Septicaemia due to *Pasteurella pneumotropica*

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SYNOPSIS The literature concerning *Pasteurella pneumotropica* infection in animals and man is briefly reviewed and a case presented in which the organism was the cause of septicaemia in a patient receiving chemotherapy for myeloid leukaemia. Bacteriological findings are recorded and compared with those of other authors.

Pasteurella pneumotropica was first described, studied, and so named by Jawetz (1948, 1950) and Jawetz and Baker (1950). This organism, latent in normal mice, and in rats, guinea pigs, and hamsters associated with infected mouse colonies, produced a necrotizing pneumonia in mice only and on serial passage only. In view of this apparent predilection for the lung, a pneumotropism having characteristically no regard for inoculation route, the name *Pasteurella pneumotropica* was proposed.

Although a survey of five laboratory animal breeding establishments in the eastern United States at that time revealed none that was free from this organism, epizootics attributable to it had not been observed. Later workers confirmed the observations on latency (Hoag, Wetmore, Rogers, and Meier, 1962; Flynn, Brennan, and Fritz, 1965; Goldstein and Green, 1967) and primary pathogenicity (Gray and Campbell, 1953; Flynn *et al.*, 1965; Brennan, Fritz, and Flynn, 1965; Innes, Garner, and Stookey, 1967; Wheeler, 1967), and contributed observations on secondary invasion and opportunism (Wheeler, 1967; Brennan, Fritz, and Flynn, 1969). Since the original observations of Jawetz, *Pasteurella pneumotropica* has been isolated from mice having pneumonia (Gray and Campbell, 1953; Brennan *et al.*, 1965; Weisbroth, Scher, and Boman, 1969), conjunctivitis, metritis, cystitis, pleural abscess, peritoneal abscess, and dermatitis (Brennan *et al.*, 1965), together with superficial abscess of the skin, panophthalmitis, and orbital abscess (Weisbroth *et al.*, 1969). Pneumonia in the rat, hamster, and dog, conjunctivitis in the kangaroo, rat, and otitis, renal abscess, septicaemia, and peritonitis in the dog have also been recorded (Brennan *et al.*, 1965).

The association of *Pasteurella pneumotropica* with

human disease, however, is rare. We have found in the literature six cases only. Two strains investigated by Henriksen and Jysum (1961) and Henriksen (1962) were isolated in large numbers in both cases from the nose of a boy aged 16 years with maxillary sinusitis, and from the throat of a boy, aged 7 years, with rhinitis, recurrent tonsillitis, and enlarged adenoids. Two further instances were mentioned by Miller (1966). The first concerned a man aged 51 years who, having been bitten by a dog, developed septicaemia within 48 hours, followed rapidly by collapse and death. Necropsy showed haemorrhage and necrosis of the adrenals as in the Waterhouse-Friderichsen syndrome. *Pasteurella pneumotropica* was recovered from the patient's blood and the dog's mouth. The second concerned a physician bitten on a finger by a dog with clinical pneumonitis, nasal discharge, and cough. Pyrexia, with pain and induration of the wound, developed within a few hours. A purulent discharge, from which *Pasteurella pneumotropica* was isolated, appeared next day. The patient, who was treated with dimethylchlortetracycline, recovered completely. No mention was made regarding isolation of *Pasteurella pneumotropica* from the dog. The fifth case, again with severe local sepsis in an adult, followed a cat bite (Olson and Meadows, 1969).

In the sixth case reported by Winton and Mair (1969) the organism was recovered from the wound of a patient who had been bitten on a finger by a dog and developed local inflammation. She was treated with oral penicillin and the wound healed without further event. No record was made of whether any lymphangitis or adenitis developed.

Case Report

An actor, aged 57 when first seen, gave a six weeks'

history of unaccustomed tiredness and breathlessness accompanied by tachycardia, even after walking short distances. Although anorexia had been present for three months there was no loss of weight. Apart from flecks of blood following nose blowing, bleeding from orifices or bruising had not occurred. The past history included an episode of haematuria 10 years previously and pneumonitis 15 years previously.

On admission he was pale and ill, with a temperature of 38°C; lymph nodes, liver, and spleen were not enlarged. The haemoglobin was 6.3 g (43%), WBC 1300/cmm, platelets 18 000/cmm. The bone marrow features were consistent with a diagnosis of acute myeloblastic leukaemia.

A remission induced with Daunorubicin and cytosine arabinoside was maintained with 6-mercaptopurine for eight months, after which relapse necessitated readmission, and further chemotherapy supported by blood and platelet transfusion. During the course of treatment and 10 days after admission, the patient developed a temperature of 40°C preceded by a sore throat. This, however, was not related to blood or platelet transfusion. Blood cultures yielded a non-motile Gram-negative bacillus having the characteristics of *Pasteurella pneumotropica* and sensitive by preliminary standard disc tests to penicillin, ampicillin, oxytetracycline, chloramphenicol, kanamycin, polymyxin, carbeni-

cillin, cephalothin, gentamicin, and sulphamethoxazole/trimethoprim combination. This organism was not isolated from throat swabs or sputum on the first or second admissions.

