A simple assay of intrinsic factor-vitamin B$_{12}$ complex employing the binding intrinsic factor antibody

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SYNOPSIS The reaction between binding intrinsic factor antibody and intrinsic factor-vitamin B$_{12}$ complex has been studied. Initially in the zone of antibody excess, the relationship between the amount of antigen present and the amount of antigen-antibody complex adsorbed onto zirconium phosphate gel was linear. With increasing amounts of antigen, the curve departed from linearity and reached a plateau. The linear portion of the reaction forms the basis of a simple and reproducible assay for quantitating intrinsic factor to which vitamin B$_{12}$ has already been bound. The assay provides a method for studying the fate of intrinsic factor-vitamin B$_{12}$ complex during digestion and absorption. In two normal subjects given radioactive vitamin B$_{12}$ orally, aspiration of ileal contents showed that only 50 to 70% of the radioactivity was bound to intrinsic factor at that level.

The study of the physiology of vitamin B$_{12}$ absorption has been hampered by the lack of a simple assay for human intrinsic factor-vitamin B$_{12}$ complex. The activity of human intrinsic factor-radioactive vitamin B$_{12}$ complex can be measured in vivo in patients with pernicious anaemia (Heine, Welch, Scharf, Meacham, and Prusoff, 1952; Schilling, 1953; Glass, Boyd, Gellin, and Stephanson, 1954; Baker and Mollin, 1955). However, this requires the cooperation of patients with pernicious anaemia, is time consuming, and comparatively insensitive. Human intrinsic factor-vitamin B$_{12}$ complex can also be estimated in vitro by techniques employing ileal segments, everted ileal sacs, or small intestinal homogenates (Wilson and Wiseman, 1954; Wolff and Nabet, 1961; Sullivan, Herbert, and Castle, 1962; Boass and Wilson, 1963; Castro-Curel and Glass, 1963). These techniques have the disadvantage that they are specialized and difficult to apply to large numbers of samples.

The recognition of two types of intrinsic factor antibodies, the blocking antibody which prevents the combination of vitamin B$_{12}$ with intrinsic factor, and the binding antibody which unites either with intrinsic factor or intrinsic factor-vitamin B$_{12}$ complex (Garrido-Pinson, Turner, Crookston, Samloff, Miller, and Segal, 1966), has made possible the development of specific in vitro intrinsic factor assays.

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Materials and Methods

The technique is based on the fact that zirconium gel adsorbs protein complexes at or below their isoelectric point. The vitamin $B_{12}$ binders in gastric juice, including intrinsic factor, are quantitatively adsorbed onto the gel at pH 5.0, but at pH above 5.5 the negative charge of the intrinsic factor and other gastric juice binders increases and they are not adsorbed onto the gel (Hansen et al., 1966). When the binding type antibody is attached to the intrinsic factor-vitamin $B_{12}$ complex, the isoelectric point changes and it is adsorbed onto the gel at pH 6.25 (Jacob, Hansen, and Miller, 1966). Substances in solution not adsorbed onto the gel, including free radioactive vitamin $B_{12}$, can be removed by washing the gel with buffer.

Zirconium phosphate gels at pH 5.0 and pH 6.25 were prepared by the method of Hansen et al. (1966). $^{57}$Co-labelled radioactive vitamin $B_{12}$ (specific activity 70 μC/μg) was obtained from the Radiochemical Centre, Amersham. This was diluted before use with non-radioactive vitamin $B_{12}$ to give a specific activity of approximately 8 μC/μg. Radioactive measurements were carried out in a Packard automatic gamma scintillation counter, at least 10,000 counts being accumulated in each case.

Sera containing the binding and blocking antibody were obtained from four patients with pernicious anaemia. The titre of the antibodies in these sera was measured by zirconium gel assay (Hansen et al., 1966; Jacob et al., 1966). Control sera were obtained from healthy adults with no demonstrable intrinsic factor antibodies. To prevent radioactive vitamin $B_{12}$ from binding to transcobalamin II in the serum and being adsorbed to the gel (Hansen, Tan, and Jacob, 1967), the vitamin $B_{12}$ binding sites of the antibody sera and the control sera were saturated by adding excess non-radioactive vitamin $B_{12}$ (5 ng/ml). When the sera were used as a source of blocking antibody, the unbound vitamin $B_{12}$ was removed by albumin-coated charcoal prepared as described by Gottlieb et al. (1965). The removal of excess unlabelled vitamin $B_{12}$ was unnecessary when the serum was used as a source of binding antibody because the serum was added to the system after radioactive vitamin $B_{12}$ was combined with intrinsic factor.

