Activity of ampicillin *in vitro* compared with other antibiotics

R. SUTHERLAND AND G. N. ROLINSON

*From the Beecham Research Laboratories Limited,
Research Division, Betchworth, Surrey*

SYNOPSIS Comparative tests *in vitro* for antibacterial activity were carried out with ampicillin, tetracycline, and chloramphenicol using 673 clinical isolates of Gram-negative bacilli and *Streptococcus faecalis*. Further comparative tests were also carried out with ampicillin, chloramphenicol, colistin sulphate, colistin methane sulphonate, cycloserine, kanamycin, nitrofurantoin, polymyxin, streptomycin, and tetracycline, using groups of 20 strains of each of the main species selected at random from the total number of isolates. Of the total number of isolates a higher percentage was inhibited by ampicillin than by tetracycline or chloramphenicol. Ampicillin showed particularly high activity against certain species of bacteria and displayed an antibacterial spectrum not shown by any of the other antibiotics tested.

Ampicillin(6[D(-)- α -aminophenylacetamido]penicillanic acid) is a penicillin with broad-spectrum activity (Rolinson and Stevens, 1961). The compound is acid stable and gives satisfactory serum concentrations after oral administration (Knudsen, Rolinson, and Stevens, 1961). Ampicillin shows a very low order of toxicity (Acred, Brown, Turner, and Wilson, 1962) and the early clinical experience with ampicillin was reported by Stewart, Coles, Nixon, and Holt (1961); Bunn (1961); Rutenberg, Greenberg, Schweinburg, and Perreault (1961); Brumfitt, Percival, and Carter (1962); and Trafford, Maclaren, Lillicrap, Barnes, Houston, and Knox (1962).

The antibacterial spectrum of ampicillin includes the pyogenic cocci, enterococci, *Neisseria* species including the gonococcus, *Haemophilus influenzae*, *Salmonella* species, and strains of *Escherichia coli*, *Shigella* and *Proteus*. Not all strains of these organisms are uniformly sensitive to ampicillin, however, and conflicting statements regarding the activity of this penicillin have appeared in the literature, particularly against coliforms and *Proteus* species.

METHODS

A total of 673 strains of bacteria was obtained from the Middlesex Hospital, the West Middlesex Hospital, and

St. Thomas's Hospital, London. Nearly all the cultures had been associated with urinary tract infections and were received as isolated routinely without selection of any kind.

CLASSIFICATION The cultures were supplied on agar slopes and were cultured on MacConkey's agar, from which mixed cultures were separated into pure cultures.

The *E. coli* group was defined as lactose-fermenting, non-citrate utilizing bacteria; this group included indole-positive (type I) and indole-negative (intermediate coliforms) organisms. Lactose-fermenting, citrate-utilizing, indole-negative bacteria were classified as *Aerobacter aerogenes*.

Proteus species were identified with some considerable care in view of the wide variation in sensitivity of these organisms to ampicillin, and classification was based on the biochemical tests as reported by Mackie and McCartney (1962). The majority of *Proteus* strains encountered in this study belonged to the species *Proteus mirabilis*; other *Proteus* species were rare.

The remainder of the non-lactose fermenting, non-urea-hydrolysing organisms were tested for phenylpyruvic acid production by the method of Stewart (1961) in order to isolate providencia (*Proteus inconstans*) organisms; these strains were also tested for β -(D)-galactosidase production (Lowe, 1962) to isolate late-lactose fermenting cultures which would be classified as paracolon bacteria. *Pseudomonas* was distinguished by the characteristic odour and production of green pigment on MacConkey's medium, although in some instances weak reactions were confirmed by culture in Hugh and Leifson's medium, and by tests to demonstrate utilization of arginine (Thornley, 1960).

Received for publication January 10 1964.

Streptococcus faecalis was identified by colonial appearance on MacConkey's agar and Gram-stain reaction.

