REVIEW

Molecular biology of squamous cell carcinoma of the head and neck

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Squamous cell carcinoma of the head and neck (HNSCC) is a heterogeneous but largely preventable disease with complex molecular abnormalities. It arises from a premalignant progenitor followed by outgrowth of clonal populations associated with cumulative genetic alterations and phenotypic progression to invasive malignancy. These genetic alterations result in inactivation of multiple tumour suppressor genes and activation of proto-oncogenes, including p16^{ink4A}, p53, cyclin D1, p14^{ARF}, *FHIT*, *RASSF1A*, epidermal growth factor receptor (EGFR), and Rb. Intramucosal migration and clonal expansion of transformed cells with formation of abnormal genetic fields appear to be responsible for local recurrences and development of second primary tumours.

non-neoplastic mucosa of patients with HNSCC in local tumour recurrences and the development of second primary HNSCCs. We also survey recent progress in the molecular classification of HNSCC provided by genomic arrays, the molecular biology of HNSCC in “young” individuals, and the relation of human papillomavirus (HPV) and HNSCC.

MOLECULAR PATHOLOGY AND TUMOUR PROGRESSION

Genetic alterations and progression of squamous dysplasia to invasive carcinoma

It is generally accepted that HNSCC arises from a common premalignant progenitor followed by outgrowth of clonal populations associated with cumulative genetic alterations and phenotypic progression to invasive malignancy.^{8–10} These genetic alterations result in inactivation of tumour suppressor genes and activation of proto-oncogenes by deletions, point mutations, promoter methylation, and gene amplification (table 1). Microsatellite marker analysis has allowed the delineation of a genetic progression model for HNSCC based on the frequency of these genetic alterations in preinvasive lesions and invasive tumours (fig 1).^{9–10} Loss of chromosomal region 9p21 is found in 70–80% of cases, thus representing the most common genetic alteration seen in squamous dysplasia and HNSCC.^{9–11–12} Loss of heterozygosity (LOH) of 9p21 appears to be an early event in squamous neoplasia of the head and neck and has been found in preneoplastic lesions, including 30% of cases of squamous hyperplasia.^{11–13} The *CDKN2A* gene locus found in chromosome 9p21 encodes two different transcripts, p16 and p14^{ARF}, which are responsible for G₁ cell cycle regulation and MDM2 mediated degradation of p53. P16 is often inactivated in HNSCC through homozygous deletion, by promoter methylation, and less commonly by point mutations.¹⁴

Loss of chromosome region 3p is another common early genetic event in squamous dysplasias and invasive HNSCC.^{15–17} The specific locus responsible for the tumour suppressor phenotype of 3p remains uncharacterised but investigators have identified at least four distinct regions of allelic loss.^{15–18} These regions include 3p14, 3p21, 3p22, 3p24, and 3p26. 3p14 contains the fragile histidine triad gene or *FHIT*, a putative tumour suppressor gene, which has been found

Squamous cell carcinoma of the head and neck (HNSCC) remains a significant cause of morbidity and mortality, with approximately 540 000 new cases annually worldwide and 271 000 deaths for a mortality of 50%.¹ An estimated 4350 new cases of oral and laryngeal carcinoma are diagnosed each year in Canada, with 1560 tumour related deaths giving a mortality of 36%.² In the USA, the use of adjuvant radiotherapy after surgery, and concurrent chemotherapy and radiotherapy, has resulted in improved five year survival rates for advanced carcinomas of the larynx and pharynx but no survival improvement has been seen for oral carcinomas and early laryngeal carcinomas.³ Conventional HNSCC is largely a preventable disease. Major risks factors include tobacco use and alcohol consumption in developed countries and betel quid chewing and bidi smoking in Southeast Asia. The past decade has witnessed significant progress in the understanding of the molecular genetic events underlying the development of HNSCC.^{4–6} The promise of these studies is the development of novel diagnostic and therapeutic procedures which can be used in the clinical management of these patients.⁷

HNSCC is a heterogeneous disease with complex molecular abnormalities. In this review we attempt to summarise the molecular mechanisms underlying certain clinicopathological characteristics of squamous neoplasia in the head and neck observed by surgical pathologists. Our goal is to highlight some of the advances made in the understanding of the molecular progression from dysplasia to invasive carcinoma, and the role played by molecular alterations found in

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Abbreviations: CIS, carcinoma in situ; HNSCC, squamous cell carcinoma of the head and neck; HPV, human papillomavirus; LOH, loss of heterozygosity; SPT, second primary tumour

Figure 1 Hypothetical model for HNSCC carcinogenesis, modified from Califano *et al.*⁹

to be inactivated by exonic deletions in many tumour types including a small percentage of HNSCC.^{19–20} *RSSF1A* is another candidate tumour suppressor gene mapped to 3p21 and found to be inactivated by hypermethylation in a small subset of HNSCC.^{16–21} Much controversy remains regarding the genes present in 3p and their significance and possible roles in HNSCC carcinogenesis. Thus therefore further studies continue.

LOH of 17p and point mutations of the p53 are seen in approximately 50% of HNSCC cases.^{8–22} Most of these mutations appear to occur late in the progression from epithelial dysplasia to invasive carcinoma.^{23–24} In a study including 102 invasive carcinomas, 13 severe dysplasias, and 24 cases of squamous carcinoma in situ (CIS), Boyle *et al.*²³ reported a prevalence of p53 mutations in 43% of the invasive carcinomas and 19% in the non-invasive lesions. In a longitudinal study of 10 patients with sequential biopsies showing progression from hyperplasia or mild dysplasia to CIS or invasive carcinoma, Shahnava *et al.*²⁴ identified p53 mutations in one of 12 dysplasias (8%), two of three CIS cases (66%), and six of eight invasive carcinomas (75%). Nearly 64% of p53 mutations in HNSCC occur in G nucleotides, consistent with exposure to carcinogens such as tobacco.²³

Amplification of 11q13 and overexpression of *cyclin D1* is seen in 30–60% of HNSCC cases and has been associated with an increased rate of lymph node metastases and overall poor prognosis.^{25–27} *Cyclin D1* induces phosphorylation of Rb, thus enabling progression from G₁ to S phase. Both amplification of *cyclin D1* and inactivation of p16 result in increased phosphorylation of Rb and progression in the cell cycle from G₁ to S phase. Amplification and overexpression of *cyclin D1* has been described in up to 40% of cases of oral squamous dysplasia, including mild dysplasia.²⁸

In contrast to some colorectal carcinomas, where microsatellite instability is common, in HNSCC this alteration has been identified in only a small subset of preinvasive lesions of the head and neck.²⁹ El-Naggar *et al.*²⁹ reported it in 15% of a

small group of head and neck dysplasias. In another study, Ha and coworkers³⁰ showed that it becomes increasingly more common as dysplastic lesions progress to HNSCCs.

