ORIGINAL ARTICLE

Microsatellite analysis of the adenomatous polyposis coli (APC) gene and immunoexpression of β catenin in nephroblastoma: a study including 83 cases treated with preoperative chemotherapy

A Ramburan, F Oladiran, C Smith, G P Hadley, D Govender

J Clin Pathol 2005;58:44–50. doi: 10.1136/jcp.2004.019752

See end of article for authors' affiliations

Correspondence to:
Dr D Govender, Division of
Anatomical Pathology,
Faculty of Health Sciences,
University of Cape Town,
Anzio Road, Observatory,
7925, Cape Town, South
Africa; dgovende@
chempath.uct.ac.za

Accepted for publication
28 June 2004

Aims: To determine whether microsatellite mutations of the adenomatous polyposis coli (APC) gene have pathological or prognostic significance in nephroblastomas and to correlate APC alterations with β catenin immunoexpression.

Methods: One hundred nephroblastomas were analysed, 83 of which received preoperative chemotherapy. Normal and tumour DNA was isolated using standard proteinase K digestion and phenol/chloroform extraction from paraffin wax embedded tissue. Polymerase chain reaction using four APC microsatellite markers—D5S210, D5S299, D5S82, and D5S346—was performed and the products analysed. Immunohistochemistry was performed using the LSAB kit with diaminobenzidine as chromogen. Results were correlated with clinicopathological data using the χ^2 test.

Results: Allelic imbalance/loss of heterozygosity was more frequent than microsatellite instability, with 30% of cases showing allelic imbalance/ loss of heterozygosity and 16% showing microsatellite instability. Although there was a significant correlation between the results for individual markers and the clinicopathological data, the overall results do not support a prognostic role for APC in nephroblastoma. Expression of β catenin was seen in 93% of cases. Staining was predominantly membranous, with epithelium, blastema, and stroma being immunoreactive. Cytoplasmic redistribution was seen in 58% of cases, but no nuclear staining was detected. No significant associations between β catenin expression and the clinicopathological parameters were found. Kaplan–Meier survival plots showed that patients with loss of membranous staining and pronounced cytoplasmic staining (score, 3) had a significantly shorter survival ($p=0.04$; median survival, 5.87 months).

Conclusion: Microsatellite analysis of APC and immunoexpression of β catenin did not provide significant pathological or prognostic information in this cohort of nephroblastomas.

The tumour suppressor adenomatous polyposis coli (APC) gene is frequently mutated or deleted in colon cancers.^{1,2} Germline mutations in the APC gene are responsible for the autosomal dominant inherited disease familial adenomatous polyposis,^{3,4} whereas somatic mutations in APC occur in approximately 80% of sporadic colorectal tumours.⁵ The APC gene is a relatively large gene, which encodes a 8.5 kb mRNA molecule and a 2843 amino acid protein in its most common isoform.⁶ The APC protein is structurally complex, with many possible protein–protein interaction sites, including binding sites for β catenin, EB1, and axin.⁷ APC participates in a variety of cellular functions including proliferation, differentiation, apoptosis, adhesion, migration, and chromosomal segregation.⁸

The APC gene product is part of a multiprotein complex that regulates the cytoplasmic concentration of β catenin, which has two major functions, cell adhesion and the transmission of the proliferation signal of the Wnt pathway.^{9,10} Under normal circumstances, APC binds directly to free β catenin and promotes its phosphorylation by glycogen synthetase kinase 3 β (GSK-3 β), thereby targeting β catenin for degradation by the proteasome system.¹¹ However, when there are mutations in the APC or β catenin gene, especially in the GSK-3 β phosphorylation region, APC fails to bind β catenin,¹² which then accumulates within the cell cytoplasm and nucleus. Within the nucleus, β catenin binds directly to the T cell factor/lymphoid enhancer factor group of

DNA binding transcription factors and stimulates the transcription of target genes.¹³ C-myc, cyclin D1, and fibronectin have been identified as possible target genes for the APC– β catenin pathway. Overexpression of c-myc and/or cyclin D1 is frequently seen in various human cancers,^{14,15} and fibronectin plays an important role in cancer metastasis.¹⁶

“APC participates in a variety of cellular functions including proliferation, differentiation, apoptosis, adhesion, migration, and chromosomal segregation”

At our institution, nephroblastoma is the most common solid malignant renal tumour in childhood. As a result of poor nutritional status, intercurrent infection, and advanced disease at presentation, our patients have a worse survival than patients from developed countries.¹⁷ Despite attempts to identify other variables of prognostic value, histology and clinicopathological stage remain the most reliable prognostic determinants for nephroblastoma.^{18–20} Molecular studies have identified abnormalities at several loci for nephroblastoma. Loss of heterozygosity for 11p13 (WT1) and 11p15 (WT2) have been implicated directly in the development of

Abbreviations: APC, adenomatous polyposis coli; DCC, deleted in colon cancer; GSK-3 β , glycogen synthetase kinase 3 β ; IL, interleukin; PCR, polymerase chain reaction

