Differentiating primary and extragenital metastatic mucinous ovarian tumours: an algorithm combining PAX8 with tumour size and laterality

Ajin Hu, Hongwei Li, Letian Zhang, Caixia Ren, Yuxiang Wang, Yan Liu, Congrong Liu

ABSTRACT

Aims Our aim was to analyse the utility of the algorithm combining PAX8 with clinicopathological characteristics (tumour size, laterality and patient age) in differentiating primary ovarian mucinous tumours (POMTs) from extragenital metastatic mucinous carcinomas involving the ovary (eMOMCs).

Methods and results Immunohistochemical staining for PAX8 was performed on formalin fixed, paraffin embedded tissues from 47 POMTs, 18 eMOMCs and 70 extragenital primary mucinous carcinomas (POMCs) using anti-PAX8 rabbit polyclonal antibody (pAb) and anti-PAX8 rabbit monoclonal antibody (mAb). PAX8 (pAb) positive signals were found in 3/18 eMOMCs and in 32/70 POMCs. PAX8 (mAb) demonstrated superior specificity, with 0% positivity in both eMOMCs and ePOMCs, but unfavourable sensitivity, with 60.9% in ovarian mucinous borderline tumours and 45.8% in POMCs. Although PAX8 (mAb) immunostaining status (66.2%), tumour size (75.4%) and laterality (84.6%) demonstrated unsatisfactory accuracy when they were evaluated individually in differentiating POMTs from eMOMCs, a combination of PAX8 (mAb) immunostaining status, tumour size and laterality markedly increased accuracy (86.2%), with a satisfactory Youden Index (63.7%).

Conclusions PAX8 (mAb) was a specific marker in differentiating POMTs from eMOMCs. As a simple, convenient and high performance to price ratio algorithm, a combination of PAX8 (mAb) immunostaining with tumour size and laterality will improve the diagnostic criteria of ovarian mucinous metastasis.

INTRODUCTION

The distinction between primary ovarian mucinous tumours (POMTs) and metastatic mucinous carcinomas involving the ovary (MOMCs) is often problematic.1 MOMCs commonly metastasise from the alimentary system, breast and cervix, which sometimes produces histological patterns as well as immunohistochemical features closely resembling well differentiated primary ovarian mucinous carcinomas (POMCs), or benign or borderline POMTs.2–6 Occasionally, the primary tumours of MOMCs may be clinically silent, only presenting with symptoms related to an ovarian mass, and may not manifest until a period of time after total abdominal hysterectomy with bilateral salpingo-oophorectomy. As a result, it is difficult to differentiate between primary and metastatic mucinous neoplasms based on clinical and histopathology features alone, even though the differential diagnosis is decisive for treatment and prognosis.2

Seidman et al1 first recommended a simple algorithm (bilateral tumours of any size, or unilateral tumour <10 cm = metastatic; unilateral tumour ≥10 cm = primary) and claimed it could accurately classify 90% of mucinous neoplasms involving the ovary. However, some cases violate the algorithm.2 To deal with an equivocal tumour, immunohistochemistry (IHC) may be helpful, although it is not definitive.7 Several candidate markers have been advocated, including PAX8, which is a member of the paired box gene family of transcription factors and plays a critical role in the organogenesis of the Müllerian system.8 In addition, PAX8 has the potential to induce tumorigenesis and is expressed in a tissue specific manner during neoplastic transformation.9 Recently, PAX8 has been shown to be constantly expressed in the majority of histological subtypes of Müllerian epithelial tumours.13 Absence of PAX8 expression in mammary carcinoma and malignant mesothelioma is especially valuable, considering that ovarian involvement of these tumours is not uncommon.14 Nevertheless, the percentage of PAX8 positivity in POMCs has ranged from 0% to 50% in previous studies.14–23

With the growing awareness that true POMCs appear to be substantially less common than previously reported, we believe that PAX8 expression in POMTs should be explored based on full compliance with stringent morphological criteria, in conjunction with detailed clinical information and close follow-up. The other reason behind the disparity may be the variety of anti-PAX8 antibodies used in different studies. The majority of the published studies used anti-PAX8 rabbit polyclonal antibody (pAb).14–22 Unfortunately, currently available commercial pAbs have cross reactions with lymphocytes, metastatic pancreatic cancers, duodenal neuroendocrine tumours and a subset of rectal, gastric and appendiceal neuroendocrine tumours, which could confuse the interpretation and thus influence the confidence and accuracy of the diagnosis.13–24

In addition, information about PAX8 expression in different POMTs categories is limited and, to date, no study has been performed to verify PAX8 expression in POMTs, in extragenital metastatic mucinous carcinomas involving the ovary (eMOMCs) or in extragenital primary mucinous carcinomas (ePOMCs).

