increasing the risk of heatstroke.^{8,9} The explanation proposed here does not preclude these possibilities. Indeed, infection is a major cause of both pyrexia and decreased arousal due to cytokine production. Thus, a number of these factors could act together to bring about a fatal outcome.

- 1 Fleming PJ, Gilbert R, Azaz Y, Berry PJ, Rudd PT, Stewart A, *et al.* Interaction between bedding and sleeping position in sudden infant death syndrome: a population based case control study. *BMJ* 1990;301:85-9.
- 2 Engelberts AC, de Jonge GA. Choice of sleeping position for infants: possible association with cot death. *Arch Dis Child* 1990;65:462-7.

- 3 Taylor BJ. A review of epidemiological studies of sudden infant death syndrome in Southern New Zealand. *J Paediatr Child Health* 1991;27:344-8.
- 4 Morris JA, Haran D, Smith A. Hypothesis: common bacterial toxins are a possible cause of the sudden infant death syndrome. *Med Hypotheses* 1987;22:211-22.
- 5 Morris JA. Sudden infant death syndrome. *BMJ* 1989;298:958.
- 6 Telford DR, Morris JA, Hughes P, Conway AR, Lee S, Barson AR, *et al.* The nasopharyngeal bacterial flora in sudden infant death syndrome. *J Infect* 1989;18:125-30.
- 7 McKendrick N, Drucker DB, Morris JA, Telford DR, Barson AJ, Oppenheim BA, *et al.* Bacterial toxins: a possible cause of cot death. *J Clin Pathol* 1992;45:49-53.
- 8 Nelson EAS, Taylor BJ, Weatherall IL. Sleeping position and infant bedding may predispose to hyperthermia and the sudden infant death syndrome. *Lancet* 1989;i:199-200.
- 9 Ponsonby A-L, Dwyer T, Gibbons LE, Cochrane JA, Wang Y-G. Factors potentiating the risk of sudden infant death syndrome associated with the prone position. *N Engl J Med* 1993;329:376-82.

J Clin Pathol 1996;49:172-173

Nosocomial empyema caused by *Clostridium difficile*

A J H Simpson, S S Das, S Tabaqchali

Abstract

Pleural infection with *Clostridium difficile* is extremely rare. A case of nosocomial empyema following chest drain insertion in a 46 year old man is described. The potential of *C difficile* to cause extra-intestinal infections should be recognised and its isolation from other sites should not be ignored.

(*J Clin Pathol* 1996;49:172-173)

Keywords: *Clostridium difficile*, empyema, nosocomial infection.

Case report

A 46 year old man was transferred from another hospital to St Bartholomew's Hospital, London, with a history of incessant atrial tachycardia secondary to alcoholic cardiomyopathy. Following admission, his tachycardia was treated and stabilised with digoxin 0.25 mg per day and verapamil 80 mg three times daily.

However, an admission chest x ray revealed bilateral pleural effusions, larger on the right. There was increased shadowing and a cavitating mass in the left upper zone, with erosion of the first and second ribs anteriorly, suggestive of malignancy. There was no previous history of tuberculosis. The patient became pyrexial (38°C) 24 hours after admission, and after collection of blood cultures, was started on intravenous cefuroxime 750 mg three times daily and oral erythromycin 500 mg four times daily for a suspected chest infection. He had not previously received any antibiotics. No sputum samples were produced. A diagnostic tap of the right pleural effusion showed pus cells, but no bacteria, and was sterile on culture. A Ziehl-Neelsen (ZN) stain for acid/alcohol fast bacilli was negative. The protein content was 37 g/l and the glucose con-

centration 6 mmol/l. Cytology showed abundant polymorphs and reactive mesothelial cells, but no malignant cells. A rapid micro-agglutination test (RMAT) titre for *Legionella pneumophila* was less than 1:8.

The patient had a low grade fever over the following week and enlarging pleural effusions. His peripheral white cell count rose from $14.8 \times 10^9/l$ to $19.6 \times 10^9/l$. A pleural biopsy was performed, followed by right chest drain insertion and drainage of 1200 ml blood stained fluid. Routine culture on blood, chocolate and cystine lactose electrolyte deficient (CLED) agar in an aerobic atmosphere with 5% CO₂ at 37°C, and on blood agar anaerobically (80% nitrogen, 10% hydrogen and 10% CO₂) at 37°C, yielded no growth after 48 hours. ZN staining was again negative and all cultures for acid fast bacilli were negative at eight weeks. Tuberculin skin testing (1:1000) was negative.

Erythromycin was stopped and intravenous metronidazole added (500 mg three times daily), in case of aspiration post-cardioversion (attempted before admission to this hospital). Three days later, drainage from the chest drain ceased; a further pleural tap and biopsy was performed, producing thickened blood stained fluid. Cytology showed fibrinous material and many white cells consistent with an empyema. Direct Gram staining showed scanty large Gram positive rods and many neutrophils; anaerobic culture, as described above, revealed a pure growth of *Clostridium difficile*. Plates incubated aerobically showed no growth. No other specimens processed during this period showed evidence of Gram positive rods on staining, nor was *C difficile* isolated from any other specimen, suggesting that laboratory contamination was extremely unlikely. The isolate was positive for toxin A using the Premier *C difficile* Toxin A EIA kit (Meridian Diagnostics

Department of
Medical Microbiology,
St Bartholomew's
Hospital Medical
College, West
Smithfield, London
EC1A 7BE
A J H Simpson
S S Das
S Tabaqchali

Correspondence to:
Dr A J H Simpson.