ANTIBIOTIC SENSITIVITIES Minimum inhibitory concentrations were determined by serial dilution in agar. A loop of an overnight broth culture was used as inoculum and the plates were incubated at 37°C. overnight.

RESULTS

COMPARATIVE ACTIVITIES OF AMPICILLIN, CHLORAMPHENICOL, AND TETRACYCLINE Of the 673 isolates tested for sensitivity to ampicillin, chloramphenicol, and tetracycline, the predominant organism was *E. coli*. This accounted for 40% of the total number. *Proteus mirabilis* was the next most common pathogen (24%); other species of *Proteus*, namely *P. vulgaris* and *P. morganii*, comprised only 2% of the total. The remainder was accounted for by *Aerobacter aerogenes* (12%), *Streptococcus faecalis* (10%), *Pseudomonas pyocyanea* (7%), and miscellaneous organisms, comprising mainly paracolon and some achromobacter bacteria, (6%). The antibiotic sensitivities of the bacteria are shown in Table I. Of the total of 269 strains of *E. coli*, 78% were inhibited by 5 µg./ml. or less of ampicillin. The corresponding figures for chloramphenicol and tetracycline were 52% and 64%, respectively. At

this concentration, therefore, ampicillin was more effective than either chloramphenicol or tetracycline. At the lower concentration of 2.5 µg./ml., however, only 19% were inhibited by ampicillin whereas with tetracycline the corresponding figure was 45% and for chloramphenicol only 8%.

Resistance of *E. coli* to ampicillin was not associated with resistance to tetracycline or chloramphenicol. The strains resistant to tetracycline or chloramphenicol were as sensitive to ampicillin as was the total number of isolates tested.

The strains of *P. mirabilis* were generally highly sensitive to ampicillin, 83% being inhibited by a concentration of 5.0 µg./ml. and 74% by 2.5 µg./ml. Both chloramphenicol and tetracycline were considerably less active at these concentrations. In contrast with *P. mirabilis*, all the strains of *P. morganii* and *P. vulgaris* were relatively resistant to ampicillin.

Ampicillin was highly active against the isolates of *Streptococcus faecalis*, 95% being inhibited by 2.5 µg./ml. whereas chloramphenicol and tetracycline were relatively ineffective. Against strains of *Aerobacter aerogenes*, ampicillin generally showed a low order of activity. Of a total of 80 strains, however, 20% were inhibited by 5.0 µg./ml. or less of ampicillin. Similarly, not all paracolon organisms were resistant to ampicillin, about half of the strains tested being inhibited by 5.0 µg./ml. ampicillin.

TABLE I

ACTIVITY OF AMPICILLIN, CHLORAMPHENICOL, AND TETRACYCLINE AGAINST 673 STRAINS OF GRAM-NEGATIVE BACTERIA AND ENTEROCOCCI

Organism	Antibiotic	Number of Strains	Percentage of Total	Minimum Inhibitory Concentrations (µg./ml.) and No. of Strains										
				>250	250	125	50	25	12.5	5.0	2.5	1.25	0.5	0.25
<i>E. coli</i>	Ampicillin	269		8	2	4	3	1	39	161	46	5		
	Chloramphenicol	270	40.0	15	3	2	4	6	101	116	12	9	2	
	Tetracycline	259		11	20	13	11	23	13	50	59	54	3	2
<i>Proteus mirabilis</i>	Ampicillin	156		11		3	2	5	5	14	80	36		
	Chloramphenicol	156	23.2	1	3	6	5	6	89	41	4	1		
	Tetracycline	156		16	3	17	98	14	1	1	4	2		
<i>Streptococcus faecalis</i>	Ampicillin	67		1		1			1		21	37	3	3
	Chloramphenicol	66	10.0			3	8	2	34	12	6	1		
	Tetracycline	67		1	10	17	3	4	3	5	7	13	4	
<i>Aerobacter aerogenes</i>	Ampicillin	80		25	7	5	13	5	9	11	2	3		
	Chloramphenicol	80	11.9	10	3	3	4	2	18	26	5	8	1	
	Tetracycline	80		5	8	6	6	15	5	6	9	17	2	1
<i>Pseudomonas pyocyanea</i>	Ampicillin	47		40	4		2			1				
	Chloramphenicol	47	7.0	4	4	25	10	1	2	1				
	Tetracycline	47		15	9	6	9	7			1			
Miscellaneous ¹	Ampicillin	40		5	3	3	1	4	5	12	6	1		
	Chloramphenicol	40	6.0	1	1	2	1	3	15	12	5			
	Tetracycline	40		2	6	1	3	2	1	12	8	5		
<i>Proteus</i> species ²	Ampicillin	13		5	2	2	2		1	1				
	Chloramphenicol	13	1.9		2	2	1		3	6	1			
	Tetracycline	13		3	2	1	1		4			1	1	