Therefore, our aim was to confirm the exact role of PAX8 in the differential diagnosis of POMTs,
and to develop an algorithm to improve the accuracy of this clinical practice.

MATERIALS AND METHODS

Tissue specimens

Formalin fixed, paraffin embedded tissues were selected from the Department of Pathology, Peking University Health Science Centre. Forty-seven POMTs (23 borderline, 24 carcinomas), 18 eMOMCs (7 from the colorectum, 6 from the appendix, 4 from the stomach and 1 from the pancreas) and 70 ePMCs (29 colorectal, 16 mammary, 13 gastric, 7 pancreatic, 3 appendiceal and 2 biliary tract) were from patients who underwent surgery from 1999 to 2011.

Twenty-one cases of POMCs were stage I while only two cases were stage II and one was stage IV. All pathological diagnoses for specimens used in this study were reviewed using established histological criteria, and were confirmed by two pathologists. Brieﬂy, tumours were diagnosed as mucinous borderline tumours (MBTs) when they showed non-invasive tumours with intraglandular or intracytic epithelial proliferation in the absence of disturbing architectural features of the lining epithelium, and with mild to moderate nuclear atypia. A tumour was classiﬁed as MBT with intraepithelial carcinoma if a borderline tumour contained foci in which the epithelial lining cells had marked atypical nuclei or had appreciable architectural proliferation but a cribriform pattern on its own or epithelial stratification of greater than three cell layers in the absence of severe atypia. MBTs with micrometastasis were diagnosed when a borderline tumour had stromal invasion of <5 mm in a contiguous area. Tumours were classiﬁed as POMCs when they exhibited stromal invasion of >5 mm, characterised by an expansile growth pattern or cribriform pattern, without having discrete nodularity. Tumours were classiﬁed as MOMCs when they exhibited surface involvement, stromal invasion characterised by a multinodular growth pattern in which the mucinous gland aggregates were separated from each other by regions of preserved ovarian stroma or areas simulating a benign/borderline appearance but displaying signiﬁcant cytological atypia.

All patients, except those who presented with ePMCs without ovary involvement, were closely followed-up for at least 3 years. All available clinical data and follow-up outcomes were added to determine the primary sites for the metastatic cases and to exclude the possibility of metastatic tumours in diagnosing POMTs. Four cases with equivocal clinicopathological data and an undetermined final diagnosis were excluded. Tumour size (if bilateral involvement, we recorded the size of the larger tumour), laterality and patient age were recorded for all cases of POMTs and eMOMCs.

Immunohistochemistry

Representative haematoxylin–eosin stained slides from the tumours were reviewed, and one representative block was selected for IHC. Immunohistochemical analyses were performed on formalin fixed, paraffin embedded, 4 μm thick tissue sections. The primary antibodies used were anti-PAX8 rabbit monoclonal antibody (mAb) (Zeta, Corporation, California, USA) and pAb (Protein Tech Group, Chicago, Illinois, USA). Tissue sections were deparafﬁnised and rehydrated through xylene and graded ethanol solutions to water. Antigen retrieval was performed using pressure cooker pretreatment in Tris EDTA buffer (pH 9.0) for 2 min. After blocking the endogenous peroxidase activity with 3% hydrogen peroxidase for

