α-Methylacyl-CoA racemase (AMACR) serves as a prognostic biomarker for the early recurrence/metastasis of HCC

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ABSTRACT

Aims Hepatocellular carcinoma (HCC) is one of the most common malignancies worldwide, and it is still lacking effective prognostic biomarkers so far. Previous results of the iTRAQ-based quantitative proteomics study (iTRAQ-2DLC-MS/MS) have shown that α-methylacyl-CoA racemase (AMACR) might be a promising prognostic biomarker for the early recurrence/metastasis of hepatocellular carcinoma (HCC). Here a large-scale cohort clinical study was performed to evaluate its prognostic potential.

Methods HCC samples from patients (n=158) were used for the construction of tissue microarray. The expression level of AMACR was determined by immunohistochemical staining. A large-scale cohort clinical study between the expression of AMACR and some major clinical parameter has been performed to assess the prognostic potential of AMACR for the early recurrence/metastasis of HCC.

Results Some important clinical parameters such as α-fetoprotein, tumour numbers, dissemination to regional lymph nodes, tumour capsule and portal vein tumour thrombosis are significantly associated with the low expression of AMACR. The expression of AMACR was an independent factor for the survival of patients with HCC. The median survival time was 17 months in the low-expression group compared with 45 months in the high-expression group.

Conclusions This study reveals that the AMACR might be a potential prognostic marker for predicting early recurrence/metastasis of HCC after hepatectomy.

INTRODUCTION

Hepatocellular carcinoma (HCC) is one of the most common malignancies worldwide and is reported to be the second leading cause of cancer death in China.1 Despite the proved feasibility and safety of surgical resection for HCC,2 3 the prognosis of patients with HCC remains poor, mainly due to the intraparenchymal recurrence and/or extrahepatic metastasis. In such cases, it is necessary to identify new biomarkers for distinguishing, predicting and treating the early and late recurrence/metastasis, which will certainly give important guides for identifying patients with HCC with high risk of early recurrence/metastasis.

α-Methylacyl-CoA racemase (AMACR) is a peroxisomal and mitochondrial enzyme that plays an important role in bile acid biosynthesis and β-oxidation of branched-chain fatty acids through the interconversion of (R)- and (S)-2-methyl branched-chain fatty acyl-CoA fragments.4 It is abundantly expressed in prostate cancer compared with the benign prostate epithelium and serves as a diagnostic biomarker.3–8 There are also some reports regarding the correlations between the expression of AMACR and the diagnostic potential for HCC. Guzman et al9 have reported that AMACR could distinguish the HCC and the dysplastic hepatocytes from the benign non-dysplastic hepatocytes based on the different staining patterns, but the sensitivity was not mentioned in this report; Li et al10 have reported that AMACR could serve as a useful marker for distinguishing the well-differentiated HCC from the hepatocellular adenoma, and the AMACR might play a role in HCC development and progression. However, there has been conflicting reports in other studies where it was found that AMACR may not be a suitable marker; Willemoe et al11 have reported that the staining patterns and staining intensities of AMACR in HCC tumour cells were not significantly different from the surrounding non-tumour cells, and the AMACR staining could not distinguish the neoplastic from non-neoplastic liver cells.

In our previous quantitative proteomics study (iTRAQ-2DLC-MS/MS),12 the results have shown that AMACR might be a very promising prognostic biomarker for the early recurrence/metastasis of HCC instead of a diagnostic biomarker. Here, a large-scale cohort clinical study between the expression of AMACR and some major clinical parameter has been performed to assess the prognostic potential of AMACR for predicting the early recurrence/metastasis of HCC.