Table 1 Chromosome 5q markers

Marker	Primer sequence	Chromosome location	Map position (cM)	Heterozygosity	Product size (bp)
D5S82	(F) ATCAGAGTATCAGAATTCT (R) CCCAATTGTATAGATTAGAAGTC	5q14-q21	118.8	0.758	169-179
D5S299	(F) GCTATTCTCTCAGGATCTTG (R) GTAAGCCAGGACAAGATGACAG	5q15-q22	127.0	0.704	156-182
D5S346	(F) ACTCACTCTAGTGATAAATCGGG (R) AGCAGATAAGACAGTATTACTAGTT	5q21-q22	128.9	0.820	96-122
D5S210	(F) AAATATTGAAGCTGAGTGCC (R) CTTAACATCCTTAAACAGC	5q31.3-q33.3	169.1	0.782	112-132

F, forward; R, reverse.

nephroblastoma, whereas losses at loci 11q, 7p, 4q, 3q, 9p, 17p, and 16q have been associated with an outcome or phenotype, but have no direct role in tumour development.²¹⁻²⁶

Based on findings from our microsatellite study on the deleted in colorectal cancer gene (DCC), where multiple microsatellite aberrations of this tumour suppressor gene conferred a worse prognosis,²⁷ we extended the study to evaluate allelic imbalance/loss of heterozygosity and microsatellite instability of the APC gene. Therefore, our present study was conducted to determine whether mutations in the microsatellite sequences of the APC gene have pathological or prognostic significance in nephroblastomas. Furthermore, the APC alterations were correlated with β catenin immun-expression.

MATERIALS AND METHODS

Our study comprised 100 cases of nephroblastoma from the files of the department of pathology, University of KwaZulu-Natal, Durban, South Africa. All cases were obtained from nephrectomies performed over an 18 year period from 1984 to 2001. Clinical data were available for all patients. Patients were managed according to the protocols of the International Society of Paediatric Oncology. Preoperative chemotherapy was administered in 83 of the 100 patients studied. Haematoxylin and eosin stained slides were reviewed and the histological type and tumour staging parameters were confirmed. In our study, nephroblastomas with unfavourable histology referred to those nephroblastomas with diffuse anaplasia.¹⁹ Normal DNA was extracted from normal renal tissue (n = 97) and normal lymph nodes (n = 3). For tumour DNA extraction and immunohistochemistry, paraffin wax embedded tissue blocks containing maximal tumour and minimal or no necrosis were selected. For the molecular study, contamination of tumour DNA by normal DNA was minimised by microdissecting the tumour meticulously. Normal and tumour tissue were marked out on the haematoxylin and eosin sections using a permanent marker. This was then superimposed on the unstained tissue section and marked out on the reverse side using a permanent marker. Normal or tumour tissue was then scraped away from the glass slides using a sterile scalpel blade and transferred into separate 1.5 ml Eppendorf tubes. This was followed by removal of the paraffin wax.

Microsatellite study

The standard proteinase K digestion and phenol/chloroform extraction method was used for DNA extraction. Amplifiable DNA was verified by polymerase chain reaction (PCR) demonstration of the insulin gene.²⁸

Primer sequences were obtained from the Genome Database (<http://www.gdb.org>). Table 1 lists the Cy5 labelled microsatellite primers (Roche Diagnostics, Mannheim,

Germany) used to amplify the APC gene. These primers were chosen for their high informativity rate.

PCR was carried out in a total volume of 25 μ l, containing 10pmol of each Cy5 labelled primer pair, 2 μ l template DNA, 200 μ M dNTPs, 50mM PCR buffer containing 1.5mM MgCl₂, and 0.75 U Taq DNA polymerase. The PCR amplification was performed using a GeneAmp PCR System 9700 (Applied Biosystems, Foster City, California, USA). The PCR consisted of an initial denaturation step of five minutes at 95°C, followed by 35 cycles of one minute at 94°C, 30 seconds at 55°C, and 30 seconds at 72°C, with a final extension step at 72°C for seven minutes. Aliquots of 3 μ l of the microsatellite PCR products were mixed with 3 μ l of STOP solution (blue dextran 2000, deionised formamide; Pharmacia Biotech, Uppsala, Sweden) and denatured at 96°C for three minutes in a GeneAmp PCR System 9700 (Applied Biosystems). The samples were held on ice before being loaded on to the gel. A Cy5 labelled 50-500 bp standard was used as the external size marker (Pharmacia Biotech).

The microsatellite PCR products were analysed on the ALF Express automated DNA sequencer using a 6% Longranger sequencing gel (FMC Bioproducts, Rockland, Maine, USA). The analysis was performed under the following conditions: 1500 V, 60 mA, 15 W, at a constant temperature of 55°C. The data were analysed using the Fragment Manager software program.

