A CULTURE MEDIUM FOR TRICHOMONAS VAGINALIS DONNÉ AND SPECIES OF CANDIDA

BY

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The laboratory methods usually employed for detecting Trichomonas vaginalis in secretions are the microscopic examination of wet films or fixed, stained smears, and the culture of the parasites in artificial media. In recent years the cultural method has become more widely used as an aid in the diagnosis of trichomoniases. Different culture media have been employed for this purpose with varying degrees of success, and this paper describes a medium which seems particularly suitable, because it is cheap, easy to prepare and use, and keeps well.

During the past 40 years many attempts have been made to grow T. vaginalis in a variety of media. At first simple liquid bacteriological media were used (Lynch, 1915; Reuling, 1921; Fischer, 1935, for example). Richer media of the type used by Boeck and Drbohlav (1925) have also given good results (Andrews, 1929; Hawes, 1947; Whittington, 1951, for example). All these media had the disadvantage of favouring the growth of the accompanying bacteria as well as that of the trichomonads. Johnson and Trussell (1943) and Sprince and Kuperberg (1947) used elaborate media for maintaining bacteria-free cultures of T. vaginalis. The value of these media for diagnostic purposes is unknown, however, and they are too complex and expensive for ordinary routine work. McEntegart (1952) simplified Johnson and Trussell's (1943) formula for use in his studies on T. vaginalis, and Feinberg (1953) substantially modified McEntegart's medium and succeeded in growing the parasites in large numbers for experimental work. A further modification of this medium by Feinberg was suggested for use in routine diagnostic work, and the present investigation has been carried out to assess its value.

Culture Medium

The formula of this medium is:

<table>
<thead>
<tr>
<th>Component</th>
<th>Amount</th>
</tr>
</thead>
<tbody>
<tr>
<td>Proteolysed liver</td>
<td>25.0 g.</td>
</tr>
<tr>
<td>Sodium chloride</td>
<td>6.5 g.</td>
</tr>
<tr>
<td>Dextrose</td>
<td>5.0 g.</td>
</tr>
<tr>
<td>Inactivated horse serum</td>
<td>80.0 ml.</td>
</tr>
<tr>
<td>Distilled water</td>
<td>1,000.0 ml.</td>
</tr>
<tr>
<td>Penicillin</td>
<td>1,000,000 units</td>
</tr>
<tr>
<td>Streptomycin</td>
<td>500,000 units</td>
</tr>
</tbody>
</table>

The solid components are dissolved in the distilled water, the serum added, and the pH adjusted to 6.4 by adding N/1 sodium hydroxide (about 9 ml per litre of medium). The mixture is sterilized by Seitz filtration and stored in screw-capped bottles in a refrigerator.

In a diagnostic medium some method of suppressing the bacteria, which would otherwise overgrow the trichomonads, is desirable. Antibiotics serve this purpose admirably, 1,000 units of penicillin and 500 units of streptomycin per ml of medium being an effective concentration. No deterioration of the anti-bacterial powers of this culture medium is evident after three months' storage in a refrigerator at +4° to +5° C.

Sources of Clinical Material

The patients from whom secretions were obtained were men and women attending the Venerous Diseases Department of the London Hospital (the Whitechapel Clinic), and women attending the Gynaecological Out-patient Department of the same hospital from June, 1954, to July, 1956.

Samples of vaginal secretions were collected in small sterile plastic spoons (Jackson, Malleson, Stallworthy, and Walker, 1948). After a fresh smear had been made and examined microscopically, the remainder of the sample was inoculated into about 7 ml of the warmed culture medium in a 6 × ½ in. test tube.

Urethral specimens from the men were obtained by Lanceley's (1954) method of gently rubbing the surface of the urethra with a platinum loop. Part

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† Panmede™ brand, in powder form, obtained from Messrs. Paines and Byrne, Ltd., Pabyrn Laboratories, Greenford, Middlesex.
of the material thus obtained was inoculated into smaller tubes (3 × 1⁄2 in.) containing 2 to 3 ml of the medium, and the rest examined microscopically.

Voided urine from both male and female patients was centrifuged, and the deposit examined microscopically and cultured in the small tubes of medium.

For direct microscopic examination of specimens, wet smears were used in preference to fixed, stained preparations because it has been found more efficient (Whittington, 1957).

