergy) (1664)
Small intestine (45)

Notes

To request permissions go to:
http://group.bmj.com/group/rights-licensing/permissions

To order reprints go to:
http://journals.bmj.com/cgi/reprintform

To subscribe to BMJ go to:
http://group.bmj.com/subscribe/