Allelic imbalance was determined by using the following ratio: $(T2 \times N1)/(T1 \times N2)$, where T/N 1 and 2 are the first and second peaks of tumour and normal DNA, respectively.²⁹ A ratio < 0.6 was regarded as allelic imbalance with loss of the larger allele, and a ratio > 1.67 was regarded as loss of the smaller allele. The presence of a novel allele in the tumour sample was interpreted as microsatellite instability.³⁰

Immunohistochemical study

The β catenin specific antibody (clone CAT-5H10; Zymed Laboratories Inc, San Francisco, California, USA) was used at a dilution of 1/300. Tissue sections (2 μ m) were picked up on poly-L-lysine (Sigma Diagnostics, St Louis, Missouri, USA) coated slides and routinely dewaxed in xylene and alcohol. To achieve optimal antigen retrieval, the sections were incubated in a microwave oven (H2500 Microwave Processor; Energy Beam Sciences, Agawam, Massachusetts, USA) for 10 minutes at 85°C in 0.01 mol/litre trisodium citrate buffer at pH 6.0. The slides were incubated in 3% H₂O₂ for five minutes to quench endogenous peroxidase activity. The peroxidase labelled streptavidin biotin kit (Dako) was used to perform the tests and 3,3 diaminobenzidine (liquid DAB; Dako) was used as a chromogen. Sections were then counterstained with Mayer's haematoxylin. Positive (normal breast tissue) and negative controls were run simultaneously. For the negative control the primary antibody was replaced by phosphate buffered saline in the staining method.

	Lane	Peak area	Base pair size		Lane	Peak area	Base pair size
	1		50–500 base pair external marker				
A	26	4453.9	180.2	D	18	1109.6	117.9
		2830.3	185.9			508.5	121.3
	27	1490.3	181.0		19	1499.5	116.0
		815.0	186.7			606.4	113.4
B	2	2814.7	172.1	E	35	255.4	128.3
	3	3119.4	172.2			194.1	113.4
C	12	922.3	121.1		36	285.7	123.1
		556.0	128.7			154.5	128.6
	13	206.0	121.3				
		583.2	128.8				

Figure 1 Allelic imbalance/loss of heterozygosity and microsatellite instability were based on electrophoretograms that were generated by the Fragment Manager Software. (A) Two peaks in the normal and tumour tissue was reflective of a heterozygous or informative case (marker D5S299), whereas (B) the appearance of a single peak in both the normal and tumour tissue (marker D5S82) was regarded as a homozygous or non-informative case. (C) A case displaying allelic imbalance (marker D5S210; ratio = 4.69, therefore the smaller allele is lost); (D) loss of heterozygosity (marker D5S346); and (E) an example of microsatellite instability (marker D5S210).

The immunostaining was graded according to the method of Jass *et al.*³¹ Briefly, a score of 1 was awarded for loss of cell membrane staining; 1 or 2 for slight and pronounced increase in cytoplasmic staining, respectively; and 1 or 2 for slight and pronounced nuclear staining, respectively. The maximum total score was 5 and cases scoring 4 or more were regarded as positive for abnormal β catenin immunolocalisation. The

grading was performed manually (DG, AR) using an Olympus BX40 microscope.

Statistical analysis

Statistical analysis was performed using the Statistical Package for Social Sciences software program (SPSS, Chicago, Illinois, USA). The results of APC microsatellite

Table 2 Data generated for the four microsatellite markers				
	D5S82	D5S299	D5S346	D5S210
No change, heterozygous	42	25	54	46
No change, homozygous	45	59	26	46
Allelic imbalance/LOH	5	11	18	7
MSI	8	5	2	1
Total cases	100	100	100	100
% Informativity	55	41	74	54

LOH, loss of heterozygosity; MSI, microsatellite instability.

Table 3 Sex versus markers D5S82 and D5S299 (informative cases)				
	D5S82		D5S299	
	F	M	F	M
No change, heterozygous	22	20	11	14
Allelic imbalance/LOH	5	0	2	9
Microsatellite instability	6	2	4	1
p Value	0.078		0.061	

LOH, loss of heterozygosity.

Table 4 Clinicopathological stage versus marker D5S82 (informative cases)

	D5S82			
	Stage 1	Stage 2	Stage 3	Stage 4
No change, heterozygous	8	13	9	12
Allelic imbalance/LOH	2	1	1	1
Microsatellite instability	0	0	0	8
p Value	0.011			

LOH, loss of heterozygosity.

analysis and β catenin immunoexpression were correlated with age at diagnosis, sex, clinicopathological stage, histological classification, preoperative chemotherapy, and patient outcome. They were assessed by cross tabulation and the significance was determined by the χ^2 test. Differences were considered significant at $p < 0.05$. Survival curves were estimated by means of the Kaplan–Meier method, and differences in survival were evaluated with the log rank test. For the molecular study, only informative cases were analysed.

RESULTS

There were 51 female and 49 male patients. The patients' ages ranged from 2 months to 14 years. The median age at diagnosis was 3 years. Eighty three patients received preoperative chemotherapy. There were 83 favourable histology and 17 unfavourable histology (diffuse anaplasia) tumours. Fourteen patients had stage I disease, 23 had stage II, 23 had stage III, and 40 had stage IV disease. There were two stage V patients who were further substaged as stage I (both kidneys) and stage IV. Twenty eight patients were alive and disease free. There were 53 tumour related deaths, two treatment related deaths, and four deaths as a result of other illnesses. Thirteen patients were lost to follow up. The overall follow up ranged from 25 days to 14 years and nine months. The median survival period was 18 months.