Gene expression microarrays suggest that most of transcriptional alterations in head and neck carcinogenesis occur during the transition from normal mucosa to premalignant lesions rather than in the transformation from premalignant lesion to invasive carcinoma.³¹ When compared with normal mucosa, premalignant lesions showed 108 upregulated and 226 downregulated genes, whereas invasive carcinomas had five upregulated and 13 downregulated genes when compared with premalignant lesions.³¹

Genetic alterations and risk of progression to malignancy

Leukoplakia and erythroplakia have a well documented risk for transformation into HNSCCs.^{32–33} The risk of malignant transformation increases with the microscopic finding of dysplasia; however, there is significant lack of agreement in the grading of oral dysplasias.³⁴ One of the promises offered by a better understanding of the molecular basis of preneoplastic lesions of the head and neck is the provision of ancillary tools to provide objective and reproducible grading and estimation of risk of evolving to invasive carcinoma. Oral premalignant tissues frequently show many genomic alterations. In a study by Mao *et al.*,¹² 51% of patients whose lesions were investigated for LOH at 9p21 and 3p14 loci showed allelic losses at either or both loci. Of the 19 patients with LOH, seven (37%) developed HNSCC while only one of 18 patients without LOH developed HNSCC. In another study of patients with oral leukoplakia, three biomarkers—chromosomal polysomy, suprabasal p53 expression, and LOH at chromosome 3p or 9p—were found to be the most significant predictors of invasive cancer development, in addition to the presence of moderate/severe dysplasia and a past history of oral cancer.³⁵ Rosin and collaborators¹³ showed differing LOH patterns between non-progressing and progressing low grade oral dysplasias. Both types of lesion were characterised by LOH at 3p or 9p or both; however, patients with progression to in situ or invasive squamous cell carcinomas showed additional losses on 4q, 8p, 11q, or 17p. The same group of investigators have shown that toluidine blue staining identifies premalignant lesions with “high risk” LOH patterns associated with the development of invasive HNSCC.³⁶

Evaluation of DNA ploidy in oral leukoplakia allows the identification of gross genomic alterations and has been shown to be a useful tool in identifying lesions with a high risk of malignant transformation.^{37–38} Sudbø *et al.*³⁷ studied the lesional DNA content in 150 patients with oral leukoplakia and epithelial dysplasia. Thirty six (24%) developed a carcinoma after a mean follow up of 49 months; however,

Table 1 Frequent molecular abnormalities in head and neck squamous cell carcinoma

LOH 9p	70–80%
LOH 3p	60–70%
LOH 17p	50–70%
LOH 11q	30%
LOH 13q	30%
Inactivation of p16 ^{ink4A} (homozygous deletion, promoter methylation, point mutation)	80%
Inactivation of <i>FHIT</i> and <i>RASSF1A</i> p53 mutation	50–80%
Cyclin D1 amplification	30%

only three of 105 diploid cases (3%) develop invasive disease, as compared with 21 of 25 aneuploid cases (84%) and 12 of 20 tetraploid cases (60%). The negative predictive value for diploid lesions was 97% and the positive predictive value for aneuploid lesions was 84%. DNA content appeared to be an independent prognostic factor, with no statistically significant correlation with the grade of dysplasia. In an earlier study, the same investigators analysed ploidy status of 45 leukoplakic lesions histopathologically negative for dysplasia.³⁸ Thirty five lesions (78%) were diploid, five (11%) were tetraploid, and five (11%) were aneuploid. Four of the five aneuploid cases developed carcinoma (80%) whereas only one of the diploid/tetraploid cases (2%) developed malignancy.³⁸ Patients with aneuploid leukoplakia have a 98% rate of developing primary oral carcinoma, an 81% rate of recurrent or second primary tumours, and a 76% risk of death from disease despite negative histology of the surgical margins.³⁹ Thus ploidy status, chromosome polysomy, and LOH analysis of oral precancerous lesions may provide important information about the risk of progression to cancer, second cancer development, and tumour specific death.

FIELD CANCERISATION, MULTIFOCAL DISEASE, AND SECOND MALIGNANCIES

Field cancerisation and second primary tumours

The concept of "field cancerisation" was first introduced in 1953 by Slaughter *et al*⁴⁰ after noting the presence of multiple "independent" tumours and "abnormal epithelium" in the mucosa adjacent to HNSCC. In his now classic study, Slaughter *et al*⁴⁰ described multiple carcinomas in 11.2% of 783 unselected cases of HNSCC. Slaughter postulated that "multicentric origin through a process of field cancerisation would seem to be an important factor in the persistence or recurrence of epidermoid carcinoma following therapy". In its classical interpretation, the concept of "field cancerisation" refers to large areas of the aerodigestive tract mucosa affected by long term exposure to carcinogens, resulting in genetically altered fields in which multifocal carcinomas can develop because of independent genetic events. Many techniques, including chromosome X inactivation, microsatellite analysis, and p53 mutational analysis have confirmed the presence of genetic alterations in the mucosa adjacent to primary HNSCCs, thus providing a genetic explanation for Slaughter's concept of field cancerisation,^{9 41-46} but in contrast to Slaughter's original assertion, these studies strongly suggest that most tumours arising in these abnormal fields are clonally related and originate from a common preneoplastic progenitor. It has been hypothesised that these multiple tumours are caused by the migration of transformed cells through the upper aerodigestive tract mucosa, either by intraepithelial migration or by saliva (micrometastases). The transformed cells have a survival advantage and eventually displace or replace the surrounding mucosa through a process known as "clonal expansion".^{9 46}

The frequency of genetically abnormal fields in mucosa surrounding HNSCC is difficult to ascertain as most studies addressing this problem have been retrospective, but there is evidence suggesting that it is significantly high. Tabor *et al*⁴² found genetically altered fields in the non-neoplastic mucosa of 36% of unselected patients with HNSCC. More importantly, in 70% of these patients, the altered field extended to the surgical margins. Brennan *et al*⁴⁷ also detected p53 mutations in 13 of 25 patients (52%) who appeared to have complete tumour resection on the basis of microscopically negative surgical margins; in a different study Tabor *et al*⁴¹ found p53 mutations in 60% of the specimen margins of 10 patients with multiple HNSCC. The size of the abnormal field is difficult to estimate but it can be large and surely varies

considerably from patient to patient. Tabor *et al*⁴¹ reported six clonally related tumours separated by 3–6 cm, Worsham *et al*⁴⁸ reported two clonally related synchronous carcinomas from the anterior floor of the mouth and pyriform sinus, and Califano *et al*⁴⁹ described a patient with multiple squamous cell carcinomas from the hypopharynx and distal oesophagus showing similar allelic loss separated by 40 cm.

Although there is substantial molecular and genetic evidence supporting the concept of field cancerisation, debate remains over the clonal relation of preneoplastic and neoplastic lesions developing in these fields and their mode of spread.^{50 51} Establishing the clonal relation of multiple tumours in the same patient is not always easy or straightforward. Unrelated tumours may lose the same allele by chance alone, and related tumours may show differing allelotypes owing to the accumulation of new alterations during tumour progression. Tabor *et al*⁴² demonstrated this conundrum when comparing the genetic alterations between the "field" and the tumours in 28 patients with HNSCC, and found that in only 20% of cases did the tumour and the "field" show similar alterations. In the remaining group, 60% of tumours showed additional losses indicative of genetic progression whereas in 40%, the "field" showed alterations not present in the index tumour.⁴² In another study, these group of investigators showed discordant genetic alterations between the primary tumour and the corresponding lymph node metastasis in nine of 20 patients (45%). Therefore, similar patterns of LOH with statistical analysis, identical breakpoints, shared microsatellite shifts, and similar p53 mutations have been recommended as evidence of common clonal origin.^{10 51} Using a combination of current techniques it appears that approximately 60–70% of carcinomas arising in a field originate from a common precursor clone^{41 46 52 53} whereas the remaining appear to represent "true" multiple primary tumours.^{45 52 54}

