‘Normal’ vaginal microbiology of women of childbearing age in relation to the use of oral contraceptives and vaginal tampons

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SYNOPSIS  The vaginal microbiology of women attending a family planning clinic was found to be unrelated to the use of oral contraceptives and vaginal tampons. Beta haemolytic streptococci isolated from this ‘normal’ population were compared with those from 1,104 women attending general practitioners complaining of vaginal discharge. There is a caution regarding the indications for antibiotic therapy.

Observations were made on the effects of contamination of vaginal swabs with yeasts and β-haemolytic streptococci from the vulva. The persistent character of the vaginal flora over a six-month period is described.

Reports of monilial vulvo-vaginitis associated with oral contraceptives (Yaffee and Grots, 1965; Porter and Lyle, 1966; Catterall, 1966) indicate that artificial alterations in the normal anatomy and physiology of the female genital tract may introduce a hazard to the patient. The indiscriminate use of antibiotics to treat vaginitis associated with bacteria of dubious pathogenicity has been said by Morison (1966) to predispose to ‘abacterial vaginitis’. These reports prompted this investigation, which was undertaken to establish a base line for variations in the ‘normal’ vaginal flora and to measure the influence on it of oral contraceptives and vaginal tampons.

MATERIAL AND METHODS

Swabs of vaginal secretions were examined from a series of 291 non-pregnant patients attending a family planning clinic between December 1965 and July 1966. Patients were excluded from the study only if they had a contraceptive cap in situ or were menstruating. Up to 10 women were examined at a clinic but when a busy session prohibited the examination of all eligible patients only the first few attending sequentially were included. No patient was under treatment for a discharge.

From each patient three vaginal swabs were taken through a sterile speculum or a glass funnel before a vaginal examination was performed. The first two, charcoal-impregnated swabs, were broken into Stuart’s thioglycollate transport medium and into a modified Feinberg trichomonas medium (Stenton, 1957); a third, but plain swab was used to prepare wet films and smears for staining by Gram’s method.

Five culture plates were inoculated with the swabs, from transport medium within four hours of sampling and were examined after 36, 48, and 72 hours’ incubation at 37°C. Two of these plates, 10% oxalated horse blood in Oxoid no. 2 blood agar base and MacConkey’s lactose bile salt agar, were incubated aerobically; two, horse blood agar and crystal violet blood agar, anaerobically and the heated horse blood agar (chocolate agar) plate in a candle jar. The ‘chocolate agar’ was a modification of the medium described by Cooper, Mayr-Harting, and McLachlan (1950) in which oxalated horse blood was substituted for rabbit blood. Trichomonas cultures were continued at 36°C for a week and examined on alternate days. The swab from transport medium was placed finally in a tube of 3% glucose broth containing 0.05 mg chloramphenicol per millilitre which was maintained at room temperature for a week before the broth was examined for the presence of yeasts.

These methods, apart from the yeast enrichment culture and prolonging the plate incubations for an additional 24 hours, are those used for the examination of routine vaginal swabs submitted to this laboratory.

The bacterial flora were defined by standard bacteriological methods and yeasts were identified by their ability to produce germ tubes in serum at 37°C. (Taschdjian, Burchall, and Kozinn, 1960), filament and chlamydo-spor formation on corn meal agar, and zymogram patterns.

In addition to examining the vaginal flora, 162 patients were investigated for the presence of yeasts and...
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β-haemolytic streptococci on the vulva; swabs from the vulva were plated onto crystal violet blood agar and incubated in glucose chloramphenicol broth.

RESULTS

THE POPULATION Of the 291 women examined, nearly three quarters (72%) belonged to social classes II or III (Census, 1961). The ages ranged from 18 to 45, with 75% aged 20 to 35. Thirty women were single and 261 married. One in four was nulliparous whereas 56% had had two to five pregnancies. Oral contraceptives were used by 104 patients, two-thirds for a period exceeding six months. Vaginal tampons were used by 184 women (Table I).

Cervical erosions were observed in 51% of patients taking oral contraceptives but in only 33% of women who were not on hormone treatment ($\chi^2 = 8.87$, $P < 0.01$).

Specimens were taken at any stage of the menstrual cycle with the result that nearly 30% were taken in each of the last three weeks of the cycle.

DIRECT SMEARS Two patients had purulent vaginal secretions, both associated with Trichomonas vaginalis infection; all other specimens were of a predominant epithelial type with few or no leucocytes.

CULTURE RESULTS The vaginal flora in relation to contraceptive and sanitary method is summarized in Table II. Isolations of two different species of pseudomonads and single strains of a klebsiella and a bacillus are not included as they do not warrant subdivision. A heterogeneous group of 11 catalase-negative microaerophilic Gram-variable bacilli is also not tabulated.