In our study, 30% of cases showed allelic imbalance/loss of heterozygosity for at least one microsatellite marker (fig 1). At the D5S82 locus, allelic imbalance/loss of heterozygosity was seen in five of 55 informative cases (table 2), at the D5S210 locus in seven of 54 cases, at the D5S299 locus in 11 of 41 cases, and at the D5S346 locus in 18 of 74 cases. In contrast, microsatellite instability of at least one marker was seen in 16% of cases.

There appeared to be an association between sex and the microsatellite changes, although this association was not significant. For marker D5S299, a greater proportion of allelic imbalance/loss of heterozygosity was seen in male patients, whereas microsatellite instability was seen more frequently among female patients ($p = 0.061$). For marker D5S82, girls had a greater propensity for both allelic imbalance/loss of

Table 5 Patient outcome versus marker D5S82 (informative cases)

	D5S82		
	Alive	Dead (TR)	Dead (NTR)
No change, heterozygous*	17	16	2
Allelic imbalance/LOH	3	1	1
Microsatellite instability	0	8	0
p Value	0.027		

LOH, loss of heterozygosity; NTR, not tumour related; TR, tumour related.

*Excluding cases lost to follow up.

Table 6 Results of the APC markers versus administration of preoperative chemotherapy (informative cases)

Marker		Preoperative chemotherapy		p Value
		No	Yes	
D5S82	AI	1	4	0.902
	MSI	1	7	
	NAI	8	34	
D5S210	AI	1	6	0.078
	MSI	1	0	
	NAI	7	39	
D5S299	AI	1	10	0.713
	MSI	1	4	
	NAI	5	20	
D5S346	AI	2	16	0.362
	MSI	1	1	
	NAI	9	45	

heterozygosity and microsatellite instability ($p = 0.078$; table 3).

For marker D5S82, allelic imbalance/loss of heterozygosity was observed in all stages, whereas the eight cases with microsatellite instability were all stage 4 tumours ($p = 0.011$; table 4). This marker also had a significant association with patient outcome ($p = 0.027$). All eight patients with microsatellite instability died (table 5).

No significant associations were found when preoperative chemotherapy (table 6), age at diagnosis, and histological classification were independently correlated with the results of microsatellite analysis. In addition, Kaplan–Meier survival plots showed no significant differences in survival between those patients with tumours that harboured APC microsatellite mutations and those that did not.

We found that β catenin was expressed in 93% of cases, and 35% of these cases showed membranous β catenin staining only (fig 2), 52% had both a membranous and cytoplasmic distribution pattern, and 6% displayed cytoplasmic staining only (table 7). No nuclear staining was seen. The distribution of staining in the histological component was as follows: the epithelium was immunoreactive in 80 cases, the blastema in 46 cases, and the stroma in 29 cases.

No significant relations were found when β catenin was independently correlated with sex ($p = 0.763$), preoperative chemotherapy ($p = 0.545$), histological classification

Figure 2 Intense membrane and focal cytoplasmic staining for β catenin in tubular epithelial cells (score, 3).

Table 7 Immunoexpression of β catenin

Score	No of cases	Type of staining	No of cases
0	35	Membranous only	35
1	39	Loss of staining (both membranous and cytoplasmic)	7
		Membranous staining with slight cytoplasmic staining	32
2	22	Loss of membranous staining with slight cytoplasmic staining	2
		Membranous staining with pronounced cytoplasmic staining	20
3	4	Loss of membranous staining with pronounced cytoplasmic staining	4
Total	100		100

($p = 0.171$), stage ($p = 0.081$), and patient outcome ($p = 0.285$). Kaplan–Meier survival plots showed that patients with loss of membranous staining and pronounced cytoplasmic staining (score, 3) had a significantly shorter survival ($p = 0.04$; median survival, 5.87 months). All four patients had stage 4 disease and tumours with favourable histology.

There were no significant correlations between β catenin expression and APC microsatellite mutations.

DISCUSSION

Kinzler and Vogelstein's "pathway to neoplasia" model subdivides genes involved in tumour development into two groups, namely: "gatekeepers" and "caretakers".³² The gatekeepers are directly involved in cell cycle regulation—the proteins they encode control tumour progression by promoting or inhibiting the processes associated with cell division. Inactivation of gatekeeper genes is a rate limiting step for the initiation of a tumour, and both copies must be altered for tumour development. Mutations in other genes may not lead to sustainable growth perturbation in the presence of normal gatekeeper function.³² The APC gene is a well known gatekeeper gene and others include DCC and TGF β IIIR (the gene encoding the transforming growth factor β II receptor). Loss or mutation of APC is an early event in the tumorigenic process known to trigger the "adenoma–carcinoma" sequence of colorectal tumours.³³

Mutations or allelic deletions of the APC gene are present in nearly 80% of sporadic colorectal carcinomas.^{34–35} In addition to colorectal cancers, somatic mutations of APC have been described in gastric cancer,³⁶ hepatoblastoma,³⁷ pilomatricoma,³⁸ synovial sarcoma,³⁹ medulloblastoma,⁴⁰ and oesophageal cancer.⁴¹ Biallelic loss of APC has also been demonstrated in hepatoblastoma⁴² and hepatocellular adenoma.⁴³ In our present study, allelic imbalance/loss of heterozygosity in at least one of four markers of the APC gene was seen in 30% of cases, whereas microsatellite instability was seen in 16% of cases. Allelic imbalance/loss of heterozygosity appears to be a more frequent aberration than microsatellite instability in nephroblastomas. Previous studies involving chromosome 5q in nephroblastomas have detected allelic loss and chromosomal translocation in rare cases.^{44–45} In contrast, our study demonstrates the more important 5q abnormalities.