MATERIALS AND METHODS

Sample collection
One hundred and fifty-eight formalin-fixed and paraffin-embedded HCC tissues from consecutive patients who underwent curative resection between 2002 and 2010 at the First Affiliated Hospital of Fujian Medical University were retrieved for immunohistochemical staining. Barcelona Clinic Liver Cancer (BCLC) of the evaluated patients were 0 (6, 3.8%), A (22, 13.9%), B (130, 82.3%). The tissues were divided into three groups according to the time of recurrence/metastasis after operation: the patients who had recurrence/metastasis within 12 months after operation (RM ≤12 months group, n=101), and the BCLC of the evaluated patients.
were A (7), B (94); the patients whose recurrence/metastasis occurred between 12 and 24 months after operation (R/M12-24 months group, n=21), and the BCLC of the evaluated patients were A (5), B (16); the patients who had no recurrence/metastasis at least 24 months after surgery (NR/M group, n=36), and the BCLC of the evaluated patients were 0 (6), A (10), B (20). Fresh tissues were collected at the time of surgery from patients and were formalin embedded for immunohistochemistry. The project was approved for the using of human biopsy by the Institution Review Board of the First Affiliated Hospital of Fujian Medical University. The written consent was received from all participants before surgery. Clinical and pathological diagnosis of patients with HCC met the diagnostic criteria of the American Association for the Study of Liver Diseases.

Construction of the tissue microarrays and immunohistochemistry

HCC samples from patients (n=158) were used for the construction of tissue microarray (TMA). By using manual tissue arrayer, tissue cores from circled areas were targeted and then transferred to the corresponding location of the recipient block to form a TMA block. Two replicated tissue points were sampled from each patient with HCC. Then the wax block was cooled at room temperature for 30 min and placed in 4°C for further usage. The cooled wax block was fixed on the auto tissue slicer and continuously sliced into 50 sections. Each section was 4 μm. Afterwards, the sections were extended in distilled water for 2 min, then directly mounted onto the superforst plus-treated import slide and dried on the section drier for 16 h at 60°C.

Immunohistochemical staining of AMACR was performed by using the two-step Envision plus staining technique. Representative 4 μm section TMA was de-paraffinised in xylene and rehydrated in graded ethanol. Then the slides were incubated in 1% hydrogen peroxide for 30 min to block endogenous peroxidase activity and then rehydrated in distilled water followed by phosphate buffered saline (PBS). Slides were incubated in 0.1 mol/L EDTA (pH 9.0) with 3 min for high-pressure antigen retrieval. Afterwards, the liver sections were incubated with the primary antibody (mouse polyclonal antihuman AMACR antibody, 1:100 dilutions, abcam) at 4°C overnight. After several rinses with PBS, it was followed by incubating with HRP-conjugated secondary antibodies (ZSGB-BIO, China) at room temperature for another 1 h. Finally, the liver sections were subjected to diaminobenzidine colouration and haematoxylin restaining. The results were independently assessed by two pathologists double-blindly. Using localised prostate cancer as the positive control (scored 3), all tissues were manually scored as 0 (negative), 1 (weak), 2 (strong) or 3 (very strong). The expression was finally considered by low expression if the final score was 0–1, while high expression if the final score was 2–3.

Statistical analysis

All statistical analyses were performed with SPSS software package (V.19.0). Comparisons of quantitative data between two groups were performed with a two-tailed paired Student’s t test. Categorical data were analysed by Fisher’s exact test between

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**Figure 1** Different α-methylacyl-CoA racemase (AMACR) expression levels in hepatocellular carcinoma tumour tissues. As described in ‘Methods’ section, AMACR were semiquantitatively categorised into four groups: (A) negative (0), (B) weak (1+), (C) moderate (2+) and (D) strong (3+). (original magnifications 200×).
AMACR expression and clinical–pathological features. Survival curves were calculated by the Kaplan–Meier method and compared using the log-rank test. A Cox proportional hazards model was used to determine the independent factors of survival and recurrence based on the variables selected in univariate analysis. p<0.05 was considered as statistically significant.

RESULTS
In our previous proteomics study, AMACR was identified to be significantly downregulated in the HCC tumour with early recurrence/metastasis. Since this protein has clear lower expression profiles in the early recurrence/metastasis patients, it might be interesting potential prognostic biomarker for the predicting of early recurrences/metastases in HCC.

To further confirm the expression profile of AMACR in patients with HCC, we constructed the TMAs and performed the IHC staining for AMACR in a large scale. After independently evaluated by two pathologists, the expression level of AMACR was scored as 0 (negative), 1 (weak), 2 (strong) or 3 (very strong). The representative images of each score are shown in figure 1. Depending on the scores, we divided the AMACR expression level into two groups: low-expression group if the final score was 0 or 1, and high-expression group if the final score was 2 or 3. Combining with the clinical history, the major factors that affect the prognosis have been collected to evaluate the relationship between the AMACR expression and the key clinical parameters.