Braakhuis *et al*⁵¹ have proposed a new classification of secondary HNSCCs arising after treatment of an oral or oropharyngeal carcinoma based on the degree of clonal relation (fig 2). They defined field cancerisation as mucosa with "the presence of one or more areas consisting of epithelial cells that have genetic alterations. A field lesion (or field) has a monoclonal origin, and does not show invasive growth or metastatic behavior".⁶ These investigators proposed that tumours with similar genetic profile should be classified as "recurrence" or a metastasis depending on whether the second lesion occurs in the same or a distant anatomical site. If the individual tumours show different genetic alterations, the secondary lesion should be regarded a "true" second primary tumour (SPT). The third group consists of lesions with a common genetic origin but which diverge in later stages; therefore some show similar allelic imbalances and mutations but others show different genotypes. This latter group is classified as "second field tumours" (SFTs).⁵¹

Molecular pathology of HNSCC and second primary tumours in lung and oesophagus

Approximately 5% of patients with HNSCC develop lung metastases. Given the morphological similarities, the distinction of a primary lung carcinoma from a solitary lung metastasis in a patient with HNSCC cannot be made reliably on histopathological grounds in most cases. To address this problem, Leong *et al*⁵⁵ examined the status of chromosomes 3p and 9p in 16 patients with paired resected primary HNSCC and solitary lung carcinomas. Ten of the 16 patients (63%) showed concordant allelic loss at all loci, consistent with a common clonal origin and strongly supporting the conclusion that the pulmonary tumours represented solitary metastases. Three additional samples had different allelic

Figure 2 Proposed classification by Braakhuis *et al*⁶¹ of second tumours after a primary HNSCC.

losses consistent with an independent tumour origin, suggesting that the lung lesions were second primary tumours. These results suggest that the majority of solitary lung squamous carcinomas in patients with HNSCC are in fact metastases rather than second primaries, with a smaller number representing second primary tumours. The clonal relation of HNSCC and second “primary” oesophageal squamous cell carcinoma has been investigated by Califano *et al*⁴⁹ using microsatellite markers located on chromosomal arms 3p, 9p, and 17p in paired tumours of the head and neck and oesophagus. In 14 cases (83%) the paired tumours had discordant patterns of allelic loss, whereas only two (13%) had identical genetic alterations. Thus most oesophageal carcinomas in patients with HNSCC appear to arise as independent neoplasms with only a small percentage being clonally related.

Detection of minimal disease in surgical margins

Approximately 3.9–32% of patients with HNSCC develop local recurrences despite the presence of microscopically negative margins.^{56–58} Several studies have investigated the presence and clinical implications of molecular alterations in seemingly tumour-free mucosa in the surgical margins of patients undergoing curative surgery for HNSCC.^{47 59 60} Detection of genotypically abnormal cells at the surgical margins of resection may be helpful in identifying individuals with a heightened risk of local recurrences after complete surgical resection. In a seminal study addressing this problem, Brennan *et al*⁴⁷ demonstrated p53 mutations in 13 of 25 patients (52%) who appeared to have complete tumour resection on the basis of microscopically negative surgical margins. Five of these 13 patients developed local recurrences whereas none of the 12 patients without mutations had recurrences of their tumour. In a study by van Houten *et al*⁵⁹ 50 of 76 patients (66%) showed p53 mutations in one or more of the surgical margins regarded as “negative” by conventional histopathology. Nine of the patients with p53 mutations developed local recurrences and four developed regional disease. In the group without p53 mutations, there were no local failures and only one patient developed regional disease.⁵⁹ These investigators found that no other clinicopathological index was predictive of development of local or regional failure. Molecular studies are also helpful in detecting small foci of abnormal mucosa overlooked by conventional histopathological assessment. In a study by van der Toom *et al*,⁶⁰ a combination of p53 immunostaining and

in situ hybridisation for chromosomes 1 and 7 demonstrated the presence of abnormal cells in 11 of 20 margins (55%) diagnosed as tumour-free in their initial microscopic assessment. After examination of the p53 stains and ISH findings, atypical clusters of 10 to 20 cells that had been diagnosed as normal or hyperplastic became evident. All these cases were upgraded to low grade dysplasia with no case being upgraded to high grade dysplasia or carcinoma.

Data from several studies by Nathan *et al*^{61–63} have suggested some prognostic value for overexpression of the proto-oncogene eIF4E in histologically negative margins of HNSCC; eIF4E is a eukaryotic protein synthesis initiation factor increased in almost all HNSCC cases.⁶³ Using immunohistochemistry, Nathan^{61 62} showed statistically significant differences in local recurrence rates and disease-free intervals between eIF4E positive and eIF4E negative margins; patients with eIF4E-positive margins had a 6.5-fold risk of developing local recurrences. eIF4E was reported to be a more sensitive prognostic indicator for local recurrences than p53 immunohistochemistry.

Gene profiling of head and neck squamous cell carcinoma

The development of high throughput, hybridisation based methods such as cDNA and oligonucleotide microarrays allows the simultaneous analysis of gene expression alterations in thousands of genes in HNSCC.^{64–71} These studies have used a variety of experimental designs, tumour types, array platforms, and statistical tools, making direct comparisons of results difficult; however, the potential biological and clinical implications of gene expression signatures of HNSCC in tumour classification, prognosis, and treatment are enormous.

The molecular heterogeneity and complexities of subcellular abnormalities in HNSCC have been clearly highlighted by expression studies; however, most of these studies have been small and need independent confirmation in larger groups of patients. In a study of 17 patients with HNSCC, Belbin *et al*⁶⁷ identified 375 genes that discriminated between two genotypic subtypes of HNSCC. Patients in group I were younger and had more early stage and paradoxically, poorly differentiated tumours with lower prevalence of nodal metastasis than those in group II; however, 38% of patients in group I died of their cancer whereas none of those in group II died as a result of their cancer after a median follow up period of four years. El-Naggar *et al*⁶⁶ analysed 12 matched pairs of normal squamous mucosa and tumour samples from six conventional HNSCC and six variants HNSCC. Parallel expression analysis of 8055 genes showed 26 differentially expressed genes between normal mucosa and tumour specimens. Most of the genes with differential expression were associated with cell differentiation and proliferation. Some of the highly expressed genes in all the tumours included keratin 5, small proline-rich protein 1B (cornifin), S-100 calcium binding protein A8, and keratin 14. Cluster analysis also showed some differences in the overall pattern of expression of certain keratins between conventional HNSCC and variant HNSCC. Conventional HNSCC showed a striking increase in keratins 5 and 14, while none of the variants HNSCC showed changes in those genes. Cornifin was highly expressed in most of the conventional HNSCC but not in the variants. Sarcomatoid and verrucous carcinomas had a pattern of gene expression that was closer to each other than to papillary and basaloid carcinomas. In a different study using a combination of cDNA subtraction and microarray analysis, Villaret *et al*⁶⁹ found overexpression of 13 genes in HNSCC compared with normal mucosa. These genes included keratin 6, keratin 16, cell adhesion related molecules (plakophilin, laminin, and connexin 26), metalloproteinases (stromelysin-2), p53 regulated elements (14-3-3