Microsatellite instability, caused by the somatic insertion or deletion of one or more copies of the repeat unit of a microsatellite, is believed to result from slippage by DNA polymerases during DNA replication.^{46–47} Microsatellite instability appears to be a late event in nephroblastoma because all eight patients with microsatellite instability at marker D5S82 had stage 4 tumours, and these eight patients

also had a poor outcome. Although microsatellite instability in our cohort was associated with a worse outcome, this may not be an independent prognostic factor because high clinicopathological stage is also a poor prognostic factor.

The other important prognostic factor for nephroblastoma is histological classification. Anaplastic or unfavourable histology nephroblastomas are associated with a poor prognosis because of the putative chemoresistance of the anaplastic tumour cells.⁴⁸ A previous microsatellite study identified a significant relation between histological classification and microsatellite changes. Genetic changes were more frequent in tumours with unfavourable histology compared with those with favourable histology.²⁷ However, we could not verify these observations.

Marker D5S346 was the most informative marker, followed by markers D5S82, D5S210, and D5S299. In addition, marker D5S346 showed the highest number of aberrations (allelic imbalance/loss of heterozygosity and microsatellite instability), followed by markers D5S299, D5S82, and D5S210. The informativity of the APC markers in our cohort was lower than that published by the Genome Database. Previous microsatellite studies in similar populations have also found very low informativity rates.^{27–49}

"Microsatellite instability appears to be a late event in nephroblastomas because all eight patients with microsatellite instability at marker D5S82 had stage 4 tumours, and these eight patients also had a poor outcome"

The APC gene may not be the target of loss of heterozygosity in nephroblastoma because the frequency of APC loss was fairly low. Deletions involving multiple chromosomal loci have been reported in several human cancers. Genes on chromosome 5q other than APC include the IRF-1 (interferon regulatory factor 1) gene, the cytokine genes interleukin 3 (IL-3), IL-4, IL-5, and GM-CSF (granulocyte–macrophage colony stimulating factor), and the mitotic inducer CDC25C gene.⁵⁰ Loss of 5q distinct from the APC locus has been reported in carcinomas of the lung,⁵¹ liver,⁵² and ovary,⁵³ and in acute myelogenous leukaemia and myelodysplastic syndrome.⁵⁴ Another tumour suppressor gene from the 5q21 region, MCC (mutated in colorectal cancer), has also been implicated in the pathogenesis of sporadic colorectal, lung,⁵⁵ and oesophageal cancers.⁴¹ The mismatch repair gene hMSH3 and tumour suppressor gene del-27 are also located on the long arm of chromosome 5.

The immunoexpression of β catenin has been investigated in a wide variety of human malignancies where loss and aberrant staining patterns have been associated with tumour dedifferentiation^{56–57} and a poor outcome.^{58–60} In our study β catenin was expressed in 93% of cases. Consistent with other studies,^{61–62} there appears to be no appreciable loss of β catenin in nephroblastoma. β Catenin had a membranous localisation in 88% of cases, and was expressed not only in the epithelium but also in the mesenchyme and blastema.

Nuclear relocalisation of β catenin has been associated with a poor outcome in tumours such as hepatoblastoma⁶⁰ and synovial sarcoma.⁶³ Previous immunohistochemical studies on nephroblastomas have also noted cytoplasmic and nuclear β catenin staining,^{61–62} but this finding was not associated with clinicopathological parameters. Although cytoplasmic β catenin staining was seen, according to Jass and colleagues³¹ this is not classed as abnormal β catenin immunolocalisation, and none of the cases achieved a score of more than 3. However, patients with a score of 3 had a shorter survival, although this finding may be stage dependent, and not necessarily related to the pronounced

Take home messages

- In the nephroblastomas studied, allelic imbalance/loss of heterozygosity of the APC (adenomatous polyposis coli) locus was a more frequent genetic aberration than microsatellite instability
- Although there was a significant correlation between some of the individual markers and the clinicopathological markers, the overall results do not support a prognostic role for the APC gene in nephroblastoma
- There were no significant correlations between the expression of β catenin and APC microsatellite mutations

cytoplasmic staining and associated loss of membranous staining.

The cytoplasmic redistribution of β catenin could be a result of either wnt signalling, inactivation of APC, or direct mutation of β catenin itself. Mutation of the β catenin gene has been demonstrated in 15% of nephroblastomas.⁶⁴ In the study by Koesters *et al*, nuclear accumulation of β catenin was detected in tumours harbouring a mutation in the β catenin gene.⁶¹ Nuclear accumulation of β catenin has also been reported in tumours that lack β catenin mutations, suggesting defects in other members of the Wnt signalling pathway.⁶¹

Interactions of β catenin with several molecular partners have been shown to be important for cell signalling and cell-cell adhesion. Molecules such as E-cadherin, α catenin, the epidermal growth factor receptor, and fascin, among many others,¹⁰ interact with β catenin. The functional consequences of some of these interactions are well established, whereas the importance of others is still unclear. Changes in β catenin concentrations will probably also impact on the function of these molecules, and vice versa. We found no significant associations between β catenin immunorexpression and APC mutations. Although β catenin retained its expression and relatively few APC mutations were seen, abnormalities in other members of the APC- β catenin signalling pathway may still exist.