As shown in tables 1 and 2, the low expression of AMACR in HCC tissues was significantly associated with multiple tumour numbers, absence of tumour capsule, undistinguishable tumour boundaries and presence of portal vein tumour thrombosis (PVTT). Larger tumour size and faster progression of cirrhosis were also more frequently observed in the AMACR low-expression group, although not statistically significantly. Tumour without encapsulation is more likely to be associated with microscopic vascular invasion and leads to a further intrahepatic recurrence or metastasis. Meanwhile, it has been reported that tumour recurrence commonly occurs in HCC patients with PVTT, which is the major cause of unsatisfactory prognosis. As shown in table 2, 79% (33/42) of total cases showed AMACR low expression in the PVTT group, and all the cases in the regional lymph nodes dissemination group had a lower AMACR expression. These data suggest that the lower expression of AMACR is well associated with the more invasive features of HCC.
The associations between the expression level of AMACR and postoperative clinical features have been further studied and shown in Table 3. The AMACR expression has been assessed in 158 patients, including 101 cases of early recurrence/metastasis (R/M ≤ 12 months), 21 cases of late recurrence/metastasis (R/M > 12–24 months) and 36 cases of none-recurrence/metastasis (NR/M). As shown in Table 4, the low expression of AMACR was detected in 12 cases of NR/M HCC patients (in a total of 36 patients, the low-expression rate was 33.3%). In contrast, the AMACR low-expression rate was significantly higher in the early recurrence/metastasis group (R/M ≤ 12 months, 71 of 101, low-expression rate 70.3%, p<0.001). Meanwhile, the patients with low expression of AMACR had very discouraging prognosis, only with a median survival of 17 months; however, the patients with high expression of AMACR had a better prognosis, with a median survival of 45 months (Figure 2).

The univariate and multivariate analysis was further performed to evaluate the prognosis potential of AMACR, as shown in Table 3; low expression of AMACR was the independent risk factor affecting the survival of patients with HCC. These data clearly suggest that AMACR could serve as an independent prognostic factor for the early recurrence/metastasis of HCC.

**DISCUSSION**

Following curative liver resection of HCC within the Milan criteria, still up to 63% of patients will develop intrahepatic recurrence or extrahepatic metastasis within 5 years, and most of the recurrence/metastasis will occur in the early stage after the surgery. The incidence of recurrence/metastasis within 12 months in the residual liver of patients with HCC, who underwent curative resection, ranges from 50% to 70%, and the median survival after recurrence/metastasis is only 13 months. There are several biomarkers for predicting the recurrence/metastasis of HCC, but still limited biomarkers point to the early stage. Here, the prognostic potential of AMACR for the prediction of early recurrence/metastasis postliver resection in HCC has been systematically analysed.

Our previous studies have shown that the AMACR was significantly downregulated in the early recurrence/metastasis group compared with the NR/M group. These results encouraged us to further verify the expression profiles of AMACR in patients with HCC in a large scale; therefore, we performed the TMA with immunohistochemistry (IHC) staining to evaluate the AMACR expression in patients.

Combining with the clinical assessment, several important tumour features have been analysed. Among those features, the preoperative α-fetoprotein level, tumour numbers, dissemination to regional lymph nodes, tumour capsule and PVTT are significantly correlated with the low expression of AMACR. Additionally, patients with HCC with lower AMACR expression are significantly associated with a poorer 5-year survival with a median survival of 17 months, but the patients with HCC with higher AMACR expression had a better survival with a median survival of 45 months. Furthermore, Cox’s multivariate proportional hazards model revealed that the AMACR expression level can serve as an independent prognostic biomarker to identify patients with poor clinical outcomes. Taken together, these data suggested that AMACR can be applied as a potential prognosis biomarker for the recurrence/metastasis of HCC in patients. Further validation of the AMACR expression by RT-PCR on mRNA level is not necessary for the clinical practice, but it would certainly promote the clinical applications of AMACR due to the easier use of the RT-PCR technique. As a promising prognosis biomarker, it would be crucial to detect in the circulation since the biopsies are not always available in clinical practice; we have some unpublished preliminary data showing that the AMACR could be detected in HCC patient’s serum, but further study is needed to clarify whether the AMACR serum levels are associated with the prognosis of HCC, as well as its sensitivity and specificity.