<table>
<thead>
<tr>
<th>Table 1</th>
<th>PAX8 expression in POMTs, eMOMCs and ePMCs</th>
</tr>
</thead>
<tbody>
<tr>
<td>PAX8 mAb</td>
<td>PAX8 pAb</td>
</tr>
<tr>
<td>Staining intensity</td>
<td>Positive/total cases (%)</td>
</tr>
<tr>
<td>POMTs 25/47 (53.2)</td>
<td>22</td>
</tr>
<tr>
<td>MBTs 14/23 (60.9)</td>
<td>9</td>
</tr>
<tr>
<td>POMCs 11/24 (45.8)</td>
<td>13</td>
</tr>
<tr>
<td>FIGO stage</td>
<td></td>
</tr>
<tr>
<td>Ia–Ic 10/21 (47.6)</td>
<td>11</td>
</tr>
<tr>
<td>II–IV 1/3 (33.3)</td>
<td>2</td>
</tr>
<tr>
<td>eMOMCs 0/18 (0)</td>
<td>0</td>
</tr>
<tr>
<td>Colorectal 0/7 (0)</td>
<td>0</td>
</tr>
<tr>
<td>Appendiceal 0/6 (0)</td>
<td>0</td>
</tr>
<tr>
<td>Gastric 0/4 (0)</td>
<td>0</td>
</tr>
<tr>
<td>Pancreatic 0/1 (0)</td>
<td>0</td>
</tr>
<tr>
<td>ePMCs 0/70 (0)</td>
<td>0</td>
</tr>
<tr>
<td>Colorectal 0/29 (0)</td>
<td>0</td>
</tr>
<tr>
<td>Gastric 0/13 (0)</td>
<td>0</td>
</tr>
<tr>
<td>Appendiceal 0/3 (0)</td>
<td>0</td>
</tr>
<tr>
<td>Pancreatic 0/7 (0)</td>
<td>0</td>
</tr>
<tr>
<td>Biliary tract 0/2 (0)</td>
<td>0</td>
</tr>
<tr>
<td>Mammary 0/16 (0)</td>
<td>0</td>
</tr>
</tbody>
</table>

Staining intensity is graded 0–3. Nuclear staining in >5% of malignant cells was considered positive.

*POMTs (MBTs vs carcinoma), Fisher’s exact test.
†POMCs (FIGO stage Ia–Ic vs FIGO stage II–IV), Fisher’s exact test.
‡POMTs vs eMOMCs, Fisher’s exact test.
§POMs vs ePMCs, Fisher’s exact test.
eMOMCs, extragenital metastatic mucinous carcinomas involving the ovary; ePMCs, extragenital primary mucinous carcinomas; mAb, monoclonal antibody; MBTs, ovarian mucinous borderline tumours; POMCs, primary ovarian mucinous carcinomas; POMTs, primary ovarian mucinous tumours.

10 min, the sections were incubated with primary antibody (mAb, dilution: 1:100; pAb, dilution: 1:100) overnight at 4°C. Detection reactions utilised the Envision kit from Dako (Dako Cytomation, Glostrup, Denmark). Diaminobenzidine was used as the chromogen and haematoxylin as the counterstain. Appropriate positive (serous ovarian carcinoma tissue) and negative (incubation with Tris buffered saline instead of the specific primary antibody) controls were run simultaneously.

Analyses of IHC results were performed by two independent authors (AH and HL). Discrepancies in analyses were reconciled following a review by a third reviewer (CL). Nuclear staining in more than 5% of malignant cells was considered positive. For positive cases, staining intensity was further scored as follows: 0, no staining; 1, unequivocal but weak intensity; 2, moderate intensity; and 3, marked intensity.

Statistical methods
The Kruskal–Wallis test, Mann–Whitney test and Fisher’s exact test were used to compare differences in PAX8 expression, tumour size, laterality and patient age in the mucinous tumours. The receiver operator characteristic (ROC) curve was used to detect the sensitivity and specificity of PAX8 (mAb) IHC status, tumour size, laterality and patient age in differentiating POMTs from eMOMCs. The value of the area under the ROC curve (AUC) was between 0 and 1. A test with an AUC value of 0.5–0.7 has weak overall diagnostic performance, an AUC of 0.7–0.9 has moderate overall diagnostic performance and an AUC of 0.9–1 has a favourable overall diagnostic performance.27 A p value <0.05 was considered statistically significant.

RESULTS
The results of IHC are summarised in table 1. PAX8 staining signals were localised on nuclei of tumour cells (figure 1). PAX8 (mAb) did not show any reactivity with eMOMCs (0/18) or ePMCs (0/70) tumour cells, while positive signals were seen in 60.9% (14/23) of MBTs and in 45.8% (11/24) of POMCs. In contrast, PAX8 (pAb) not only stained a high percentage of POMTs (82.6% (19/23) MBTs and 75% (18/24) POMCs), but also showed positive signals in 16.7% (3/18) of eMOMCs and in a high proportion of ePMCs, including 41.4% (12/29) colorectal, 68.8% (11/16) mammary, 23.1% (3/13) gastric, 42.9% (3/7) pancreatic, 66.7% (2/3) appendiceal and 50% (1/2) biliary tract. Additionally, unlike PAX8 pAb, PAX8 mAb did not exhibit any cross reaction with B cells or pancreatic cells. Although both antibodies demonstrated meaningful roles in the differential diagnosis of mucinous tumours involving the ovary (table 1), our data strongly suggested that PAX8 (mAb) was far more specific than PAX8 (pAb), especially in differentiating POMTs from eMOMCs. Consequently, only PAX8 (mAb) IHC results were analysed in the following studies.