The administration of chemotherapy or radiotherapy may result in diverse morphological changes in tumours.⁶⁵⁻⁶⁷ These include maturation and cytodifferentiation in primitive embryonal tumours of childhood, including nephroblastomas. Although the exact mechanism for this phenomenon has not been identified, underlying genetic changes appear to be a prerequisite for these phenotypic alterations. In our study, most patients were treated with preoperative chemotherapy and this may have induced genetic alterations. Although no significant difference was noted when the results of APC analysis and β catenin expression were correlated with the administration of preoperative chemotherapy, we are uncertain about the existence or nature of genetic changes attributable to preoperative chemotherapy in our patients because no prechemotherapy tumour tissue was available for analysis.

In conclusion, our study has shown that allelic imbalance/loss of heterozygosity of the APC locus is a more frequent aberration than microsatellite instability in this cohort of nephroblastomas. Although there was a significant correlation between some of the individual markers and the clinicopathological markers, the overall results do not support a prognostic role for the APC gene in nephroblastoma. In addition, there are no significant correlations between the expression of β catenin and APC microsatellite mutations.

ACKNOWLEDGEMENTS

This study was funded by a grant awarded to DG by the Cancer Association of South Africa.

Authors' affiliations

A Ramburan, D Govender, Molecular Biology Research Facility, Nelson R Mandela School of Medicine, University of KwaZulu-Natal, Congella, 7925 Durban, South Africa

F Oladiran, C Smith, Department of Pathology, Nelson R Mandela School of Medicine, University of KwaZulu-Natal

G P Hadley, Department of Paediatric Surgery, Nelson R Mandela School of Medicine, University of KwaZulu-Natal

REFERENCES

- 1 Su L, Steinbach G, Sawyer J, *et al*. Genomic rearrangements of the APC tumor-suppressor gene in familial adenomatous polyposis. *Hum Genet* 2000;**106**:101-7.
- 2 Rees M, Leigh S, Delhanty J, *et al*. Chromosome 5 allele loss in familial and sporadic colorectal adenomas. *Br J Cancer* 1989;**59**:361-5.
- 3 Bodmer W, Bailey C, Bodmer J, *et al*. Localization of the gene for familial adenomatous polyposis on chromosome 5. *Nature* 1987;**328**:614-16.
- 4 Leppert M, Dobbs M, Scambler P, *et al*. The gene for familial polyposis coli maps to the long arm of chromosome 5. *Science* 1987;**238**:1411-13.
- 5 Beroud C, Soussi T. APC gene: database of germline and somatic mutations in human tumors and cell lines. *Nucleic Acids Res* 1996;**24**:121-4.
- 6 Horrii A, Nakatsuru S, Ichii S, *et al*. Multiple isoforms of the APC gene transcripts and their tissue-specific expression. *Hum Mol Genet* 1993;**2**:283-7.
- 7 Rubinfeld B, Albert I, Porfiri E, *et al*. Loss of beta-catenin regulation by the APC tumor suppressor protein correlates with loss of structure due to common somatic mutations of the gene. *Cancer Res* 1997;**57**:4624-30.
- 8 van Es J, Giles R, Clevers H. The many faces of the tumor suppressor gene APC. *Exp Cell Res* 2001;**264**:126-34.
- 9 Gumbiner B. Signaling transduction of beta-catenin. *Curr Opin Cell Biol* 1995;**7**:634-70.
- 10 Bullions L, Levine A. The role of beta-catenin in cell adhesion, signal transduction, and cancer. *Curr Opin Oncol* 1998;**10**:81-7.
- 11 Aberle H, Bauer A, Stappert J, *et al*. Beta-catenin is a target for the ubiquitin-proteasome pathway. *EMBO J* 1997;**16**:3797-804.
- 12 Rubinfeld B, Albert I, Porfiri E, *et al*. Binding of GSK3- β to the APC- β -catenin complex and regulation of complex assembly. *Science* 1996;**272**:1023-5.
- 13 Nathke I. The adenomatous polyposis coli protein. *Mol Pathol* 1999;**52**:169-73.
- 14 Nesbit C, Tersak J, Prochownik E. MYC oncogenes and human neoplastic disease. *Oncogene* 1999;**18**:3004-16.
- 15 Nishida N, Fukuda Y, Komeda T, *et al*. Amplification and overexpression of the cyclin D1 gene in aggressive human hepatocellular carcinoma. *Cancer Res* 1994;**54**:3107-10.
- 16 Hynes R, Yamada K. Fibronectins: multifunctional modular glucoproteins. *J Cell Biol* 1982;**95**:369-77.
- 17 Hadley G, Jacobs C. The clinical presentation of Wilms' tumour in black children. *S Afr Med* 1990;**77**:565-7.
- 18 Barrantes J, Muir K, Toyn C, *et al*. Thirty-year population-based review of childhood renal tumours with an assessment of prognostic features including tumour DNA characteristics. *Med Pediatr Oncol* 1993;**21**:24-30.
- 19 Faria P, Beckwith J, Mishra K. Focal versus diffuse anaplasia in Wilms' tumor—new definitions with prognostic significance. A report from the National Wilms' tumor study group. *Am J Surg Pathol* 1996;**20**:909-20.
- 20 Ordóñez N, Rosai J. Kidney, renal pelvis and ureter. In: Rosai J, ed. *Ackerman's surgical pathology*. St Louis: Mosby, 1996:465-78.
- 21 Feinberg A. Multiple genetic abnormalities of 11p15 in Wilms' tumour. *Med Pediatr Oncol* 1996;**27**:484-9.
- 22 Reeve A, Sih S, Raizis A. Loss of allelic heterozygosity at a second locus on chromosome 11 in sporadic Wilms' tumor cells. *Nat Rev Mol Cell Biol* 1989;**9**:1799-803.
- 23 Grundy P, Telzerow P, Breslow N, *et al*. Loss of heterozygosity for chromosome 16q and 1p in Wilms' tumors predicts an adverse outcome. *Cancer Res* 1994;**54**:2331-3.
- 24 Grundy P, Pritchard J, Scambler P, *et al*. Loss of heterozygosity on chromosome 16 in sporadic Wilms' tumour. *Br J Cancer* 1998;**78**:1181-7.
- 25 Coppes M, Haber D, Grundy P. Genetic events in the development of Wilms' tumor. *N Engl J Med* 1994;**331**:586-90.
- 26 Bardeesy N, Beckwith J, Pelletier J. Clonal expansion and attenuated apoptosis in Wilms' tumors are associated with p53 gene mutations. *Cancer Res* 1995;**55**:215-19.
- 27 Ramburan A, Chetty R, Hadley G, *et al*. Microsatellite analysis of the DCC gene in nephroblastomas: pathologic correlations and prognostic implications. *Mod Pathol* 2004;**17**:89-95.
- 28 Olansky L, Janssen R, Willing C. Variability of the insulin gene in American blacks with NIDDM. *Diabetes* 1992;**41**:742-9.
- 29 Canzian F, Salovaara R, Hemminki A. Semiautomated assessment of loss of heterozygosity and replication error in tumors. *Cancer Res* 1996;**56**:3331-7.
- 30 Cawkwell L, Li D, Lewis F. Microsatellite instability in colorectal cancer: improved assessment using fluorescent polymerase chain reaction. *Gastroenterology* 1995;**109**:465-71.