It has been reported that lower AMACR expression was also associated with worse outcome in the prostate cancer. Meanwhile, the AMACR downregulation in the prostate carcinoma metastases has also been identified; similar findings also have been reported in colorectal carcinoma and in adenocarcinomas of the small and large intestines as well. However, on the other hand, the AMACR overexpression has also been identified in various cancers. As described previously, the AMACR regulates the entry of branched-chain lipids into the peroxisomal and mitochondrial β-oxidation pathways, and then regulates the production of reactive oxygen species, which could result in DNA damage due to oxidative stress. It seems that the upregulation of AMACR may be involved in the early tumourigenesis, but the downregulation might relate to the development of tumours with high capacity of recurrence/metastasis. Although

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**Table 4** The expression of α-methylacyl-CoA racemase (AMACR) in different groups according to the time of recurrence/metastasis postoperation

<table>
<thead>
<tr>
<th>Groups</th>
<th>Low expression (%)</th>
<th>P Values</th>
</tr>
</thead>
<tbody>
<tr>
<td>R/M group ≤ 12 months</td>
<td>71</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>R/M group &gt; 12–24 months</td>
<td>9</td>
<td>0.1</td>
</tr>
<tr>
<td>NR/M group</td>
<td>12</td>
<td></td>
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</tbody>
</table>

*P<0.01.

**Figure 2** Kaplan–Meier’s analysis of the correlation between the α-methylacyl-CoA racemase (AMACR) expression and the survival of hepatocellular carcinoma (HCC) patients. Patients with HCC with low-expression level of AMACR (n=95) had significantly lower survival rate than those with high-expression level of AMACR (n=63), (p<0.001).
there are some mechanistic studies on AMACR,29 30 the underlying molecular mechanisms that explain the aggressive behaviour and early recurrence/metastasis feature of HCC with lower AMACR expression are still unclear. Further prospective studies with molecular mechanism are necessary to fully clarify the clinical implications of AMACR in HCC.

In conclusion, we applied TMA-based immunohistochemistry staining to perform a large-scale cohort clinical study and showed that the low expression of AMACR was significantly associated with key clinical features and hence leads to the early recurrence/metastasis of HCC. Our findings indicate that the AMACR might be a potential prognostic biomarker for predicting the early recurrences/metastases of HCC after hepatectomy.

### Take home messages

- α-Methylacyl-CoA racemase (AMACR) is downregulated in the early recurrence/metastasis hepatocellular carcinoma (HCC) tumour tissues.
- The downregulation of AMACR is significantly associated with poor prognosis of patients with HCC.
- AMACR might be a promising prognostic biomarker for patients with HCC instead of a diagnostic biomarker.

### Contributors
BX, YZ, ZC and LC performed the experiments; BX and XD collected the clinical samples; BX and XL wrote the paper; XL, AH and JL designed the project.

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### REFERENCES


### Table 5

Univariate and multivariate analysis of α-methylacyl-CoA racemase (AMACR) and other clinicopathological characteristics at 5 years of follow-up