Figure 3 Unsupervised hierarchical cluster analysis of gene expression of 20 oral squamous cell carcinomas on the basis of 1482 genes. Clustering of gene expression is shown on the left, where each row represents one sequence. Genes are shown on the right besides each row. The dendrogram shows two groups of tumours. Cluster 1 (left) shows patients with lymph node metastasis compared with cluster 2 (right), which contains patients with early disease stage and no lymph node metastases. Red represents overexpression, green represents underexpression, and black represents no change in gene expression in comparison with a baseline sample (universal RNA, control). (Courtesy of Dr Suzanne Kamel-Reid, Toronto, Canada.)

sigma protein), and a S-100 calcium binding protein (CaN19). In a study of 60 HNSCCs, Chung *et al*⁷² identified four distinct subtypes of HNSCC, based on gene expression. These subtypes included tumours with an EGFR pathway signature, a mesenchymal enriched subtype, a normal epithelium-like subtype, and a subtype with high levels of antioxidant enzymes. The first or EGFR pathway subtype was characterised by overexpression of the genes for transforming growth factor α (TGF α), fibroblast growth factor binding protein (FGF-BP), MMK6, bullous pemphigoid antigen 1, P-cadherin, laminin γ 2, and collagen XVII- α , in addition to genes involved in desmosome function (desmoglein 3, desmocollin 2) and keratin 14. The mesenchymal enriched subtype showed high expression of vimentin, syndecan 2, lysyl-oxidase, and four collagen subunits. The normal epithelium-like subtype had the highest expression of microsomal glutathione s-transferase 2 and keratins 15 and 4. The last group included tumours with high expression of glutathione s-transferase M3, thioredoxin reductase 1, and glutathione peroxidase 2 among others. The median follow up period in this study was short, but the investigators reported no significant association between these tumour groups and clinical findings.⁷²

Another line of investigation suggests that gene profiling by cDNA microarrays is capable of identifying upregulated and downregulated genes correlated with tumour recurrences and lymph node metastases (fig 3).^{73–74} In a study of 20 previously untreated patients, Schmalbach *et al*⁷³ found significant differential expression of 101 genes between metastatic and non-metastatic oral and pharyngeal carcinomas. The differential profile included genes related to extracellular matrix, cell adhesion and motility, inflammation, and protease inhibition. Collagen type 11 α -1 (COL11A1) and tissue inhibitor of metalloproteinase 1 (TIMP-1) were upregulated in metastatic tumours, whereas serine protease inhibitor-2 (SERPINB2) was downregulated. In a study of 16 oral squamous cell carcinomas, O'Donnell *et al*⁷⁵ concluded

that metastatic tumours could be differentiated from non-metastatic tumours on the basis of their differential expression of 116 genes. COL2 upregulation was one of the genes associated a “metastatic signature”.⁷⁵ Ginos *et al*⁷⁶ described a “recurrent disease signature” seen in tumours with overexpression of genes implicated in stromal invasion (snail homologue 2, met proto-oncogene), RAS signalling (r-ras homologue), cell surface/extracellular matrix interactions (collagen type IV α 5 and α 6, laminin γ 2, α 3, β 3; integrin α 3, α 6, β 4, β 6) and angiogenesis. All seven patients whose tumours expressed the “recurrent disease signature” failed treatment within 12 months of resection of their primary lesion.⁷⁶ Using a different technology and data analysis, Warner *et al*⁷⁴ found that claudin-1 (CLDN1) overexpression was significantly correlated with lymph node metastases and advanced tumour stage. Roepman and associates⁷⁷ have recently reported a predictor of lymph node metastases of HNSCC based on the differential expression of 102 genes. Among the predictor genes were genes encoding extracellular matrix components, genes involved in cell adhesion, cell death genes, cell growth and maintenance genes, and genes encoding proteins involved in the degradation of the extracellular matrix. The predictor had an initial overall accuracy of 86% and correctly identified 19 of 22 cases on a subsequent validation set. Interestingly, long term storage of tumour samples affected the predictor performance negatively.

HUMAN PAPILLOMAVIRUS AND HNSCC

The causal link between human papillomavirus (HPV) and a subset of HNSCC cases has become established in recent years.^{78–80} HPV DNA has been identified in approximately 15% of HNSCC cases.^{80–83} The most commonly detected HPV in HNSCC is HPV-16, which has been demonstrated in 90–95% of all HPV positive HNSCC cases, followed by HPV-18, HPV-33, and HPV-33.^{83–86} The prevalence of HPV DNA in HNSCC varies according to tumour site. The most common location of HPV associated HNSCC (HPV-HNSCC) is the oropharynx.

Figure 4 Poorly differentiated squamous cell carcinoma of tonsil with no evidence of keratinisation (A). Polymerase chain reaction products of DNA extracted from formalin fixed, paraffin embedded tissue show the presence of HPV-16 DNA in lanes 3 to 5 (B). Strong immunohistochemical expression of p16 in same cases (C) with markedly reduced expression of Rb protein (D).

HPV is especially common in carcinomas of the lingual and palatine tonsils, where its reported prevalence has ranged from 45% to 67%.^{80-83 87 88} A detailed review of HPV infections and tonsillar carcinoma by Syrjänen has been published recently in this journal.⁸⁹ HPV DNA has been found less often in squamous cell carcinomas of the oral cavity (12–18%), tongue (12%), hypopharynx (13–25%), and larynx (3–7%).⁸² Epidemiological and case matched studies have also shown that individuals who are seropositive for HPV-16 are at an increased risk for the development of oral or oropharyngeal squamous cell carcinomas.^{79 86 90} After adjustment for age, sex, and serum cotinine levels, HPV-16 seropositive individuals have a greater risk of development of oral squamous cell carcinomas (odds ratio 2.8) and oropharyngeal squamous cell carcinomas (odds ratio 14.4) than seronegative individuals.⁷⁹

There is a growing body of evidence suggesting that oropharyngeal HPV-HNSCC is a distinctive clinicopathological and molecular entity.^{82 91} The risk factors for HPV-HNSCC appear to be different from those of non-HPV HNSCC. Although not universally reported, patients with HPV-HNSCC are less likely to have a history of tobacco use, are more likely to be diagnosed at a younger age, and have a

higher frequency of cervical lymph node metastases at presentation. They are also more likely to report a high lifetime number of heterosexual partners, young age at first intercourse, and a history of orogenital sex.^{85 86 90 91} Most tonsillar HPV-HNSCCs are associated with poorly differentiated, non-keratinising, basaloid morphology, wild-type p53 status, p16^{INK4A} overexpression, and decreased cyclin D1 and Rb expression.^{80 92-96} High risk oncogenic HPVs such as HPV-16 and HPV-18 encode two oncoproteins, E6 and E7, which induce cellular transformation and dysregulation of cell cycle control. E7 interacts and induces proteolytic degradation of pRb and other RB related “pocket” proteins, whereas E6 inactivates p53 by accelerating its ubiquitin mediated proteolysis.⁹⁷ In accordance with the oncogenic properties of E6, approximately 90% of HPV-HNSCCs have wild type p53 and lack immunohistochemical overexpression of p53.^{80 92 98 99} Given the ability of E7 to target Rb for degradation, Rb expression is decreased in HPV-HNSCC (fig 4).^{92 100} Absence of p16 mutations and overexpression of p16^{INK4A} are also molecular hallmarks of HPV-HNSCC.⁹⁹⁻¹⁰² It has been suggested that overexpression of p16 may be used as a surrogate marker to identify HPV-HNSCC, and when

detected in lymph node metastases is a reliable way to establish an oropharyngeal origin.^{96–103}