Accepted for publication
5 September 1995

Europe, Milan, Italy). No stool samples were available for culture or direct toxin testing, but the patient did not have diarrhoea at any time during this admission.

The patient improved clinically over the next 72 hours, became afebrile and was sent back to his referring hospital as his cardiological problems had been controlled. He received metronidazole for six days in total. The chest drain was removed prior to transfer, although he still had a right pleural effusion. He did not receive any further antibiotics. Appearances on chest x ray film did not change further and it was concluded that these were caused by a chronic inflammatory process. The patient was stable six months after discharge and refused further investigation of his chest disease, including further pleural taps.

Discussion

Although *C. difficile* is well established as the major cause of pseudomembranous colitis (PMC), antibiotic associated colitis (AAC) and antibiotic associated diarrhoea (AAD),¹⁻³ it is very rarely isolated from extraintestinal specimens, especially pleuropulmonary specimens,⁴ and is very rarely a cause of extraintestinal infection.⁵ Large outbreaks of AAC/AAD have occurred in hospitals⁶ and most cases of intestinal infection follow nosocomial acquisition of the organism.⁷ Such acquisition is likely in this patient, causing an empyema secondary to attempted drainage of a pleural effusion, although a primary aspiration pleuropneumonia cannot be excluded. The latter possibility seems less likely as the previous pleural fluid specimens were sterile, but we have no other satisfactory explanation for his chest disease. However, there were no cases of *C. difficile* associated diarrhoea on the same ward during his stay in this hospital.

Isolation of *C. difficile* from extraintestinal sites is frequently of little significance,⁵ but in this case isolation in pure culture and the response to aspiration and treatment with metronidazole suggest a pathogenetic role.

Risk factors for *C. difficile* associated intestinal disease include prior use of antibiotics, with the elderly and those with serious underlying conditions additionally predisposed.² The number of reported cases of pleuropulmonary infection due to *C. difficile* is insufficient to draw reliable conclusions about risk factors for this disease, but in this case the patient had been receiving broad spectrum antibiotics.

Pleuropulmonary infections with other clostridial species, usually *C. perfringens*, have been reported occasionally.⁸ Very few cases of pleural infection with *C. difficile* have been reported, the first two cases being described in 1962.⁹

Pleural infections caused by clostridial species seem to behave clinically like empyemas due to other causes.¹⁰ In this case the process seems to have been relatively benign, assuming that this was a secondary infection and not the cause of the patient's underlying chest disease.

We suggest that isolation of *C. difficile* from extraintestinal sites should not be ignored and that its recognition as a nosocomial pathogen be extended beyond its role in AAD and PMC.

We thank Dr A W Nathan for permission to report this case.

- 1 Bartlett JG. *Clostridium difficile*: clinical considerations. *Rev Infect Dis* 1990;12(Suppl 2):S243-51.
- 2 Tabaqchali S, Jumaa P. Diagnosis and management of *Clostridium difficile* infection. *BMJ* 1995;310:1375-80.
- 3 Kelly CP, Pothoulakis C, LaMont JT. *Clostridium difficile* colitis. *N Engl J Med* 1994;330:257-62.
- 4 Marina M, Strong CA, Civen R, Molitoris E, Finegold SM. Bacteriology of anaerobic pleuropulmonary infections: preliminary report. *Clin Infect Dis* 1993;16(Suppl 4):S256-62.
- 5 Levett PN. *Clostridium difficile* in habitats other than the human gastro-intestinal tract. *J Infect* 1986;12:253-63.
- 6 Cartmill TDI, Panigrahi H, Worsley MA, McCann DC, Nice CN, Keith E. Management and control of a large outbreak of diarrhoea due to *Clostridium difficile*. *J Hosp Infect* 1994;27:1-15.
- 7 Heard SR, O'Farrell S, Holland D, Crook S, Barnett MJ, Tabaqchali S. The epidemiology of *Clostridium difficile* infection with use of a typing scheme: nosocomial acquisition and cross infection among immunocompromised patients. *J Infect Dis* 1986;153:159-62.
- 8 Lorber B. Gas gangrene and other *Clostridium*-associated diseases. In: Mandell GL, Bennett JE, Dolin R, eds. *Mandell, Douglas and Bennett's Principles and practice of infectious diseases*. 4th edn. New York: Churchill Livingstone; 1994:2182-95.
- 9 Smith LDS, King EO. Occurrence of *Clostridium difficile* in infections of man. *J Bacteriol* 1962;84:65-7.
- 10 Spagnuolo PJ, Payne VD. *Clostridium* pleuropulmonary infection. *Chest* 1980;78:622-5.

Gastroenteritis caused by *Aeromonas trota* in a child

J Reina, A Lopez

Clinical Microbiology Service, University Hospital Son Dureta, Universitat Illes Balears (UIB), 07014-Palma de Mallorca, Spain
J Reina
A Lopez

Correspondence to:
Dr Jordi Reina.

Accepted for publication
5 September 1995

Abstract

A case of acute diarrhoea caused by *Aeromonas trota* (formerly HG 13 group) in a Spanish child is reported. The strain was isolated in the faeces using the CIN agar (cefsulodin-irgasan-novobiocin) culture media. The strain was initially identified as *A. sobria* by the commercial GNI card

and API 20E biochemical systems. The strain was, however, Voges-Proskauer and sucrose negative, so complementary tests of cellobiose fermentation and gluconate oxidation were performed. These tests, together with the strain susceptibility to ampicillin (MIC 1 µg/ml) and carbenicillin (MIC <16 µg/ml) led to the final iden-