<table>
<thead>
<tr>
<th>Variables</th>
<th>Survival</th>
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<th></th>
<th>p Value</th>
<th></th>
<th></th>
<th>p Value</th>
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<tbody>
<tr>
<td></td>
<td>Univariate analysis</td>
<td>Multivariate analysis</td>
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<tr>
<td></td>
<td>HR</td>
<td>95% CI</td>
<td>p Value</td>
<td>HR</td>
<td>95% CI</td>
<td>p Value</td>
<td></td>
</tr>
<tr>
<td>Age (&lt;55/ ≥55)</td>
<td>0.743</td>
<td>0.488 to 1.132</td>
<td>0.167</td>
<td>2.620</td>
<td>1.200 to 5.722</td>
<td>0.016</td>
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<tr>
<td>Sex (male/female)</td>
<td>2.865</td>
<td>1.327 to 6.185</td>
<td>0.007</td>
<td>1.054</td>
<td>0.770 to 1.442</td>
<td>0.744</td>
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<td>HBV infection (absent/present)</td>
<td>1.411</td>
<td>0.929 to 2.142</td>
<td>0.106</td>
<td>1.167</td>
<td>0.781 to 1.745</td>
<td>0.451</td>
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</tr>
<tr>
<td>Cirrhosis (absent/present)</td>
<td>1.054</td>
<td>0.770 to 1.442</td>
<td>0.744</td>
<td>0.765</td>
<td>0.386 to 1.519</td>
<td>0.445</td>
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<tr>
<td>Alanine transaminase (ALT) (UI/L) (&lt;54/≥54)</td>
<td>1.167</td>
<td>0.781 to 1.745</td>
<td>0.451</td>
<td>1.634</td>
<td>0.991 to 2.696</td>
<td>0.054</td>
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<tr>
<td>Total bilirubin (μmol/L) (&lt;25/≥25)</td>
<td>0.985</td>
<td>0.626 to 1.549</td>
<td>0.946</td>
<td>2.280</td>
<td>0.993 to 5.234</td>
<td>0.052</td>
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<tr>
<td>GGTP (UI/L) (&lt;60/≥60)</td>
<td>1.417</td>
<td>0.737 to 2.723</td>
<td>0.296</td>
<td>1.628</td>
<td>1.050 to 2.524</td>
<td>0.030</td>
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<tr>
<td>Prothrombin time (s) (≤14/&gt;14)</td>
<td>0.851</td>
<td>0.505 to 1.434</td>
<td>0.544</td>
<td>1.833</td>
<td>1.203 to 2.795</td>
<td>0.005</td>
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</tr>
<tr>
<td>Tumour encapsulation (absent/present)</td>
<td>1.833</td>
<td>1.203 to 2.795</td>
<td>0.005</td>
<td>0.258</td>
<td>0.105 to 0.635</td>
<td>0.003</td>
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<tr>
<td>Tumour differentiation (I/II–III)</td>
<td>0.258</td>
<td>0.105 to 0.635</td>
<td>0.003</td>
<td>0.418</td>
<td>0.167 to 1.050</td>
<td>0.063</td>
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<tr>
<td>Extrahepatic metastasis (yes/no)</td>
<td>0.985</td>
<td>0.626 to 1.549</td>
<td>0.946</td>
<td>0.851</td>
<td>0.505 to 1.434</td>
<td>0.544</td>
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<tr>
<td>Intraoperative ascites (absent/present)</td>
<td>1.628</td>
<td>1.050 to 2.524</td>
<td>0.030</td>
<td>1.251</td>
<td>0.776 to 2.015</td>
<td>0.358</td>
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<tr>
<td>Tumour number (single/multiple)</td>
<td>0.992</td>
<td>0.530 to 1.857</td>
<td>0.980</td>
<td>0.992</td>
<td>0.530 to 1.857</td>
<td>0.980</td>
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<tr>
<td>Maximal tumour size(&lt;5/&gt;5 cm)</td>
<td>4.439</td>
<td>1.084 to 10.925</td>
<td>0.001</td>
<td>2.368</td>
<td>0.931 to 6.026</td>
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<tr>
<td>PVTT (absent/present)</td>
<td>1.833</td>
<td>1.203 to 2.795</td>
<td>0.005</td>
<td>1.301</td>
<td>0.822 to 2.058</td>
<td>0.262</td>
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<tr>
<td>AMACR (low expression/high expression)</td>
<td>0.501</td>
<td>0.326 to 0.770</td>
<td>0.002</td>
<td>0.612</td>
<td>0.319 to 0.957</td>
<td>0.031</td>
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</table>

Prognostic significance of AMACR: univariable analysis revealed that presence of intraoperative ascites and PVTT, maximal tumour size ≥5 cm, absence of tumour encapsulation, poor differentiation, low expression of AMACR were significantly associated with poor prognosis. On multivariate analysis of the prognostic factors for overall survival, presence of PVTT (relative risk (RR)=1.801; 95% CI 1.169 to 2.773; p=0.008) and low expression of AMACR (RR=0.612; 95% CI 0.319 to 0.957; p=0.031) were independent predictors for poor overall survival of hepatocellular carcinoma patients.

GGTP, glutamyltranspeptidase; HBV, Hepatitis B virus; PVTT, portal vein tumour thrombosis.

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