### Table I

**VAGINAL FLORA SURVEY IN A FAMILY PLANNING CLINIC POPULATION**

<table>
<thead>
<tr>
<th>Patients</th>
<th>Percentage Population</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total population</td>
<td>291</td>
</tr>
<tr>
<td>Marital status</td>
<td></td>
</tr>
<tr>
<td>Single</td>
<td>30</td>
</tr>
<tr>
<td>Married</td>
<td>261</td>
</tr>
<tr>
<td>Contraceptive method</td>
<td></td>
</tr>
<tr>
<td>(a) Hormonal</td>
<td></td>
</tr>
<tr>
<td>Lyndiol</td>
<td>40</td>
</tr>
<tr>
<td>Anovlar</td>
<td>25</td>
</tr>
<tr>
<td>Ovulen</td>
<td>25</td>
</tr>
<tr>
<td>Other</td>
<td>14</td>
</tr>
<tr>
<td>(b) No method</td>
<td></td>
</tr>
<tr>
<td>Mechanical</td>
<td>83</td>
</tr>
<tr>
<td>Chemical</td>
<td>3</td>
</tr>
<tr>
<td>Mechanical and chemical</td>
<td>52</td>
</tr>
<tr>
<td>Sanitary method</td>
<td>Tampons</td>
</tr>
<tr>
<td>Towels</td>
<td>107</td>
</tr>
</tbody>
</table>

1 Lyndiol, 5 mg. lynoestrol + 0.15 mg. mestranol
2 Anovlar, 4 mg. norethisterone acetate + 0.05 mg. ethinyloestradiol

### Table II

**VAGINAL FLORA OF FAMILY PLANNING PATIENTS IN RELATION TO CONTRACEPTIVE AND SANITARY METHODS**

<table>
<thead>
<tr>
<th>Isolation (%) of Patients in Each Category</th>
<th>Total Population</th>
<th>Oral Contraceptives</th>
<th>Vaginal Tampons</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>Used</td>
<td>Not Used</td>
<td>Used</td>
</tr>
<tr>
<td>A Culture Classification</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Abundant mixed flora</td>
<td>24.4</td>
<td>23.1</td>
<td>25.0</td>
</tr>
<tr>
<td>Bacteria of faecal origin</td>
<td>16.8</td>
<td>12.5</td>
<td>19.3</td>
</tr>
<tr>
<td>B Bacteria</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Clostridium welchii</td>
<td>1.4</td>
<td>1.0</td>
<td>1.6</td>
</tr>
<tr>
<td>Corynebacteria</td>
<td>21.3</td>
<td>19.2</td>
<td>22.4</td>
</tr>
<tr>
<td>Lactobacilli (predominant)</td>
<td>49.1</td>
<td>52.9</td>
<td>47.0</td>
</tr>
<tr>
<td>Staphylococci and micrococci</td>
<td>37.3</td>
<td>37.5</td>
<td>37.4</td>
</tr>
<tr>
<td>Staphylococcus pyogenes</td>
<td>2.7</td>
<td>4.8</td>
<td>1.6</td>
</tr>
<tr>
<td>Streptococci</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Beta haemolytic</td>
<td>10.9</td>
<td>9.7</td>
<td>11.8</td>
</tr>
<tr>
<td>Strep. faecalis</td>
<td>9.6</td>
<td>7.7</td>
<td>10.7</td>
</tr>
<tr>
<td>Other aerobic</td>
<td>18.2</td>
<td>22.1</td>
<td>14.0</td>
</tr>
<tr>
<td>Anaerobic</td>
<td>4.8</td>
<td>3.8</td>
<td>5.3</td>
</tr>
<tr>
<td>Bacteroides</td>
<td>1.0</td>
<td>0.0</td>
<td>1.6</td>
</tr>
<tr>
<td>Escherichia coli</td>
<td>5.8</td>
<td>3.8</td>
<td>6.9</td>
</tr>
<tr>
<td>Proteus</td>
<td>2.7</td>
<td>1.0</td>
<td>3.7</td>
</tr>
<tr>
<td>C Yeasts</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Candida albicans</td>
<td>10.7</td>
<td>13.5</td>
<td>9.1</td>
</tr>
<tr>
<td>Other yeasts</td>
<td>6.9</td>
<td>3.9</td>
<td>8.5</td>
</tr>
</tbody>
</table>

1 Two or more species, with no one species forming more than 60% colonies isolated
2 Sum of Strep. faecalis, Esch. coli and Cl. welchii
3 80% or more of colonies isolated
TABLE III