¹Mainly paracolon organisms but including achromobacter sp. and *Proteus inconstans*.

²*Proteus morganii*, 10 strains; *Proteus vulgaris*, three strains.

TABLE II

COMPARISON OF AMPICILLIN WITH NINE ANTIBIOTICS AGAINST GRAM-NEGATIVE BACTERIA AND ENTEROCOCCI

Organism	Number of Strains	Antibiotic	Minimum Inhibitory Concentrations ($\mu\text{g./ml.}$) and No. of Strains										
			>500	500	250	125	50	25	12.5	5.0	2.5	1.25	0.5
<i>E. coli</i>	20	Ampicillin	2				1		2	9	6		
		Chloramphenicol		2			1		4	13			
		Colistin sulphate	1			1				3	15		
		Colistin methane sulphonate	1			1			17	1			
		Cycloserine				4	12	4					
		Kanamycin						1		3		16	
		Nitrofurantoin				3	3	2	10	2			
		Polymyxin	1							2	10	7	
		Streptomycin	2					1		1	6	9	1
		Tetracycline		1	1	2					14	2	
<i>Proteus mirabilis</i>	20	Ampicillin	1								2	15	2
		Chloramphenicol						2	16	2			
		Colistin sulphate	19							1			
		Colistin methane sulphonate	18	1				1					
		Cycloserine			1	19							
		Kanamycin							6	14			
		Nitrofurantoin				8	12						
		Polymyxin	19						1				
		Streptomycin	1							8			
		Tetracycline		2	18								
<i>Streptococcus faecalis</i>	20	Ampicillin										20	
		Chloramphenicol				5	1	3	11				
		Colistin sulphate	20										
		Colistin methane sulphonate	20										
		Cycloserine			2	18							
		Kanamycin						5	15				
		Nitrofurantoin				1		19					
		Polymyxin	5	15									
		Streptomycin	10				8	2					
		Tetracycline			11	1	1			1	6		
<i>Aerobacter aerogenes</i>	10	Ampicillin	2	2	1		3	1	1				
		Chloramphenicol	1						2	3	2	2	
		Colistin sulphate								4	4	2	
		Colistin methane sulphonate						2	8				
		Cycloserine			2	6	2						
		Kanamycin						1	1		2	6	
		Nitrofurantoin				5	2	2	1				
		Polymyxin								6	4		
		Streptomycin	1	1							1	7	
		Tetracycline			2				1	3	1	3	
<i>Pseudomonas pyocyanea</i>	20	Ampicillin	19							1			
		Chloramphenicol			1	18			1				
		Colistin sulphate								19		1	
		Colistin methane sulphonate					18	1	1				
		Cycloserine		1	11	8							
		Kanamycin			10	6	4						
		Nitrofurantoin			18	2							
		Polymyxin								19	1		
		Streptomycin	7			2	2	8	1				
		Tetracycline				15	4				1		
Mixed <i>proteus</i> ¹	9	Ampicillin	1		1		1		2		2	2	
		Chloramphenicol		1		1			4	3			
		Colistin sulphate	6						1	2			
		Colistin methane sulphonate	5	1				2	1				
		Cycloserine	1			8							
		Kanamycin								2	3	4	
		Nitrofurantoin				6	2	1					
		Polymyxin	7							1	1		
		Streptomycin	3							3	1	2	
		Tetracycline			1	1	3	1	1			2	

¹Three of *P. morganii*, one of *P. inconstans*, four of *P. vulgaris*, and one of *P. rettgeri*.