Compared with POMTs, eMOMCs were more frequently bilateral (55.6% vs 4.2%, p<0.01), showed a much smaller

Figure 1  PAX8 (mAb) immunohistochemistry in POMTs, eMOMCs and ePMCs. (A) MBTs: marked PAX8 nuclear reaction. (B) POMCs: moderate PAX8 nuclear reaction. (C) Colorectal MOMCs: negative PAX8 reaction. (D) Colorectal PMCs: negative PAX8 reaction. eMOMCs, extragenital metastatic mucinous carcinomas involving the ovary; ePMCs, extragenital primary mucinous carcinomas; mAb, monoclonal antibody; MBTs, ovarian mucinous borderline tumours; MOMCs, metastatic mucinous carcinomas involving the ovary; PMCs, primary mucinous carcinomas; POMCs, primary ovarian mucinous carcinomas; POMTs, primary ovarian mucinous tumours.
mean tumour size (11.1 vs 18.8 cm, \( p < 0.01 \)) and presented at a higher mean age (51.8 vs 40.6 years, \( p < 0.05 \)) (Figure 2). Based on AUC, PAX8 (mAb) IHC status (\( 0.766, 95\% \) CI 0.654 to 0.878; \( p < 0.01 \)), tumour size (\( 0.870, 95\% \) CI 0.779 to 0.961; \( p < 0.01 \)), laterality (\( 0.757, 95\% \) CI 0.605 to 0.908; \( p < 0.01 \)) and patient age (\( 0.645, 95\% \) CI 0.497 to 0.792; \( p = 0.073 \)) were assessed (Figure 3). All of the above parameters, except patient age (AUC <0.7), demonstrated some success in distinguishing POMTs from eMOMCs. However, the accuracy of each individual parameter (66.2\% for PAX8 (mAb) IHC status, 75.4\% for tumour size and 84.6\% for laterality) and the accuracy of the algorithm proposed by Seidman et al. \(^3\) (76.6\% for POMTs and 72.2\% for eMOMCs) were both unsatisfactory in our cohort. Therefore, we tried combinations of the above parameters. Interestingly, when PAX8 (mAb) IHC status was combined with tumour size and laterality, diagnostic accuracy increased markedly to 86.2\%, with the highest Youden Index (63.7\%), which was superior to all other combinations (table 2).

**DISCUSSION**

Our study confirmed the most frequent primary sources of eMOMCs as colorectum (38.9\%, 7/18), followed by appendix (33.3\%, 6/18) and upper gastrointestinal tract (27.8\%, 5/18). MOMCs from these organs can be potentially difficult to diagnose, as their primary tumours may be insidious and clinically silent, and they are often diagnosed synchronously with or even after the ovarian tumours.\(^1\) In our cohort, 6 of 18 cases of eMOMCs were misdiagnosed as POMCs at presentation and were proved to be metastatic lesions 1–3 years later. MOMCs are always associated with increased aggressiveness, and their behaviour is diametrically opposite to that of the majority of POMCs, which are in most cases at an early stage of diagnosis.\(^2\)

Most (95.8\%, 23/24) of our POMCs were FIGO stages I and II at presentation. Consistent with previous studies, POMCs appear to develop from benign and borderline POMTs, which tend to be the largest ovarian masses overall and usually involve only one ovary; MOMCs are often much smaller and bilateral. As a result, it would be of great value to distinguish POMTs from MOMCs in the intraoperative diagnosis. However, difficulty distinguishing POMTs from MOMCs can extend beyond the intraoperative period, even to when permanent sections are evaluated. Some MOMCs, especially from pancreatic and intestinal primary tumours, can closely mimic POMTs morphologically and can occur in the absence of a known extra-ovarian
primary site. Thus, to correctly and properly diagnose problematic cases, it is essential to use more advanced molecular markers in clinical practice.