There is strong evidence suggesting that patients with HPV-HNSCC have a more favourable prognosis with lower disease specific mortality than those with HPV-negative HNSCC.^{80–84–104–105} Gillison *et al*⁸⁰ reported a 59% reduction in risk of death from cancer in patients with HPV-HNSCC compared with HPV-negative HNSCC. The detection of HPV in HNSCC has potentially significant therapeutic implications, given the development of vaccines targeting HPV oncogenic proteins E6 and E7 for the prevention of HPV related malignancies.¹⁰⁶

MOLECULAR BIOLOGY OF SQUAMOUS CELL CARCINOMA IN YOUNG INDIVIDUALS

Unlike their counterparts in older patients, HNSCC affecting young individuals show an increased incidence in females with no known risk factors, and occur more frequently in the oral cavity and hypopharynx.^{107–109} The influence of young age on survival has been controversial, with some investigators reporting improved clinical outcome in comparison with older patients,¹¹⁰ whereas most have found no differences on survival.^{107–109} Such clinicopathological differences suggest that these tumours represent a biologically distinct entity; however, only a few studies have focused on the molecular biology of HNSCC arising in young individuals. In a study comparing “young” and “old” patients, Wang *et al*¹⁰⁸ found a significant greater frequency of microsatellite instability in the young group (88% *v* 36%). Microsatellite instability was also detected at a higher frequency in tongue (71%) than in laryngeal cancers (23%). The mechanism of microsatellite instability in these tumours was unclear as there were no mutations or promoter methylation of mismatch repair (MMR) genes.

With the possible exception of Fanconi anaemia, the role of HPV in the pathogenesis of HNSCC of young individuals appears to be related to anatomical site of the tumour rather than to age on its own. Sisk and collaborators¹¹¹ compared the prevalence of HPV between “young” and “old” patients but found no significant differences in the presence of HPV DNA in these two groups. El-Mofty and Lu⁹³ investigated the prevalence of HPV-16 in 33 HNSCCs from young patients. They found a high prevalence of HPV-16 in tonsillar carcinomas (91%). No HPV was found in any of the oral carcinomas and only two of seven laryngeal carcinomas showed HPV DNA.

Fanconi anaemia is an autosomal recessive genomic instability syndrome associated with congenital abnormalities, bone marrow failure, and a predisposition to the development of acute myeloid leukaemia and other solid cancers.¹¹² The relative risk of developing HNSCC in individuals with Fanconi anaemia is 700 times greater than in the general population, the median age at diagnosis of their carcinoma being 26 years.^{113–114} The risk is greater in patients undergoing haematopoietic stem cell transplantation and in those with acute and chronic graft-versus-host diseases.¹¹⁵ The molecular mechanisms involved in Fanconi anaemia associated head and neck carcinogenesis are not completely understood; however, Kutler *et al*¹¹⁶ found HPV-16 DNA in a high proportion of HNSCCs affecting individuals with this condition. As is the case with other HPV related HNSCCs, these tumours lacked p53 mutations. These findings support a significant role for HPV in the pathogenesis of HNSCCs arising in the context of Fanconi anaemia.¹⁰⁶ The role played by the Fanconi anaemia/BRCA pathway in sporadic HNSCCs, particularly in young individuals with no known risk factors, has not been widely explored; however, in a recent study, Marsit *et al*¹⁰⁰ demonstrated promoter

methylation of the *FANCF* gene in 15% of sporadic HNSCC cases.

CONCLUSION

More traditional molecular studies and genomic arrays will afford us the opportunity for molecular classification of HNSCCs, which will ultimately provide additional insights into HNSCC and possibly new targeted molecular treatments. Despite all these exciting advances, the most effective and cost-efficient tools in reducing the worldwide morbidity and mortality caused by HNSCC are public health policies aimed at reducing or eliminating the use of tobacco products.

Concerns have been recently raised regarding the validity of the results and conclusions regarded in references 37, 38 and 39.

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REFERENCES

- 1 Stewart BW, Kleihues P. *World Cancer Report*. Geneva: International Agency for Research on Cancer, 2003:232–6.
- 2 Canadian Cancer Society. Canadian Cancer Statistics 2005 (www.nccic.ca/cancer.ca).
- 3 Carvalho AL, Nishimoto IN, Califano JA, *et al*. Trends in incidence and prognosis for head and neck cancer in the United States: a site-specific analysis of the SEER database. *Int J Cancer* 2005;**114**:806–16.
- 4 Forastiere A, Koch W, Trotti A, *et al*. Head and neck cancer. *N Engl J Med* 2001;**345**:1890–900.
- 5 Williams HK. Molecular pathogenesis of oral squamous carcinoma. *Mol Pathol* 2000;**53**:165–72.
- 6 Braakhuis BJ, Tabor MP, Kummer JA, *et al*. A genetic explanation of Slaughter's concept of field cancerization: evidence and clinical implications. *Cancer Res* 2003;**63**:1727–30.
- 7 Grandis JR, Pietenpol JA, Greenberger JS, *et al*. Head and neck cancer: meeting summary and research opportunities. *Cancer Res* 2004;**64**:8126–9.
- 8 Nawroz H, van der Riet P, Hruban RH, *et al*. Allelotype of head and neck squamous cell carcinoma. *Cancer Res* 1994;**54**:1152–5.
- 9 Califano J, van der RP, Westra W, *et al*. Genetic progression model for head and neck cancer: implications for field cancerization. *Cancer Res* 1996;**56**:2488–92.
- 10 Califano J, Westra WH, Meininger G, *et al*. Genetic progression and clonal relationship of recurrent premalignant head and neck lesions. *Clin Cancer Res* 2000;**6**:347–52.
- 11 van der Riet P, Nawroz H, Hruban RH, *et al*. Frequent loss of chromosome 9p21–22 early in head and neck cancer progression. *Cancer Res* 1994;**54**:1156–8.
- 12 Mao L, Lee JS, Fan YH, *et al*. Frequent microsatellite alterations at chromosomes 9p21 and 3p14 in oral premalignant lesions and their value in cancer risk assessment. *Nat Med* 1996;**2**:682–5.
- 13 Rosin MP, Cheng X, Poh C, *et al*. Use of allelic loss to predict malignant risk for low-grade oral epithelial dysplasia. *Clin Cancer Res* 2000;**6**:357–62.
- 14 Reed AL, Califano J, Cairns P, *et al*. High frequency of p16 (CDKN2/MTS-1/INK4A) inactivation in head and neck squamous cell carcinoma. *Cancer Res* 1996;**56**:3630–3.
- 15 Garnis C, Baldwin C, Zhang L, *et al*. Use of complete coverage array comparative genomic hybridization to define copy number alterations on chromosome 3p in oral squamous cell carcinomas. *Cancer Res* 2003;**63**:8582–5.
- 16 Hogg RP, Honorio S, Martinez A, *et al*. Frequent 3p allele loss and epigenetic inactivation of the RASSF1A tumour suppressor gene from region 3p21.3 in head and neck squamous cell carcinoma. *Eur J Cancer* 2002;**38**:1585–92.
- 17 Rowley H, Jones A, Spandidos D, *et al*. Definition of a tumor suppressor gene locus on the short arm of chromosome 3 in squamous cell carcinoma of the head and neck by means of microsatellite markers. *Arch Otolaryngol Head Neck Surg* 1996;**122**:497–501.
- 18 Masayeva BG, Ha P, Garrett-Mayer E, *et al*. Gene expression alterations over large chromosomal regions in cancers include multiple genes unrelated to malignant progression. *Proc Natl Acad Sci USA* 2004;**101**:8715–20.
- 19 Kisielewski AE, Xiao GH, Liu SC, *et al*. Analysis of the FHIT gene and its product in squamous cell carcinomas of the head and neck. *Oncogene* 1998;**17**:83–91.
- 20 Mao L, Fan YH, Lotan R, *et al*. Frequent abnormalities of FHIT, a candidate tumor suppressor gene, in head and neck cancer cell lines. *Cancer Res* 1996;**56**:5128–31.

