

a low production of pigment on normal media, according to all the other biochemical tests showed them to be normal *P. aeruginosa* strains; this was confirmed by the results of the growth on the milk agar, both hydrolysis of the casein and pigment production being observed. In the case of the other two strains, 950 and 5940, the position was not so straightforward. Neither produced pigment on any of the solid media, including the milk agar. They did, however, show some signs of hydrolysis of the casein, and this increased on incubation for a further 48 hours, but no pigment was demonstrable. The results of the biochemical tests likewise showed that these two strains did not conform precisely to the *P. aeruginosa* or the *P. fluorescens* patterns, but in fact have some characters common to both, and these can be regarded as intermediate *fluorescens/aeruginosa* types. This is supported by the results of the milk agar media previously discussed. Indeed, in view of the recently demonstrated high transformation frequency within the genus *Pseudomonas* (Khan and Sen, 1967), the existence of such intermediate strains is not surprising. In either case, the milk agar has shown itself to be as reliable as the more conventional biochemical tests.

We wish to thank Dr S. P. Lapage, of the National Collection of Type Cultures, for kindly supplying the poorly pigmented strains of *P. aeruginosa*, and Dr M. T. Parker, of Central Public Health Laboratory, Colindale, London, NW9, for kindly carrying out the phage typing. We also wish to thank The Medical Research Council for a grant which supported part of this work.

#### References

- Colwell, R. R. (1964). *J. gen. Microbiol.*, **37**, 181-194. A study of features used in the diagnosis of *Pseudomonas aeruginosa*.
- Cowan, S. T., and Steel, K. J. (1965). *Manual for the Identification of Medical Bacteria*. Cambridge University Press, London.
- Gaby, W. L., and Free, E. (1953). Occurrence and identification of nonpigmented strains of *Pseudomonas aeruginosa* in the clinical laboratory. *J. Bact.*, **65**, 746.
- Gaby, W. L., and Free, E. (1958). Differential diagnosis of *Pseudomonas*-like microorganisms in the clinical laboratory. *J. Bact.*, **76**, 442-444.
- Haynes, W. C. (1951). *Pseudomonas aeruginosa*—its characterization and identification. *J. gen. Microbiol.*, **5**, 939-950.
- Hugh, R., and Leifson, E. (1953). The taxonomic significance of fermentative versus oxidative metabolism of carbohydrates by various gram negative bacteria. *J. Bact.*, **66**, 24-26.
- Khan, N. C., and Sen, S. P. (1967). Genetic transformation in *Pseudomonas*. *J. gen. Microbiol.*, **49**, 201-209.
- King, E. O., Ward, M. K., and Raney, D. E. (1954). Two simple media for the demonstration of pyocyanin and fluorescein. *J. Lab. clin. Med.*, **44**, 301-307.
- Kovacs, N. (1956). Identification of *Pseudomonas pyocyanea* by the oxidase reaction. *Nature (Lond.)*, **178**, 703.
- Lysenko, O. (1961). *Pseudomonas*—An attempt at a general classification. *J. gen. Microbiol.*, **25**, 379-408.
- Phillips, I. (1969). Identification of *Pseudomonas aeruginosa* in the clinical laboratory. *J. Med. Microbiol.*, **2**, 9-16.
- Preston, N. W., and Morrell, A. (1962). Reproducible results with the gram stain. *J. Path. Bact.*, **84**, 241-243.
- Rhodes, M. E. (1959). The characterization of *Pseudomonas fluorescens*. *J. gen. Microbiol.*, **21**, 221-263.
- Rhodes M. E., (1961). The characterization of *Pseudomonas fluorescens* with the aid of an electronic computer. *J. gen. Microbiol.*, **25**, 331-345.
- Rogers, K. B. (1963). Oxidase reaction. (Letter), *Lancet*, **2**, 682.
- Stanier, R. Y., Palleroni, N. J., and Doudoroff, M. (1966). The aerobic pseudomonads: a taxonomic study. *J. gen. Microbiol.*, **43**, 159-271.
- Wahba, A. H., and Darell, J. H. (1965). The identification of atypical strains of *Pseudomonas aeruginosa*. *J. gen. Microbiol.*, **38**, 329-342.

#### Errata

In Table II in the paper entitled, 'Comparison of quick and slow thaw methods of producing cryoprecipitate antihaemophilic factor from fresh and 24-hour-old blood' A. L. Bloom (*J. clin. Path.*, **22**, 447-452) the P values for the supernatant have been printed under the wrong headings. The correct table 'Factor VIII content of cryoprecipitate and supernatant plasma', is printed below.

	Fresh Blood		Twenty-four Hour Blood	
	Quick-Thaw (A)	Slow-Thaw (B)	Quick-Thaw (C)	Slow-Thaw (D)
Number of samples	101	67	102	76
Factor VIII in cryoprecipitate units) Mean ± SD	83 ± 32	112 ± 44	53 ± 25	72 ± 30
Factor VIII in supernatant (units) Mean ± SD	42 ± 21	30 ± 21	31 ± 19	25 ± 12
Statistical significance	A v B	C v D	A v C	B v D
Supernatant	P = <0.001 P = <0.001			
Cryoprecipitate	P = <0.001 P = <0.001 P = <0.001 P = <0.001			

In Table IIa of the paper by Davis *et al.*, *J. Clin. Path.*, 1969, **22**, 634, the figures for *Proteus mirabilis* under the columns for tetracycline should read: S: 9.7%, 'S': 0.9% and R: 89.4%; the mean percentages of total should therefore read: S: 30.8%, 'S': 6.1%; R: 63.1%.