PAX8 has been shown to be a sensitive and specific differential diagnostic marker for carcinomas of ovarian origin. Chu et al. proved that PAX8 expression was only found in gynaecology mucinous carcinomas but not in other types of mucinous carcinomas. However, the amount of PAX8 positivity (ie, <50%) demonstrated great discrepancy due to various morphological criteria and the multifarious anti-PAX8 antibodies used in different studies of POMTs (table 3). The diagnoses of POMTs and eMOMCs of our cohort were rigorously based on stringent histological criteria and were validated by detailed clinical records as well as long term (≥3 years) follow-up.

Compared with previously published data, our study demonstrated unusually high expression of PAX8 (pAb) in POMTs and should be further validated in more studies. Also, pAb demonstrated unsatisfactory specificity in our study and was eliminated from our final analysis. Consistent with previous studies, less than half (45.8%, 11/24) of our POMCs reacted with PAX8 (mAb). The lower positivity of PAX8 in POMCs compared with ovarian serous (93–100%), endometrioid (83–100%) and clear cell (100%) carcinomas supports the theory that POMCs are unique among other epithelial ovarian carcinomas subtypes. Moreover, PAX8 is a lineage restricted transcription factor that plays an essential role in the organogenesis of the Müllerian system. In the reproductive tract, PAX8 expression was shown to be restricted to secretory cells of the fallopian tube epithelium which has recently been suggested to be cell origins for most serous ovarian carcinomas. Recent studies have provided evidence that microscopic transitional cell nests at the tubal–peritoneal junction or fimbrial epithelial cells of the fallopian tube could be the potential source of POMCs. This would be consistent with their non-Müllerian appearance and

Table 2

<table>
<thead>
<tr>
<th>POMTs (n)</th>
<th>MOMCs (n)</th>
<th>Sensitivity (%)</th>
<th>Specificity (%)</th>
<th>PPV (%)</th>
<th>NPV (%)</th>
<th>Accuracy (%)</th>
<th>Youden Index (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>unilateral=primary, bilateral=metastatic</td>
<td>45</td>
<td>10</td>
<td>95.7</td>
<td>55.6</td>
<td>84.9</td>
<td>83.3</td>
<td>84.6</td>
</tr>
<tr>
<td>≥10 cm=primary, &lt;10 cm=metastatic</td>
<td>38</td>
<td>11</td>
<td>80.9</td>
<td>61.1</td>
<td>84.4</td>
<td>55.0</td>
<td>75.4</td>
</tr>
<tr>
<td>PAX8 mAb (+)=primary, PAX8 mAb (−)=metastatic</td>
<td>25</td>
<td>18</td>
<td>54.2</td>
<td>100.0</td>
<td>100.0</td>
<td>45.0</td>
<td>66.2</td>
</tr>
<tr>
<td>Unilateral ≥10 cm=primary, bilateral tumours of any size or unilateral tumour &lt;10 cm=metastatic</td>
<td>36</td>
<td>13</td>
<td>76.6</td>
<td>72.2</td>
<td>87.8</td>
<td>51.2</td>
<td>75.4</td>
</tr>
<tr>
<td>PAX8 mAb (+) or PAX8 mAb (−) and ≥10 cm=primary, PAX8 mAb (−) and &lt;10 cm=metastatic</td>
<td>43</td>
<td>11</td>
<td>91.5</td>
<td>61.1</td>
<td>86</td>
<td>73.3</td>
<td>83.1</td>
</tr>
<tr>
<td>PAX8 mAb (+) or PAX8 mAb (−) and unilateral=primary, PAX8 mAb (−) and bilateral=metastatic</td>
<td>45</td>
<td>10</td>
<td>95.7</td>
<td>55.5</td>
<td>84.9</td>
<td>83.3</td>
<td>84.6</td>
</tr>
<tr>
<td>PAX8 mAb (+) or PAX8 mAb (−) and unilateral ≥10 cm=primary, PAX8 mAb (−) and bilateral tumours of any size or PAX8 mAb (−) and unilateral tumour &lt;10 cm=metastatic</td>
<td>43</td>
<td>13</td>
<td>91.5</td>
<td>72.2</td>
<td>89.6</td>
<td>87</td>
<td>86.2</td>
</tr>
</tbody>
</table>

eMOMCs, extragenital metastatic mucinous carcinomas involving the ovary; mAb, monoclonal antibody; MOMCs, metastatic mucinous carcinomas involving the ovary; NPV, negative predictive value; POMTs, primary ovarian mucinous tumours; PPV, positive predictive value.
low PAX8 expression compared with ovarian non-mucinous carcinomas. Moreover, PAX8 (mAb) should be considered as having limited reliability in distinguishing POMTs from eMOMCs due to its good specificity but poor sensitivity.