With the strains of *Pseudomonas pyocyanea*, ampicillin, and also chloramphenicol and tetracycline, showed a very low order of activity.

ACTIVITY OF AMPICILLIN AND NINE OTHER ANTIBIOTICS

Groups of 20 strains of each of the main species were selected at random from the total number of isolates and tested for antibiotic sensitivity to ampicillin, chloramphenicol, colistin sulphate, colistin methane sulphonate, cycloserine, kanamycin, nitrofurantoin, polymyxin, streptomycin and tetracycline. The minimum inhibitory concentration values are shown in Table II. Against the group of *E. coli*, colistin sulphate, kanamycin, polymyxin, streptomycin and tetracycline were all more active than ampicillin. Colistin methane sulphonate, however, was less active and cycloserine and nitrofurantoin were relatively ineffective.

Against *P. mirabilis*, ampicillin was markedly more active than any of the other antibiotics tested. *Proteus mirabilis* was tested as a separate group because this was the species of proteus most frequently encountered clinically. Results for a mixed group of nine proteus cultures comprising *P. morganii*, *P. inconstans*, *P. vulgaris*, and *P. rettgeri* are also shown in Table II. The minimum inhibitory concentration values were generally very variable for most of the antibiotics tested. At a level of 5.0 µg./ml. only two strains were inhibited by tetracycline and three by chloramphenicol. Four strains were inhibited by ampicillin at this level. Kanamycin and streptomycin were the most effective of the antibiotics tested against this group of organisms.

Ampicillin was outstandingly active against *Streptococcus faecalis* in comparison with the other antibiotics. Ampicillin, however, showed little activity against 10 strains of *Aerobacter aerogenes*, and chloramphenicol, colistin sulphate, kanamycin, polymyxin, and streptomycin all showed greater activity although resistant strains were observed with most of these antibiotics. Against *Pseudomonas pyocyanea* only polymyxin and colistin sulphate were active at concentrations of 5 µg./ml. Colistin methane sulphonate showed a considerably lower order of activity against this organism and none of the other antibiotics demonstrated significant activity.

DISCUSSION

The results obtained in this report with a fairly large number of clinical isolates from three different hospitals give some indication of the general sensitivity of Gram-negative bacilli and enterococci

to ampicillin. Some comparisons can also be drawn between the activity of ampicillin and that of other antibiotics in current use.

Taking the group of 673 isolates tested as a whole, 448 (67%) were inhibited by ampicillin at a concentration of 5 µg./ml. The corresponding figures for chloramphenicol and tetracycline were 42% and 38%, respectively. Among the coliforms, strains of *E. coli* were fairly sensitive to ampicillin whereas strains of aerobacter were mostly resistant. Similarly, with *Proteus* species, strains of *Proteus mirabilis* were mostly sensitive to ampicillin whereas strains of other *Proteus* species, such as *P. morganii*, *P. vulgaris* and *P. rettgeri* were generally resistant. Ampicillin was notably active against *Strep. faecalis*. Strains of *E. coli*, *Proteus mirabilis*, and *Strep. faecalis* appear as the organisms most sensitive to ampicillin and these account for 75% of the total number of isolates tested in this work. They are also reported by Brumfitt *et al.* (1962) and Garrod, Shooter, and Curwen (1954) to be the organisms most frequently encountered in urinary tract infections.