- 21 **Dong SM**, Sun DI, Benoit NE, *et al*. Epigenetic inactivation of RASSF1A in head and neck cancer. *Clin Cancer Res* 2003;**9**:3635–40.
- 22 **van Houten VM**, Tabor MP, van den Brekel MW, *et al*. Mutated p53 as a molecular marker for the diagnosis of head and neck cancer. *J Pathol* 2002;**198**:476–86.
- 23 **Boyle JO**, Hakim J, Koch W, *et al*. The incidence of p53 mutations increases with progression of head and neck cancer. *Cancer Res* 1993;**53**:4477–80.
- 24 **Shahnava SA**, Regezi JA, Bradley G, *et al*. p53 gene mutations in sequential oral epithelial dysplasias and squamous cell carcinomas. *J Pathol* 2000;**190**:417–22.
- 25 **Meredith SD**, Levine PA, Burns JA, *et al*. Chromosome 11q13 amplification in head and neck squamous cell carcinoma. Association with poor prognosis. *Arch Otolaryngol Head Neck Surg* 1995;**121**:790–4.
- 26 **Michalides R**, van Veelen N, Hart A, *et al*. Overexpression of Cyclin D1 correlates with recurrence in a group of forty-seven operable squamous cell carcinomas of the head and neck. *Cancer Research* 1995;**55**:975–8.
- 27 **Maruya S**, Issa JP, Weber RS, *et al*. Differential methylation status of tumor-associated genes in head and neck squamous carcinoma: incidence and potential implications. *Clin Cancer Res* 2004;**10**:3825–30.
- 28 **Rousseau A**, Lim MS, Lin Z, *et al*. Frequent cyclin D1 gene amplification and protein overexpression in oral epithelial dysplasias. *Oral Oncol* 2001;**37**:268–75.
- 29 **El-Naggar AK**, Hurr K, Huff V, *et al*. Microsatellite instability in preinvasive head and neck squamous carcinoma. *Am J Pathol* 1996;**148**:2067–72.
- 30 **Ha PK**, Pilkington TA, Westra WH, *et al*. Progression of microsatellite instability from premalignant lesions to tumors of the head and neck. *Int J Cancer* 2002;**102**:615–17.
- 31 **Ha PK**, Benoit NE, Yochem R, *et al*. A transcriptional progression model for head and neck cancer. *Clin Cancer Res* 2003;**9**:3058–64.
- 32 **Hellquist H**, Lundgren J, Olofsson J. Hyperplasia, keratosis, dysplasia and carcinoma in situ of the vocal cords. Follow-up study. *Clin Otolaryngol* 1982;**7**:11–27.
- 33 **Silverman S Jr**, Gorsky M, Lozada F. Oral leukoplakia and malignant transformation. A follow-up study of 257 patients. *Cancer* 1984;**53**:563–8.
- 34 **Brothwell DJ**, Lewis DW, Bradley G, *et al*. Observer agreement in the grading of oral epithelial dysplasia. *Community Dent Oral Epidemiol* 2003;**31**:300–5.
- 35 **Lee JJ**, Hong WK, Hittelman WN, *et al*. Predicting cancer development in oral leukoplakia: ten years of translational research. *Clin Cancer Res* 2000;**6**:1702–10.
- 36 **Zhang L**, Williams M, Poh CF, *et al*. Toluidine blue staining identifies high-risk primary oral premalignant lesions with poor outcome. *Cancer Res* 2005;**65**:8017–21.
- 37 **Sudbo J**, Kildal W, Risberg B, *et al*. DNA content as a prognostic marker in patients with oral leukoplakia. *N Engl J Med* 2001;**344**:1270–8.
- 38 **Sudbo J**, Ried T, Bryne M, *et al*. Abnormal DNA content predicts the occurrence of carcinomas in non-dysplastic oral white patches. *Oral Oncol* 2001;**37**:558–65.
- 39 **Sudbo J**, Lippman SM, Lee JJ, *et al*. The influence of resection and aneuploidy on mortality in oral leukoplakia. *N Engl J Med* 2004;**350**:1405–13.
- 40 **Slaughter DP**, Southwick HW. "Field cancerization" in oral stratified squamous epithelium. Clinical implications of multicentric origin. *Cancer* 1953;**6**:963–8.
- 41 **Tabor MP**, Brakenhoff RH, Ruijter-Schippers HJ, *et al*. Multiple head and neck tumors frequently originate from a single preneoplastic lesion. *Am J Pathol* 2002;**161**:1051–60.
- 42 **Tabor MP**, Brakenhoff RH, van Houten VM, *et al*. Persistence of genetically altered fields in head and neck cancer patients: biological and clinical implications. *Clin Cancer Res* 2001;**7**:1523–32.
- 43 **Tabor MP**, Brakenhoff RH, Ruijter-Schippers HJ, *et al*. Genetically altered fields as origin of locally recurrent head and neck cancer: a retrospective study. *Clin Cancer Res* 2004;**10**:3607–13.
- 44 **Lydiatt WM**, Anderson PE, Bazzano T, *et al*. Molecular support for field cancerization in the head and neck. *Cancer* 1998;**82**:1376–80.
- 45 **van Oijen MG**, Leppers Vd Straat FG, Tilanus MG, *et al*. The origins of multiple squamous cell carcinomas in the aerodigestive tract. *Cancer* 2000;**88**:884–93.
- 46 **Bedi GC**, Westra WH, Gabrielson E, *et al*. Multiple head and neck tumors: evidence for a common clonal origin. *Cancer Res* 1996;**56**:2484–7.
- 47 **Brennan JA**, Mao L, Hruban RH, *et al*. Molecular assessment of histopathological staging in squamous-cell carcinoma of the head and neck. *N Engl J Med* 1995;**332**:429–35.
- 48 **Worsham MJ**, Wolman SR, Carey TE, *et al*. Common clonal origin of synchronous primary head and neck squamous cell carcinomas: analysis by tumor karyotypes and fluorescence in situ hybridization. *Hum Pathol* 1995;**26**:251–61.
- 49 **Califano J**, Leong PL, Koch WM, *et al*. Second esophageal tumors in patients with head and neck squamous cell carcinoma: an assessment of clonal relationships. *Clin Cancer Res* 1999;**5**:1862–7.
- 50 **van Oijen MG**, Slootweg PJ. Oral field cancerization: carcinogen-induced independent events or micrometastatic deposits? *Cancer Epidemiol Biomarkers Prev* 2000;**9**:249–56.
- 51 **Braakhuis BJ**, Tabor MP, Leemans CR, *et al*. Second primary tumors and field cancerization in oral and oropharyngeal cancer: molecular techniques provide new insights and definitions. *Head Neck* 2002;**24**:198–206.
- 52 **Scholes AG**, Woolgar JA, Boyle MA, *et al*. Synchronous oral carcinomas: independent or common clonal origin? *Cancer Res* 1998;**58**:2003–6.
- 53 **Partridge M**, Pateronichelakis S, Phillips E, *et al*. Profiling clonality and progression in multiple premalignant and malignant oral lesions identifies a subgroup of cases with a distinct presentation of squamous cell carcinoma. *Clin Cancer Res* 2001;**7**:1860–6.