- 31 Jass J, Barker M, Fraser L, et al. APC mutation and tumour budding in colorectal cancer. *J Clin Pathol* 2003;**56**:69–73.
- 32 Kinzler K, Vogelstein B. Cancer-susceptibility genes. Gatekeepers and caretakers. *Nature* 1997;**386**:761–83.
- 33 Vogelstein B, Fearon E, Hamilton S, et al. Genetic alterations during colorectal-tumour development. *N Engl J Med* 1988;**319**:525–32.
- 34 Miyoshi Y, Nagase H, Ando H, et al. Somatic mutations of the APC gene in colorectal tumors: mutation cluster region in the APC gene. *Hum Mol Genet* 1992;**1**:229–33.
- 35 Powell S, Zilz N, Beazer-Barclay Y, et al. APC mutations occur early during colorectal tumorigenesis. *Nature* 1992;**359**:235–7.
- 36 Fang D, Luo Y, Yang S, et al. Mutation analysis of APC gene in gastric cancer with microsatellite instability. *World J Gastroenterol* 2002;**8**:787–91.
- 37 Oda H, Imai Y, Nakatsuru Y, et al. Somatic mutations of the APC gene in sporadic hepatoblastomas. *Cancer Res* 1996;**56**:3320–3.
- 38 Ha S, Kim J, Seo E, et al. Low frequency of beta-catenin gene mutations in pilomatricoma. *Acta Derm Venereol* 2002;**82**:428–31.
- 39 Saito T, Oda Y, Sakamoto A, et al. APC mutations in synovial sarcoma. *J Pathol* 2002;**196**:445–9.
- 40 Huang H, Mahler-Araujo B, Sankila A, et al. APC mutations in sporadic medulloblastomas. *Am J Pathol* 2000;**156**:433–7.
- 41 Boynton R, Blount P, Yin J, et al. Loss of heterozygosity involving the APC and MCC genetic loci occurs in the majority of human esophageal cancers. *Proc Natl Acad Sci U S A* 1992;**89**:3385–8.
- 42 Kurahashi H, Takami K, Oue T. Biallelic inactivation of the APC gene in hepatoblastoma. *Cancer Res* 1995;**55**:5007–11.
- 43 Bala S, Wunsch P, Ballhausen W. Childhood hepatocellular adenoma in familial adenomatous polyposis: mutations in adenomatous polyposis coli gene and p53. *Gastroenterology* 1997;**112**:919–22.
- 44 Mannens M, Devilee P, Bliet J, et al. Loss of heterozygosity in Wilms' tumors, studied for six putative tumor suppressor regions, is limited to chromosome 11. *Cancer Res* 1990;**50**:3279–83.
- 45 Devilee P, van den Broek M, Mannens M, et al. Differences in patterns of allelic loss between two common types of adult cancer, breast and colon carcinoma, and Wilms' tumor of childhood. *Int J Cancer* 1991;**47**:817–21.
- 46 Bennet P. Demystified: microsatellites. *Mol Pathol* 2000;**53**:177–83.
- 47 Christians F, Newcomb T, Loeb L. Potential sources of multiple mutations in human cancers. *Prev Med* 1995;**24**:329–32.
- 48 Bardeesy N, Falkoff D, Petrucci M, et al. Anaplastic Wilms' tumor, a subtype displaying poor prognosis, harbors p53 gene mutations. *Nat Genet* 1994;**7**:91–7.
- 49 Chetty R, Naidoo R, Tarin M, et al. Chromosome 2p, 3p 5q and 18q status in sporadic gastric cancer. *Pathology* 2002;**34**:275–81.
- 50 Ogasawara S, Tamura G, Maesawa C, et al. Common deleted region on the long arm of chromosome 5 in esophageal carcinoma. *Gastroenterology* 1996;**110**:52–7.
- 51 D'Amico D, Carbone D, Johnson B, et al. Polymorphic sites within the MCC and APC loci reveal frequent loss of heterozygosity in human small cell lung cancer. *Cancer Res* 1992;**52**:1996–9.
