Stimulation of growth on a deficient medium of a *Klebsiella aerogenes* species by sulphonamide, para-amino benzoic acid, nitrofurantoin, and some other substances

D. McGHIE, J. G. P. HUTCHISON, AND R. G. FINCH

From the Regional Public Health Laboratory, East Birmingham Hospital, Birmingham

SYNOPSIS A strain of *Klebsiella aerogenes* is described which was unable to grow from light inocula on 5% lysed horse blood diagnostic sensitivity test (DST) agar. It was able to grow on DST agar without added horse cells and MacConkey and nutrient agar. Growth on a deficient medium was promoted by sulphonamides and nitrofurantoin as well as by para-amino benzoic acid, para-amino salicylic acid, procaine hydrochloride, thymine, and uracil.

The organism, *Klebsiella aerogenes*, was first isolated from a urine specimen on MacConkey agar (Oxoid). When antimicrobial sensitivity testing was attempted using DST agar (Oxoid) enriched with 5% lysed horse blood and a Mastring (Mast Laboratories), bacterial growth took place as shown in Figure 1. The centre of the plate was free from growth and what appeared to be stimulated growth was seen associated with the sulphathiazole (no. 8) and nitrofurantoin (no. 5) positions. There follows an account of some features of this organism.

**Materials and Methods**

**MATERIALS**

Diagnostic sensitivity test (Oxoid) agar with 5% saponin lysed horse blood was used throughout to test the effect of growth-promoting substances. Other media such as MacConkey agar (Oxoid) or 1% peptone water (Oxoid) were not critical. The organism, *Klebsiella aerogenes*, was isolated from the urine of a male inpatient aged 55 who was a chronic bronchitic with asthma, from whom urine was taken as a routine (no clinical evidence of infection). We have no evidence of co-trimoxazole treatment at any time, but in retrospect the possibility of cross infection from the patients on the same ward, treated with co-trimoxazole cannot be discounted.

Fig. 1  Antibiotic sensitivity test of Kl. aerogenes, DST lysed horse blood agar. Growth of the organism is limited to a small area centred about (8) sulphathiazole (200 µg), and (5) nitrofurantoin (200 µg) on the Mastring. The other antimicrobial agents are (1) ampicillin 25 µg (2); colistin methane sulphonate 200 µg; (3) kanamycin 30 µg; (4) nalidixic acid 30 µg; (6) streptomycin 25 µg; and (7) tetracycline 100 µg.

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METHODS
From an overnight broth or peptone water culture of the organism a 4 ml tube of peptone water was inoculated with a straight wire. This was incubated at 37°C for about four hours until turbidity was just visible: a 1/1000 dilution of this culture was then made in sterile phosphate-buffered saline, pH 7·3 (Dulbecco A Oxoid). The number of organisms in this suspension was of the order of 10⁸/ml; variations between 10³ and 10⁴ organisms/ml were permissible. The suspension was flooded onto the test plate, the excess fluid sucked off, and the plates were allowed to dry. The test plates had been previously matured by incubation at 37°C for four hours (Harper and Cawston, 1945).Appearances in the accompanying figures were not always easy to reproduce in so clear-cut a way as shown. A light inoculum of a young culture gave the best results: older cultures produce aggregations of growth akin to pellicle formation which made them unsuitable for use, and if the inoculum was too heavy 'break through' growth took place, which, however, was usually light and easily distinguished from the stimulation effects to be described. Furthermore the inhibition of bacterial growth on different batches of horse-blood DST agar varied; some batches were less inhibitory than others, but we can suggest no reason for this.

Inhibitions between substances were tested using impregnated blotting paper strips applied to the medium at an angle of 90° to each other. For titration of the extinction point, solutions were applied in porcelain insulator beads (fish-spine beads) and the zone diameters plotted in the usual way.

Experimental Results
Stimulation of growth was found with the sulphonamides (sulphamethoxazole and sulphanilamide), and substances chemically similar (Fig. 2) such as para-aminobenzoic acid (PABA), para-aminosalicylic acid (PAS), and procaine HCl. Sulphanilamide and PABA were titrated decimally to extinction; sulphanilamide stimulation could be detected at a concentration of 10⁻⁴ molar, whereas PABA (or its more soluble sodium salt) stimulated only down to 10⁻² molar concentration. In other words, as a growth stimulator, sulphanilamide was 10³ times more active than PABA mole for mole. Para-aminobenzoic acid and procaine HCl interfered with the actions of sulphonamide and PAS (Fig. 2). Figure 2 also shows that PABA, at a concentration insufficient to stimulate growth, inhibited sulphonamide stimulation.

Two further effects were seen, which are illustrated in Figures 3 and 4. In Fig. 3 a sulpha-methoxazole

Fig. 2 Inhibition of sulphonamide activity by para-aminobenzoic acid on DST lysed horse-blood agar seeded with Klebsiella. Blotting paper strips soaked in solutions of the two substances (sulphanilamide, M/1000; PABA, M/50) are surrounded by stimulated growth. Para-aminobenzoic acid has, however, inhibited sulphonamide stimulation where present in a concentration too low to promote growth.

Fig. 3 Sensitivity plate seeded with Klebsiella. The central disc contains sulphamethoxazole 200 µg. Growth of the organism is first inhibited then stimulated by lower concentrations of the sulphonamide.
A disturbing feature of this organism is that we are unable to judge from *in-vitro* tests whether it is or is not sensitive to co-trimoxazole or the sulphamides. The latter appear to inhibit in high concentration and stimulate in low. By adjusting the amounts of thymine (or other substances) the organism may appear sensitive or resistant at will and the *in-vitro* tests cannot therefore be considered to be a useful guide to therapy.

Turning to the stimulating effect of sulphonamide, this seems to be a rather uncommon finding. We can discover references only to stimulation of the *Rickettsia* of murine typhus (Snyder, Maier, and Anderson, 1942) and conversely some benefit to clinical typhus ensuing on high dosage of PABA (Yeomans, Snyder, Murray, Zarafonetis, and Ecke, 1944). The present organism, however, stimulated *in vitro* by both substances whilst the competitive inhibition between them remains unaltered. As the patient appeared not to have been treated with co-trimoxazole we do not know if his condition would or would not have been aggravated by giving these drugs, but the possibility of adverse effects should be borne in mind if similar organisms become common. Dr R. M. Maskell (personal communication) has also isolated co-trimoxazole-resistant, sulphonamide-stimulated *Proteus mirabilis* strains in Portsmouth so that the organism we are reporting is not unique.

Finally we think that this and similar organisms may prove of use as a metabolic tool and of interest for work *in vivo*. It has certainly made us look very carefully at our routine procedures so that we do not miss, as we may have in the past, significant growth of coliforms and perhaps other organisms, which may exhibit peculiarities similar to those reported here.

### Discussion

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### References


Yeomans, A., Snyder, J. C., Murray, E. S., Zarafonetis, C. J. D., and Ecke, R. S. (1944). The therapeutic effect of *p* 26th. *Amer. med. Ass.*, 126, 349-

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*Fig. 4* Sensitivity plate identical to *Fig. 1* but a disc impregnated with co-trimoxazole has been placed centrally. The effect of the chance application of this disc has been to stimulate growth around it and so apparently to make the test readable.