- 54 **Jang SJ**, Chiba I, Hirai A, *et al*. Multiple oral squamous epithelial lesions: are they genetically related? *Oncogene* 2001;**20**:2235–42.
- 55 **Leong PP**, Rezaei B, Koch WM, *et al*. Distinguishing second primary tumors from lung metastases in patients with head and neck squamous cell carcinoma. *J Natl Cancer Inst* 1998;**90**:972–7.
- 56 **Slootweg PJ**, Hordijk GJ, Schade Y, *et al*. Treatment failure and margin status in head and neck cancer. A critical view on the potential value of molecular pathology. *Oral Oncol* 2002;**38**:500–3.
- 57 **Looser KG**, Shah JP, Strong EW. The significance of "positive" margins in surgically resected epidermoid carcinomas. *Head Neck Surg* 1978;**1**:107–11.
- 58 **Chen TY**, Emrich LJ, Driscoll DL. The clinical significance of pathological findings in surgically resected margins of the primary tumor in head and neck carcinoma. *Int J Radiat Oncol Biol Phys* 1987;**13**:833–7.
- 59 **van Houten VM**, Leemans CR, Kummer JA, *et al*. Molecular diagnosis of surgical margins and local recurrence in head and neck cancer patients: a prospective study. *Clin Cancer Res* 2004;**10**:3614–20.
- 60 **van der Toorn PP**, Veltman JA, Bot FJ, *et al*. Mapping of resection margins of oral cancer for p53 overexpression and chromosome instability to detect residual (pre)malignant cells. *J Pathol* 2001;**193**:66–72.
- 61 **Nathan CA**, Amirghahri N, Rice C, *et al*. Molecular analysis of surgical margins in head and neck squamous cell carcinoma patients. *Laryngoscope* 2002;**112**:2129–40.
- 62 **Nathan CA**, Sanders K, Abreo FW, *et al*. Correlation of p53 and the proto-oncogene *elF4E* in larynx cancers: prognostic implications. *Cancer Res* 2000;**60**:3599–604.
- 63 **Nathan CA**, Franklin S, Abreo FW, *et al*. Analysis of surgical margins with the molecular marker *elF4E*: a prognostic factor in patients with head and neck cancer. *J Clin Oncol* 1999;**17**:2909–14.
- 64 **Alevizos I**, Mahadevappa M, Zhang X, *et al*. Oral cancer in vivo gene expression profiling assisted by laser capture microdissection and microarray analysis. *Oncogene* 2001;**20**:6196–204.
- 65 **Leethanakul C**, Patel V, Gillespie J, *et al*. Distinct pattern of expression of differentiation and growth-related genes in squamous cell carcinomas of the head and neck revealed by the use of laser capture microdissection and cDNA arrays. *Oncogene* 2000;**19**:3220–4.
- 66 **el Naggar AK**, Kim HW, Clayman GL, *et al*. Differential expression profiling of head and neck squamous carcinoma: significance in their phenotypic and biological classification. *Oncogene* 2002;**21**:8206–19.
- 67 **Belbin TJ**, Singh B, Barber I, *et al*. Molecular classification of head and neck squamous cell carcinoma using cDNA microarrays. *Cancer Res* 2002;**62**:1184–90.
- 68 **Mendez E**, Cheng C, Farwell DG, *et al*. Transcriptional expression profiles of oral squamous cell carcinomas. *Cancer* 2002;**95**:1482–94.
- 69 **Villaret DB**, Wang T, Dillon D, *et al*. Identification of genes overexpressed in head and neck squamous cell carcinoma using a combination of complementary DNA subtraction and microarray analysis. *Laryngoscope* 2000;**110**:374–81.
- 70 **Choi P**, Chen C. Genetic expression profiles and biologic pathway alterations in head and neck squamous cell carcinoma. *Cancer* 2005;**104**:1113–28.
- 71 **Baker H**, Patel V, Malinolo AA, *et al*. Proteome-wide analysis of head and neck squamous cell carcinomas using laser-capture microdissection and tandem mass spectrometry. *Oral Oncol* 2005;**41**:183–99.
- 72 **Chung CH**, Parker JS, Karaca G, *et al*. Molecular classification of head and neck squamous cell carcinomas using patterns of gene expression. *Cancer Cell* 2004;**5**:489–500.
- 73 **Schmalbach CE**, Chepeha DB, Giordano TJ, *et al*. Molecular profiling and the identification of genes associated with metastatic oral cavity/pharynx squamous cell carcinoma. *Arch Otolaryngol Head Neck Surg* 2004;**130**:295–302.
- 74 **Warner GC**, Reis PP, Jurisica I, *et al*. Molecular classification of oral cancer by cDNA microarrays identifies overexpressed genes correlated with nodal metastasis. *Int J Cancer* 2004;**110**:857–68.
- 75 **O'Donnell RK**, Kupferman M, Wei SJ, *et al*. Gene expression signature predicts lymphatic metastasis in squamous cell carcinoma of the oral cavity. *Oncogene* 2005;**24**:1244–51.
- 76 **Ginos MA**, Page GP, Michalowicz BS, *et al*. Identification of a gene expression signature associated with recurrent disease in squamous cell carcinoma of the head and neck. *Cancer Res* 2004;**64**:55–63.
- 77 **Roepman P**, Wessels LF, Kettelarij N, *et al*. An expression profile for diagnosis of lymph node metastases from primary head and neck squamous cell carcinomas. *Nat Genet* 2005;**37**:182–6.
- 78 **Herrero R**. Human papillomavirus and cancer of the upper aerodigestive tract. *J Natl Cancer Inst Monogr*, 2003;47–51.
- 79 **Mork J**, Lie AK, Glatte E, *et al*. Human papillomavirus infection as a risk factor for squamous-cell carcinoma of the head and neck. *N Engl J Med* 2001;**344**:1125–31.
- 80 **Gillison ML**, Koch WM, Capone RB, *et al*. Evidence for a causal association between human papillomavirus and a subset of head and neck cancers. *J Natl Cancer Inst* 2000;**92**:709–20.
- 81 **Fourt P**, Martin F, Flahault A, *et al*. Human papillomavirus infection in the malignant and premalignant head and neck epithelium. *Diagn Mol Pathol* 1995;**4**:122–7.
- 82 **Paz IB**, Cook N, Odom-Maryon T, *et al*. Human papillomavirus (HPV) in head and neck cancer. An association of HPV 16 with squamous cell carcinoma of Waldeyer's tonsillar ring. *Cancer* 1997;**79**:595–604.
- 83 **Ringstrom E**, Peters E, Hasegawa M, *et al*. Human papillomavirus type 16 and squamous cell carcinoma of the head and neck. *Clin Cancer Res* 2002;**8**:3187–92.