The cultures were incubated immediately after inoculation at 36° C. for four or five days and examined microscopically three times during that period. It was found desirable to centrifuge the cultures before examining them, as if only a few trichomonads are present they may escape detection in a sample taken from the tube without previous spinning.

Results

The results of comparing the detection of *T. vaginalis* in fresh smears and in cultures of specimens from 207 men and 563 women are given in Table I.

### Table I

<table>
<thead>
<tr>
<th>Nature of Sample</th>
<th>Total No. of Positive Samples</th>
<th>Positive Smears and Positive Cultures</th>
<th>Positive Smears and Negative Cultures</th>
<th>Negative Smears and Positive Cultures</th>
</tr>
</thead>
<tbody>
<tr>
<td>354 urethral samples from 200 men</td>
<td>90</td>
<td>62 (69% of total positive samples)</td>
<td>17 (19% of total positive samples)</td>
<td>11 (12% of total positive samples)</td>
</tr>
<tr>
<td>378 urine samples from 207 men</td>
<td>109</td>
<td>80 (73% ..)</td>
<td>4 (4% ..)</td>
<td>25 (23% ..)</td>
</tr>
<tr>
<td>747 vaginal samples from 563 women</td>
<td>165</td>
<td>129 (78% ..)</td>
<td>6 (4% ..)</td>
<td>30 (18% ..)</td>
</tr>
<tr>
<td>225 urine samples from 70 women</td>
<td>128</td>
<td>73 (57% ..)</td>
<td>5 (4% ..)</td>
<td>50 (39% ..)</td>
</tr>
<tr>
<td>Total</td>
<td>492</td>
<td>344 (70% ..)</td>
<td>32 (6-5% ..)</td>
<td>116 (23-5% ..)</td>
</tr>
</tbody>
</table>

It was found that species of *Candida* (Monilia) also grew well in this medium. Table II gives the results of comparing direct microscopic examination of wet smears with cultures in detecting *Candida* infections in the 563 women referred to in Table I.

### Table II

<table>
<thead>
<tr>
<th>Sample</th>
<th>Total No. of Positive Samples</th>
<th>Positive Smears and Positive Cultures</th>
<th>Positive Smears and Negative Cultures</th>
<th>Negative Smears and Positive Cultures</th>
</tr>
</thead>
<tbody>
<tr>
<td>747 vaginal samples from 563 women</td>
<td>91</td>
<td>34 (37% of total positive samples)</td>
<td>0</td>
<td>57 (63% of total positive samples)</td>
</tr>
</tbody>
</table>

Discussion

As the tables show, the cultures provided a useful supplement to the examination of fresh smears in the routine diagnosis of *T. vaginalis* infections. In a few instances, negative cultures were obtained from samples in which trichomonads had been found in the fresh smears, but in many more cases the reverse obtained. This is particularly striking in the specimens of urine, in nearly one-third of which the diagnosis of trichomoniasis was based upon cultures.

*Candida*, when present in the fresh smears, always grew in the cultures, and in more than half the cases positive cultures were found although smears were negative. The presence of *Candida* did not interfere with the concomitant growth of trichomonads in the cultures.

Although no valid comparison can be made between the efficacy of the culture medium described in this paper and those previously used, because of the different techniques employed by other workers, the present medium has advantages which render it particularly suitable for diagnostic purposes. It supports a good growth of trichomonads; it is simple, cheap, easily made, and keeps for some time; and it can be used for the simultaneous detection of both *T. vaginalis* and *Candida*. A freeze-dried form of this medium may facilitate the preparation and storage of large quantities. The freeze-dried medium in individual tubes may be reconstituted with distilled water just before use. Feinberg (unpublished data) has found that this medium so dried and reconstituted will support the growth of laboratory strains of *T. vaginalis*.

Summary

A simple, cheap, and easily prepared culture medium is described for routine use in the clinical laboratory diagnosis of *Trichomonas vaginalis* and *Candida* infections.

A comparison of the examination of fresh smears and culturing in the medium described for detecting *T. vaginalis* infestations was made on 1,704 genito-urinary specimens, and for *Candida* on 747 specimens.

In the case of *T. vaginalis* a combination of the two methods gave the highest number of positive findings, but culturing was the more efficient single method. *T. vaginalis* would not have been detected in 23.5% of the samples had they not been cultured.

For the detection of *Candida*, culturing appears the better method. Sixty-three per cent. of the samples which were positive for *Candida* would
have been dismissed as negative without it. For this organism, culturing alone appears sufficient.

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References