Treatment with chloramphenicol followed by ampicillin caused the patient to become afebrile and blood cultures to become sterile in four days. Two days later the temperature rose to 40.6°C accompanied by severe hypotension. He died the same day. Antemortem blood cultures yielded *Klebsiella aerogenes*. Postmortem appearances were those of left ventricular failure and pulmonary oedema.

Bacteriology

Primary isolation was achieved on blood cultures at 37°C from Castaneda's medium (Southern Group Laboratories) incubated aerobically and in carbon dioxide, and also from brain-heart infusion broth containing 0.1% agar.

Subsequent growth at 37°C was abundant on horse's blood or chocolate agar, less so, on nutrient agar, and absent on MacConkey's or desoxycholate medium. Colonies on blood agar at 18 hours were 1 mm in diameter, not haemolytic, greyish with a smooth surface, convex, and round with an entire edge. A strong odour resembling that of *Haemophilus influenzae* was also noted. Morphologically

Biochemical Characteristics	Results in Other Series										
	1	2	3	4	5	6	7	8	9	10	11
Gelatin	-	-	...	-	-	-	...	-	-	...	-
Indol	+	+	+	+	+	+	+	+	+	+	+
Nitrate reduction	+	+	+	+	...	...	+	+	+	...	+
Urease	...	+	+	+	+	+	+	+	+	+	+
Catalase	...	+	...	+	+	+	+	+	+	+	+
Oxidase	...	+	...	+	+	+	+	+	+	+	+
H ₂ S	d ³	+ ² - ⁴	+ ²	- ⁴	...	- ¹ + ²	+ ²	+ ³	+ ²	- ¹ + ²	- ¹ + ²
MR/VP	-/...	-/-	-/-	-/-	-/-	-/-	-/-	-/-	-/-	...	-/-
MacConkey agar	-	-	-	-	-	-	-	-	-	-	-
Malonate	...	...	...	-	...	...	-	-	-	...	-
Arabinose	d	...	-	d	...	...	+	+	+	...	-
Glucose	+	+	+	+	+	+	+	+	+	+	+
Inositol	+	...	...	-	-	-	...	-	-	...	-
Lactose	+	+	-	+	-	+	+	+	-	-	-
Maltose	+	+	+	+	+	+	+	+	+	+	-
Mannitol	...	d	-	-	-	-	-	-	-	-	-
Mannose	+	...	...	...	+	...	+	+	+	...	...
Rhamnose	-	...	-	-	-	...	-	-	-	...	-
Salicin	-	d	-	...	-	...	-	-	-	...	-
Sorbitol	d	...	-	-	-	...	-	-	-	...	-
Sucrose	+	+	+	+	+	+	+	+	+	+	+
Xylose	-	...	+	+	-	+	+	+	-	...	-
Trehalose	...	+	...	...	+	...	...	...	+	...	+

Table I Comparison of biochemical characteristics of *Pasteurella pneumotropica*

1 Jawetz (1950), 2 Hoag, Wetmore, Rogers, and Meier (1962), 3 Henriksen (1962), 4 Heyl (1963), 5 Smith and Thal (1965), 6 Brennan, Fritz, and Flynn (1965), 7 Wheeler (1967), 8 Weisbroth, Scher, and Boman (1969), 9 Winton and Mair (1969), 10 Olson and Meadows (1969), 11 present strain.

*Small amount of gas present, d variable, H₂S, 1 tested on Kligler medium, 2 lead acetate strip, 3 method not stated, 4 iron salts incorporated in medium.

Benzyl- penicillin	Ampicillin	Cephaloridine	Erythromycin	Tetracycline	Chloram- phenicol	Kanamycin	Gentamycin	Polymyxin
0.3	0.15	0.15	0.25	0.8	1.25	1.25	0.6	1.0

Table II MIC of antibiotics for *Pasteurella pneumotropica* ($\mu\text{g/ml}$)

the organism was a small Gram-negative coccobacillus non-motile at 37 and 22°C, which did not show bipolar staining. Biochemical activities are shown in table I, antibiotic sensitivities in table II. We concluded that the organism was *Pasteurella pneumotropica*.

Confirmation of the identity of our strain was subsequently obtained at the National Collection of Type Cultures Laboratory in a computer-assisted identification and given the number A 141/71.

Comparisons between the biochemical activities of *Pasteurella pneumotropica* strains obtained by various workers are shown in table I. Those of Jawetz (1950) utilized inositol but not xylose. By contrast those of Heyl (1963), Brennan *et al* (1965), Wheeler (1967), and Weisbroth *et al* (1969); utilized xylose but not inositol or mannitol. While our own results are in harmony with those of Smith and Thal (1965), in that neither inositol or xylose were utilized, they differed from all others in the production of small amounts of gas in glucose, sucrose, and trehalose.

Comment

Although in our case *Pasteurella pneumotropica* was undoubtedly acting as an opportunist invader, the other examples of human infection so far recorded, scanty though they may be, would suggest that its abilities as a primary pathogen should not be underestimated.

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