- 84 **Schwartz SR**, Yueh B, McDougall JK, *et al*. Human papillomavirus infection and survival in oral squamous cell cancer: a population-based study. *Otolaryngol Head Neck Surg* 2001;**125**:1–9.
- 85 **Smith EM**, Ritchie JM, Summersgill KF, *et al*. Age, sexual behavior and human papillomavirus infection in oral cavity and oropharyngeal cancers. *Int J Cancer* 2004;**108**:766–72.
- 86 **Herrero R**, Castellsague X, Pawlita M, *et al*. Human papillomavirus and oral cancer: the International Agency for Research on Cancer multicenter study. *J Natl Cancer Inst* 2003;**95**:1772–83.
- 87 **Klussmann JP**, Weissenborn SJ, Wieland U, *et al*. Human papillomavirus-positive tonsillar carcinomas: a different tumor entity? *Med Microbiol Immunol (Berl)* 2003;**192**:129–32.
- 88 **Strome SE**, Savva A, Brissett AE, *et al*. Squamous cell carcinoma of the tonsils: a molecular analysis of HPV associations. *Clin Cancer Res* 2002;**8**:1093–100.
- 89 **Syrjanen S**. HPV infections and tonsillar carcinoma. *J Clin Pathol* 2004;**57**:449–55.
- 90 **Dahlstrom KR**, Adler-Storthz K, Etzel CJ, *et al*. Human papillomavirus type 16 infection and squamous cell carcinoma of the head and neck in never-smokers: a matched pair analysis. *Clin Cancer Res* 2003;**9**:2620–6.
- 91 **Gillison ML**. Human papillomavirus-associated head and neck cancer is a distinct epidemiologic, clinical, and molecular entity. *Semin Oncol* 2004;**31**:744–54.
- 92 **Wilczynski SP**, Lin BT, Xie Y, *et al*. Detection of human papillomavirus DNA and oncoprotein overexpression are associated with distinct morphological patterns of tonsillar squamous cell carcinoma. *Am J Pathol* 1998;**152**:145–56.
- 93 **el Mofly SK**, Lu DW. Prevalence of human papillomavirus type 16 DNA in squamous cell carcinoma of the palatine tonsil, and not the oral cavity, in young patients: a distinct clinicopathologic and molecular disease entity. *Am J Surg Pathol* 2003;**27**:1463–70.
- 94 **Poetsch M**, Lorenz G, Bankau A, *et al*. Basaloid in contrast to nonbasaloid head and neck squamous cell carcinomas display aberrations especially in cell cycle control genes. *Head Neck* 2003;**25**:904–10.
- 95 **Li W**, Thompson CH, Cossart YE, *et al*. The expression of key cell cycle markers and presence of human papillomavirus in squamous cell carcinoma of the tonsil. *Head Neck* 2004;**26**:1–9.
- 96 **Begum S**, Gillison ML, Ansari-Lari MA, *et al*. Detection of human papillomavirus in cervical lymph nodes: a highly effective strategy for localizing site of tumor origin. *Clin Cancer Res* 2003;**9**:6469–75.
- 97 **Munger K**, Howley PM. Human papillomavirus immortalization and transformation functions. *Virus Res* 2002;**89**:213–28.
- 98 **Dai M**, Clifford GM, Le Calvez F, *et al*. Human Papillomavirus Type 16 and TP53 Mutation in Oral Cancer: Matched Analysis of the IARC Multicenter Study. *Cancer Res* 2004;**64**:468–71.
- 99 **Hafkamp HC**, Speel EJ, Haesevoets A, *et al*. A subset of head and neck squamous cell carcinomas exhibits integration of HPV 16/18 DNA and overexpression of p16INK4A and p53 in the absence of mutations in p53 exons 5–8. *Int J Cancer* 2003;**107**:394–400.
- 100 **Wiest T**, Schwarz E, Enders C, *et al*. Involvement of intact HPV16 E6/E7 gene expression in head and neck cancers with unaltered p53 status and perturbed pRb cell cycle control. *Oncogene* 2002;**21**:1510–17.
- 101 **Olshan AF**, Weissler MC, Pei H, *et al*. Alterations of the p16 gene in head and neck cancer: frequency and association with p53, PRAD-1 and HPV. *Oncogene* 1997;**14**:811–18.
- 102 **Klussmann JP**, Gultekin E, Weissenborn SJ, *et al*. Expression of p16 protein identifies a distinct entity of tonsillar carcinomas associated with human papillomavirus. *Am J Pathol* 2003;**162**:747–53.
- 103 **Begum S**, Cao D, Gillison M, *et al*. Tissue distribution of human papillomavirus 16 DNA integration in patients with tonsillar carcinoma. *Clin Cancer Res* 2005;**11**:5694–9.
- 104 **Li W**, Thompson CH, O'Brien CJ, *et al*. Human papillomavirus positivity predicts favourable outcome for squamous carcinoma of the tonsil. *Int J Cancer* 2003;**106**:553–8.
- 105 **Ritchie JM**, Smith EM, Summersgill KF, *et al*. Human papillomavirus infection as a prognostic factor in carcinomas of the oral cavity and oropharynx. *Int J Cancer* 2003;**104**:336–44.
- 106 **Devaraj K**, Gillison ML, Wu TC. Development of HPV vaccines for HPV-associated head and neck squamous cell carcinoma. *Crit Rev Oral Biol Med* 2003;**14**:345–62.
- 107 **Verschuur HP**, Irish JC, O'Sullivan B, *et al*. A matched control study of treatment outcome in young patients with squamous cell carcinoma of the head and neck. *Laryngoscope* 1999;**109**:249–58.
- 108 **Wang Y**, Irish J, MacMillan C, *et al*. High frequency of microsatellite instability in young patients with head-and-neck squamous-cell carcinoma: lack of involvement of the mismatch repair genes hMLH1 AND hMSH2. *Int J Cancer* 2001;**93**:353–60.
- 109 **Pitman KT**, Johnson JT, Wagner RL, *et al*. Cancer of the tongue in patients less than forty. *Head Neck* 2000;**22**:297–302.
- 110 **Annertz K**, Anderson H, Biorklund A, *et al*. Incidence and survival of squamous cell carcinoma of the tongue in Scandinavia, with special reference to young adults. *Int J Cancer* 2002;**101**:95–9.
- 111 **Sisk EA**, Bradford CR, Jacob A, *et al*. Human papillomavirus infection in “young” versus “old” patients with squamous cell carcinoma of the head and neck. *Head Neck* 2000;**22**:649–57.
- 112 **D'Andrea AD**, Grompe M. The Fanconi anaemia/BRCA pathway. *Nat Rev Cancer* 2003;**3**:23–34.
- 113 **Kutler DI**, Auerbach AD, Satagopan J, *et al*. High incidence of head and neck squamous cell carcinoma in patients with Fanconi anemia. *Arch Otolaryngol Head Neck Surg* 2003;**129**:106–12.
- 114 **Alter BP**, Joenje H, Oostra AB, *et al*. Fanconi anemia: adult head and neck cancer and hematopoietic mosaicism. *Arch Otolaryngol Head Neck Surg* 2005;**131**:635–9.
- 115 **Rosenberg PS**, Socie G, Alter BP, *et al*. Risk of head and neck squamous cell cancer and death in patients with Fanconi anemia who did and did not receive transplants. *Blood* 2005;**105**:67–73.
- 116 **Kutler DI**, Wreesmann VB, Goberdhan A, *et al*. Human papillomavirus DNA and p53 polymorphisms in squamous cell carcinomas from Fanconi anemia patients. *J Natl Cancer Inst* 2003;**95**:1718–21.