- 52 Ding S, Delhanty J, Dooley J, et al. The putative tumor suppressor gene on chromosome 5q for hepatocellular carcinoma is distinct from the MCC and APC genes. *Cancer Detect Prev* 1993;**17**:405–9.
- 53 Allan G, Cottrell S, Trowsdale J, et al. Loss of heterozygosity on chromosome 5 in sporadic ovarian carcinoma is a late event and is not associated with mutations in APC at 5q21–22. *Hum Mutat* 1994;**3**:283–91.
- 54 Horrigan S, Westbrook C, Kim A, et al. Polymerase chain reaction-based diagnosis of del (5q) in acute myeloid leukemia and myelodysplastic syndrome identifies a minimal deletion interval. *Blood* 1996;**88**:2665–70.
- 55 Ashton-Rickardt P, Wyllie A, Bird C, et al. MCC, a candidate familial polyposis gene in 5q21, shows frequent allele loss in colorectal and lung cancer. *Oncogene* 1991;**6**:1881–6.
- 56 Nakanishi Y, Ochiai A, Akimoto S, et al. Expression of E-cadherin, alpha-catenin, beta-catenin and plakoglobin in esophageal carcinomas and its prognostic significance. *Oncology* 1997;**54**:158–65.
- 57 Pirinen R, Hirvikoski P, Johansson R, et al. Reduced expression of alpha-catenin, beta-catenin and gamma-catenin is associated with high cell proliferative activity in non-small cell lung cancer. *J Clin Pathol* 2001;**54**:391–5.
- 58 Jawhari A, Jordan S, Poole S, et al. Abnormal immunoreactivity of the E-cadherin-catenin complex in gastric carcinoma: relationship with patient survival. *Gastroenterology* 1997;**112**:46–54.
- 59 Krishnadath K, Tilanus H, van Blankenstein M, et al. Reduced expression of the cadherin-catenin complex in oesophageal adenocarcinoma correlates with poor prognosis. *J Pathol* 1997;**182**:331–8.
- 60 Park W, Oh R, Park J, et al. Nuclear localization of beta-catenin is an important prognostic factor in hepatoblastoma. *J Pathol* 2001;**193**:483–90.
- 61 Koesters R, Niggli F, von Knebel Doeberitz M, et al. Nuclear accumulation of beta-catenin protein in Wilms tumours. *J Pathol* 2003;**199**:68–76.
- 62 Alami J, Williams B, Yeger H. Differential expression of E-cadherin and beta catenin in primary and metastatic Wilms's tumours. *J Clin Pathol* 2003;**56**:218–25.
- 63 Saito T, Oda Y, Sakamoto A, et al. Prognostic value of the preserved expression of the E-cadherin and catenin families of adhesion molecules and of beta-catenin mutations in synovial sarcoma. *J Pathol* 2000;**192**:342–50.
- 64 Koesters R, Ridder R, Kopp-Schneider A, et al. Mutational activation of the beta-catenin proto-oncogene is a common event in the development of Wilms' tumors. *Cancer Res* 1999;**59**:3880–2.
- 65 Govender D. The pathology of nephroblastoma. *Curr Diagn Pathol* 2000;**6**:45–54.
- 66 Zuppan CW, Beckwith JB, Weeks DA, et al. The effect of preoperative therapy on the histologic features of Wilms' tumor. An analysis of cases from the third national Wilms' tumor study. *Cancer* 1991;**68**:385–94.
- 67 Brisigotti M, Cozzutto C, Fabbretti G, et al. Wilms' tumor after treatment. *Pediatr Pathol* 1992;**12**:397–406.