Urinary inorganic phosphorus determinations

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SYNOPSIS The Parekh-Jung method for determination of inorganic phosphorus in serum was applied to its determination in urine. Accuracy is good. The mean percentage recovery of added phosphorus was 100.2%. Forty analyses of a sample gave a mean value of 46.71 mg/100 ml ± 0.76 standard deviation (between-batch precision). The relative usefulness of the Parekh-Jung method compared with those other methods is discussed.

In an earlier publication Parekh and Jung (1970a, 1970b) introduced the use of p-phenylenediamine dihydrochloride as a reducing agent for the determination of inorganic phosphorus in serum. This new method possessed advantages over the existing methods in terms of simplicity, speed, reproducibility, reagent stability, etc. Recently, the authors applied this method to the determination of inorganic phosphorus in urine.

We now describe our experience with various interfering substances which may be present in urine in considerable concentrations. The results are compared with those obtained by employing the methods of Dryer, Tammes, and Routh (1957), Fiske and Subbarow (1925), and Gomori (1941-42).

Materials and Methods

REAGENTS
The reagents used are the same as those used for the determination of inorganic phosphorus in serum.

Molybdic-trichloracetic acid reagent
One volume of molybdic acid solution is mixed with 2 volumes of 10% trichloracetic acid solution.

Reducing agent
p-Phenylenediamine dihydrochloride (Eastman Kodak No. 207) 0.5 g is dissolved in 100 ml of a 5% sodium meta-bisulphite solution in water.

Phosphorus standard
For the stock solution, 0.3514 g of anhydrous monopotassium phosphate is dissolved in 10 ml of 10 N sulphuric acid and diluted to 1 litre with water to give 8 mg inorganic phosphorus/100 ml. For the standard solution, an aliquot of the stock solution is diluted with an equal volume of water to give 4 mg inorganic phosphorus/100 ml.

Hyland urine control
A commercial reference preparation is available from Hyland, Division of Travenol Laboratories, Inc, Costa Mesa, California, USA.

PROCEDURE
(1) Dilute an aliquot of a 24-hour urine specimen 1:10 (v/v) with water. (2) Deliver 0.1 ml of diluted urine to a 3 ml centrifuge tube, and add 0.9 ml of molybdic-trichloracetic acid reagent. (3) Substitute standard solution and water for urine in the standard and blank tubes respectively in step 2 above. (4) Mix well, allow the tubes to stand for five minutes and centrifuge for five minutes. (5) Transfer 0.5 ml of the supernatant to a 12 × 75 mm cuvette and add 2 ml of the reducing reagent. Mix by inversion and allow to stand for 20 minutes for colour development. (6) Read against blank in a spectrophotometer at 680-700 mM.

No precipitation occurs with urine in step 4, except in cases of proteinuria.
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OD Unknown \times \text{concentration of standard} \times 10
OD Standard
\times \frac{\text{ml of 24-hour specimen}}{1\ 000} = \text{g P/24-hr urine.}

Results

Known quantities of inorganic phosphorus in the form of monobasic potassium phosphate were added to aliquots of a urine pool. Analyses were performed in triplicate for each set of the recovery study outlined in Table I. The percentage recoveries ranged from 98·8 to 102·0 with a mean percentage recovery of 100·2.

<table>
<thead>
<tr>
<th>Substance Added</th>
<th>Concentration of Added Substrate (mg/100 ml)</th>
<th>Method</th>
<th>Dryer (Percentage difference from true value)</th>
<th>Fiske-Subbarrow</th>
<th>Mean</th>
<th>SD</th>
<th>Coefficient of Variation</th>
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<tbody>
<tr>
<td>L-Ascorbic acid</td>
<td>20</td>
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<td>+0·85</td>
<td>+1·27</td>
<td>+1·06</td>
<td>+0·85</td>
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<tr>
<td>Creatinine</td>
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<tr>
<td>Inositol</td>
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<td>D-Glucose</td>
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<td></td>
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<tr>
<td>Oxalate</td>
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<td>-0·6</td>
<td>-0·4</td>
<td>-0·4</td>
</tr>
</tbody>
</table>

Table III Effects of interfering substances with different methods when added to urine containing 362 mg phosphorus per 100 ml

Discussion

In the literature, a separate study of urinary inorganic phosphorus determinations has not usually been described. However, interference by a number of substances in serum or plasma has been reported by Denis and von Meysenburg (1922), Negrin (1964), and Baginski, Foa, and Zak (1967). Such problems encountered in the measurement of inorganic phosphorus in normal urine could be considerably magnified owing to the very variable composition of urine.

Very little or no interference was observed on applying the proposed method to urinary inorganic phosphorus analyses in the presence of 20 mg/100 ml of creatinine, L-histidine, inositol, D-glucose, salicylate, or 50 mg/100 ml of oxalate or 100 mg/100 ml of citrate. Maximum interference was caused by the addition of 20 mg/100 ml of L-ascorbic acid and this amounted to less than 1% of difference from the true value. It is noteworthy that seven of the eight compounds investigated are normally present in urine. The results in Table III are therefore indicative of the accuracy of the method.

The results shown in Tables II and III indicate that the Fiske-Subbarow and Gomori methods are closely comparable. Similarly, it can be concluded that the Dryer method compares favourably with the Parekh-Jung method. However, the Dryer method has certain serious shortcomings. Not only does it lack the simplicity of the Parekh-Jung method, but the insolubility as well as the instability
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of its reducing agent (Henry, 1964) is such that, for all practical purposes, it would have a limited application. The reagents of the Parekh-Jung method are stable for up to at least six months on the shelf. Also the reducing agent, p-phenylene-diamine dihydrochloride, is a stable chemical and is freely soluble in water. The final reaction yields a blue colour, free of turbidity, which is very stable and obeys Beer's law.

The Parekh-Jung method is therefore at least as accurate and precise as the other methods. The procedure is simple and the reagents are stable. After the preliminary dilution, the urine sample is treated exactly as the serum sample thus facilitating simultaneous analyses. Urinary proteins, when present, are removed and hence cause no interference in this method. The Parekh-Jung method is thus ideal for routine use.

References