Resistance to ampicillin was encountered, particularly among strains of *Aerobacter* and *Proteus* species other than *P. mirabilis*. Unlike the resistance of staphylococci to penicillin G, the resistance of Gram-negative bacilli to ampicillin can be due in certain cases to penicillinase production, and in other cases to intrinsic resistance, and some bacteria exhibit both of these characteristics. Penicillinase production, however, does not always result in a high degree of resistance to ampicillin. This is because ampicillin is significantly more stable than penicillin G to the penicillinase produced by certain bacteria, and in particular certain strains of *E. coli* (Auhagen, Gloxhuber, Hecht, Knott, Otten, Rauenbusch, Risse, Schmid, Scholtan, and Walter, 1962; Ayliffe, 1963; Percival, Brumfitt, and de Louvois, 1963; Smith and Hamilton-Miller, 1963; Sutherland, 1964). On the other hand ampicillin is generally less stable than penicillin G to the penicillinase produced by aerobacter strains (Smith and Hamilton-Miller, 1963) and the resistance which these strains generally show to ampicillin is usually associated with inactivation of the drug (Ayliffe, 1963; Sutherland, 1964). In contrast, the resistance of *Pseudomonas pyocyanea* to ampicillin is due entirely to intrinsic insensitivity, ampicillin being extremely stable to the penicillinase produced by these organisms (Sutherland, 1964).

The authors are indebted to Dr. A. Knudsen, of the West Middlesex Hospital, Isleworth, Dr. M. Ridley, of St Thomas's Hospital, London, and Dr. R. E. M. Thompson, Bland Sutton Institute of Pathology, Middlesex Hospital.

Medical School, London, for the cultures used in this work.

REFERENCES

- Acrod, P., Brown, D. M., Turner, D. H., and Wilson, M. J. (1962). *Brit. J. Pharmacol.*, **18**, 356.
- Auhagen, E., Gloxhuber, Ch., Hecht, G., Knott, Th., Otten, H., Rauenbusch, E., Risse, K. H., Schmid, J., Scholtan, W., and Walter, A. M. (1962). *Arzneimittel-Forsch.*, **12**, 791.
- Ayliffe, G. A. J. (1963). *J. gen. Microbiol.*, **30**, 339.
- Brumfitt, W., Percival, A., and Carter, M. J. (1962). *Lancet*, **1**, 130.
- Bunn, P. A. (1961). *Antimicrobial Agents and Chemotherapy*, 739.
- Garrod, L. P., Shooter, R. A., and Curwen, M. P. (1954). *Brit. med. J.*, **2**, 1003.
- Knudsen, E. T., Rolinson, G. N., and Stevens, S. (1961). *Ibid.*, **2**, 198.
- Lowe, G. H. (1962). *J. med. Lab. Technol.*, **19**, 21.
- Mackie, T. J., and McCartney, J. E. (1962). *Handbook of Bacteriology*, 10th ed., edited by R. Cruickshank, p. 619. Livingstone, Edinburgh and London.
- Percival, A., Brumfitt, W., and de Louvois, J. (1963). *J. gen. Microbiol.*, **32**, 77.
- Rolinson, G. N., and Stevens, S. (1961). *Brit. med. J.*, **2**, 191.
- Rutenberg, A. M., Greenberg, H. L., Schweinburg, F. B., and Perreault, M. A. (1961). *Antimicrobial Agents and Chemotherapy*, 749.
- Smith, J. T., and Hamilton-Miller, J. M. T. (1963). *Nature (Lond.)*, **197**, 976.
- Stewart, D. J. (1961). *Ibid.*, **191**, 521.
- Stewart, G. T., Coles, H. M. T., Nixon, H. H., and Holt, R. J. (1961). *Brit. med. J.*, **2**, 200.
- Sutherland, R. (1964). *J. gen. Microbiol.*, **34**, 85.
- Thornley, M. J. (1960). *J. appl. Bact.*, **23**, 37.
- Trafford, J. A. P., Maclaren, D. M., Lillicrap, D. A., Barnes, R. D. S., Houston, J. C., and Knox, R. (1962). *Lancet*, **1**, 987.