Unfortunately, in our cohort, the algorithm proposed by Seidman et al.® correctly classified only 75.4% of mucinous tumours involving the ovary (76.6% of POMTs and 72.2% of eMOMCs). Our ROC curve analysis supported the fact that PAX8 (mAb) IHC status, tumour size and laterality were useful in differentiating POMTs and eMOMCs, even though their individual accuracy was disappointing. The combination of these three parameters correctly classified 86.2% of ovarian mucinous tumours, including 91.5% of POMTs and 72.2% of eMOMCs, with the highest Youden Index (63.7%).

In conclusion, our study has verified that PAX8 (mAb) is a highly specific marker and, when combined with tumour size and laterality, can significantly improve the accuracy of discriminating POMTs from eMOMCs. Although several other candidate molecular markers are available for this differential diagnosis, few are favourably specific and commercially available, thus greatly limiting their application.® We propose a simple, convenient and high performance to price ratio algorithm combining PAX8 (mAb) immunostaining with tumour size and laterality. Although it should be validated in further studies, our algorithm may have a role in improving the diagnostic criteria for distinguishing between primary and metastatic mucinous ovarian carcinomas.

Take home messages

- Anti-PAX8 rabbit monoclonal antibody (mAb) demonstrated superior specificity than anti-PAX8 rabbit polyclonal antibody by immunohistochemistry.
- PAX8 (mAb), laterality and tumour size were useful in differentiating primary from extragenital metastatic mucinous ovarian tumours.
- Combining PAX8 (mAb) immunostaining status with tumour size and laterality is strongly recommended as an effective algorithm in routine practice.

Table 3  Previously published results of PAX8 expression in ovary mucinous tumours (positive/total cases)

<table>
<thead>
<tr>
<th>Previous studies</th>
<th>Antibodies</th>
<th>Ovary benign mucinous tumours</th>
<th>Ovary benign/borderline mucinous tumours</th>
<th>Ovary mucinous carcinomas</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bowen et al.®</td>
<td>α-mPax8-bill,pAb*</td>
<td>ND</td>
<td>ND</td>
<td>3/18</td>
</tr>
<tr>
<td>Nonaka et al.®</td>
<td>RpAbt</td>
<td>ND</td>
<td>ND</td>
<td>1/12</td>
</tr>
<tr>
<td>Köbel et al.®</td>
<td>α-mPax8-bill,pAb*</td>
<td>ND</td>
<td>ND</td>
<td>2/31</td>
</tr>
<tr>
<td>Tabrizi et al.®</td>
<td>pAb*</td>
<td>ND</td>
<td>ND</td>
<td>0/30</td>
</tr>
<tr>
<td>Laury et al.®</td>
<td>RpAbt</td>
<td>ND</td>
<td>ND</td>
<td>10/25</td>
</tr>
<tr>
<td>Tong et al.®</td>
<td>RpAbt</td>
<td>ND</td>
<td>ND</td>
<td>0/2</td>
</tr>
<tr>
<td>Tacha et al.®</td>
<td>RpAbt</td>
<td>0/1</td>
<td>ND</td>
<td>27/54</td>
</tr>
<tr>
<td>Ozcan et al.®</td>
<td>RpAbt</td>
<td>ND</td>
<td>6/31</td>
<td>2/9</td>
</tr>
<tr>
<td>Chu et al.®</td>
<td>RpAb§</td>
<td>ND</td>
<td>ND</td>
<td>9/19</td>
</tr>
<tr>
<td>Liliac et al.®</td>
<td>RpAbt</td>
<td>ND</td>
<td>ND</td>
<td>4/13</td>
</tr>
<tr>
<td>Total (%)</td>
<td>0/1(0)</td>
<td>6/31 (19.4)</td>
<td>57/213 (26.76)</td>
<td></td>
</tr>
</tbody>
</table>

*Non-commercial.
†Proteintech Group Inc.
§Cell Marque.
ND, not done; pAb, polyclonal antibody; RpAb, recombinant polyclonal antibody.

Handling editor Runjan Chetty

Contributors AH, HL and LZ performed the experiments. AH, HL and YL collected the clinical samples. AH, CR and YW analysed the data. AH and CL wrote the paper. CL designed the project.

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Competing interests None.

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Provenance and peer review Not commissioned; externally peer reviewed.

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