ISOLATIONS OF BETA HAEMOLYTIC STREPTOCOCCI IN THE VAGINA FROM FAMILY PLANNING CLINIC AND GENERAL PRACTICE PATIENTS

<table>
<thead>
<tr>
<th>Lancefield's Group</th>
<th>Total No. of Strains</th>
<th>No. of Strains in Profuse Growth</th>
<th>No. of Strains</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total</td>
<td>32 (10.9%)</td>
<td>12 (4.0%)</td>
<td>66 (6.0%)</td>
</tr>
<tr>
<td>A</td>
<td>1 (0.3%)</td>
<td>1 (0.3%)</td>
<td>15 (1.4%)</td>
</tr>
<tr>
<td>B</td>
<td>18 (6.2%)</td>
<td>9 (3.1%)</td>
<td>20 (1.8%)</td>
</tr>
<tr>
<td>C</td>
<td>2 (0.7%)</td>
<td>1 (0.3%)</td>
<td>6 (0.5%)</td>
</tr>
<tr>
<td>D</td>
<td>10 (3.4%)</td>
<td>1 (0.3%)</td>
<td>21 (1.9%)</td>
</tr>
<tr>
<td>F</td>
<td>—</td>
<td>1 (0.1%)</td>
<td>1 (0.1%)</td>
</tr>
<tr>
<td>G</td>
<td>1 (0.3%)</td>
<td>1 (0.3%)</td>
<td>3 (0.3%)</td>
</tr>
</tbody>
</table>

The serological groups of β-haemolytic streptococci are shown in Table III.

Eighteen per cent of patients harboured vaginal yeasts of which just over half (10.7%) were Candida albicans (Table IIIC); other yeasts isolated included three strains of Torulopsis glabrata, two of Saccharomyces cerevisiae, and 15 Candida species including C. krusei, C. parakrusei, C. pseudotropicalis, and C. guillermondii. The mode of primary isolation of yeasts is given in Table IV.

TABLE IV

FACILITY OF YEAST DETECTION AS A QUANTITATIVE ESTIMATE OF YEASTS FROM THE VAGINA OF FAMILY PLANNING CLINIC PATIENTS

<table>
<thead>
<tr>
<th>Contraceptive No. of Patients Yeast</th>
<th>Method of Identification</th>
<th>Culture</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Direct Microscopy 100%</td>
<td>Direct Enrichment</td>
</tr>
<tr>
<td>Oral 104</td>
<td>C. albicans 7 4 11 3</td>
<td>1 3</td>
</tr>
<tr>
<td>Other 14</td>
<td>3 2 3 1</td>
<td></td>
</tr>
<tr>
<td>Non-oral 187</td>
<td>C. albicans 7 4 9 8</td>
<td>2 6</td>
</tr>
<tr>
<td>Other 17</td>
<td>7 2 10 8</td>
<td></td>
</tr>
</tbody>
</table>

Wet preparations and stained films

Clostridium welchii was isolated from four specimens and always as occasional colonies in a mixed culture.

Staphylococcus pyogenes was recovered in profuse growth from one specimen only.

Escherichia coli was isolated more often from samples taken during the first half of the menstrual cycle (8.6%) than the second half (4.3%) whereas Streptococcus faecalis and corynebacteria were isolated with uniform frequency at all stages of the cycle.

Trichomonas vaginalis was found in three patients, two in association with symptoms. The third patient had no symptoms and the protozoa were demonstrated by culture only.

Mycoplasma sp. are not recorded in Table III. Colony forms morphologically consistent with pleuropneumonia-like organisms were present on impression smears of some cultures but it was not found practicable to record their numbers in mixed culture with any certainty. To assess accurately the prevalence of mycoplasmas will require a special investigation.

SWABS FROM THE VULVA Examination of swabs from the vagina and vulva of 162 patients showed that 16.6% of these women had yeasts (10.5% C. albicans) at both sites. An additional 5% of patients harboured yeasts (1.2% C. albicans) on the vulva but not in the vagina. Beta haemolytic streptococci were present in both sites in 9.8% of patients, with an additional 3% demonstrated from the vulva.

RE-EXAMINATIONS Nineteen of 22 patients re-examined six months after their first visit showed no change of flora; this group included two patients treated with C. albicans, one with S. cerevisiae, and one with group B streptococci present on both occasions. Fifteen patients, including seven who had started taking oral contraceptives after their first visit, retained a flora in which lactobacilli predominated. Three patients showed a change, losing a group A and group C streptococcus and a pseudomonad respectively.

DISCUSSION

The striking feature of the results is the similarity of the vaginal flora irrespective of sanitary and contraceptive methods. Although patients taking oral contraceptives had a significantly higher incidence of cervical erosions than that among patients not on hormones, this was not associated with alterations in the vaginal flora or with an increased incidence of a discharge. Since these two groups of patients were not fully comparable for age and parity and the type of erosion was not recorded, it should not be inferred from this study that oral contraceptives necessarily predisposed these patients to cervical erosions.

The insertion of tampons is likely to carry bacteria into the vagina from the vulva and perianal skin but Table II A shows that patients who used tampons were not specially liable to have a mixed bacterial flora or to carry faecal organisms in the vagina. Since most specimens were taken more than a week...
after the first day of the last menstrual period, it
may be that usually the vaginal lactobacillary flora
was re-established and contaminants displaced
before samples were collected.

The figures for β-haemolytic streptococci warrant
further analysis and comment because of the high
frequency with which they occurred in profuse
culture (Table III). Nearly 11% of patients harboured
β-haemolytic streptococci in the vagina. The
predominance of group B and group D strains is
consistent with their acid-resistant properties
relative to the other groups. If these patients had
been treated because of the presence of numerous
streptococci without prior determination of the
serological group, 3% of patients would have
received antibiotics for group B infections which
were presumably of no significance. That this is
relevant to the routine examination of vaginal
swabs is shown by the similar incidence of 1·8%
of group B streptococci (profuse growth) isolated in
our laboratory from 1,104 vaginal swabs submitted
from patients complaining of a discharge. In view of
reports of fatal acute and subacute endocarditis
caused by haemolytic streptococci other than group
A (Rosenthal and Stone, 1940; Ramsay and
Gillespie, 1941), treatment may be indicated for
antenatal and puerperal patients and women with
cardiac abnormalities in whom any haemolytic
streptococcus may be a potential pathogen.

Liston and Cruickshank (1940) recorded an
increased incidence of vaginal moniliasis occurring in
pregnancy; this predisposition has been ascribed
(Davis and Pearl, 1938) to glycogen deposition in
the vaginal epithelium, a phenomenon said by Cruick-
shank and Sharman (1934) to be under the influence
of oestrogen. Porter and Lyle (1966) described
difficulties in curing patients of C. albicans vulvo-
vaginitis when the patients were using oral contraceptives.
Three of their 13 patients taking Enovid
(norethynodrel with mestranol) failed to respond to
treatment until taken off the 'pill'. Their observations
are supported by those of Yaffee and Grots (1965)
and Catterall (1966).

The incidence of 13.5% C. albicans (Table IIC)
in clinic patients using oral contraceptives is not
significantly higher than that of 9-1% among
patients not on hormones. Three of the 30 single
women had latent C. albicans infection. Neither
marital state nor parity appeared to influence the
frequency of Candida isolations.

Oestrogen supplementation occurring in a patient
taking oral contraceptives may promote yeast
multiplication in the vagina; this, however, may be
prevented from reaching a pathological and sympto-
matic level by the monthly withdrawal of the 'pill'
to allow 'menstruation'. If this is so, then a measure
of the enhancement of yeast multiplication is the
ease with which yeasts are found by direct microscopy
and culture as compared with enrichment culture
alone. Table IV records the ease with which yeasts
were found; the difference between those using and
not using the 'pill' is not significant.

In this study, patients using oral contraceptives
showed no significantly increased incidence of
C. albicans and did not have a more severe infection
once yeasts had reached the vagina. All these patients
were treated with combinations of oestrogens and
progestogens; the particular preparation prescribed
was chosen, after assessment of the individual, as
the one least likely to impose side effects. Lyndiol is
expected to exert a predominant oestrogenic-like
effect and Anovlar and Ovulen are predominantly
progestrone-like in action.

The persistence over a six-month period of S.
cerevisiae in the vagina of a patient with a chronic
low-grade discharge deserves note. This strain grew
well at 37°C. It may have originated from the
intestine but in this patient was never associated
with bacteria of faecal origin.

Asymptomatic trichomoniasis occurred in one
patient. Direct films of the vaginal secretions from
this patient showed numerous epithelial cells with
only occasional leucocytes and gave no indication of
the underlying infection. Retrospective re-examina-
tion of smears failed to demonstrate the protozoan.

A general practitioner may have difficulty in
obtaining samples of vaginal secretions uncontami-
nated by vulval flora. Comparison of the numbers
of yeasts and β-haemolytic streptococci isolated from
paired vulval and vaginal swabs suggests that
accidental contamination from the vulva may
increase the isolations of commensal yeasts and
β-haemolytic streptococci but only rarely of C.
albicans. Group A streptococci were present on
the vulva of one patient but not demonstrated in the
vagina.

Difficulties in gauging the significance of the
vaginal flora are increased by the patient's assessment
of her discharge. None of the clinic patients had
sought advice for a discharge. On direct questioning
55% considered they had a discharge in excess of
what the patient considered 'normal'. This was
confirmed by examination in only 11 patients, 3·8% of
the total population studied, and from these a
microbiological cause for the discharge was
demonstrated in less than half.

We wish to thank Dr. June Davy and Dr. Mary Farr for
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REFERENCES