Gastric juices were obtained from healthy adult volunteers and patients with pernicious anaemia. The fasting juice was discarded. An intramuscular injection of pentagastrin (6 μg/kg body weight) was given and the juice aspirated at 15, 30, and 45 minutes thereafter. The juice was immediately filtered through glass wool, the pH adjusted to 10.4 with 1N NaOH to inactivate the pepsin and 30 minutes later brought back to 7.4 by the addition of 1N HCl. The juice was divided into aliquots and stored at −20°C until further use.

The intrinsic factor content of each sample of gastric juice was estimated by the use of the binding antibody in the assay described below and by the blocking antibody employing either albumin-coated charcoal (Gottlieb et al., 1965) or zirconium gel at pH 5.0 (Hansen et al., 1966).

Results

Observations on the combination of binding antibody with intrinsic factor-vitamin $B_{12}$ complex

To determine whether the combination of the binding antibody with intrinsic factor-vitamin $B_{12}$ complex could be used for assay of the complex, a detailed study of the reaction was undertaken using a fixed amount of antibody and increasing amounts of the complex. Radioactive vitamin $B_{12}$ was added to gastric juice in excess of its total binding capacity to saturate all available intrinsic factor molecules. The juice was diluted 1 in 10 with 0.15M NaCl and duplicate amounts ranging from 0.05 to 1.0 ml were added to two sets of test tubes. To one set, 0.1 ml of serum containing 0.5 ng units of the binding antibody was added, and to the other set 0.1 ml of control serum was added. Ten ml of zirconium gel (pH 6.25) was then added to each tube. The tubes were mixed in a vortex mixer, centrifuged at 1000 g for five min and the supernatants discarded. The gel was washed three times with 15 ml of 0.1M am-
monium acetate buffer at pH 6.25. The radioactivity adsorbed onto the gel in the tube with antibody serum minus that in the corresponding tube with the control serum represented the amount of intrinsic factor-\(^{57}\)Co vitamin B\(_{12}\)-antibody complex adsorbed onto the gel. The results are shown in Figure 1. Initially, the relationship between the amount of gastric juice and the amount of intrinsic factor-\(^{57}\)Co vitamin B\(_{12}\)-antibody complex adsorbed onto the gel was linear up to about 3-4 ng units. As the amount of intrinsic factor-\(^{57}\)Co vitamin B\(_{12}\) complex was further increased, the amount of radioactivity bound to the gel for a given increment in intrinsic factor-vitamin B\(_{12}\) complex decreased. Finally a plateau was reached when the amount bound to the gel was equal to the antibody content of the serum. Increasing the intrinsic factor-vitamin B\(_{12}\) complex still further produced no change in the amount of radioactivity bound to the gel.

THE ASSAY

It has been found that the relationship between the amount of gastric juice saturated with \(^{57}\)Co vitamin B\(_{12}\) used and the amount of intrinsic factor-\(^{57}\)Co vitamin B\(_{12}\) adsorbed onto the gel is always linear up to a value equivalent to approximately half the antibody content of the volume of serum employed. Using a volume of serum containing 5 ng units, the relationship is therefore linear to at least 2 ng units of intrinsic factor-\(^{57}\)Co vitamin B\(_{12}\) complex, although, at times, the linear range may extend further (Fig. 2). The protocol of a typical assay, for 1 volume of gastric juice to which excess \(^{57}\)Co vitamin B\(_{12}\) has been added, is shown in Table I, and the value obtained from this, together with values obtained from other volumes of the gastric juice, is shown in Figure 2.

When this assay is used simply to quantitate intrinsic factor in gastric juice, a single dilution of gastric juice giving a value within the linear range is adequate. For gastric juices with a probable normal content of intrinsic factor 0-1 ml of a 1 in 10 dilution gives adequate results. Juices which are likely to contain less than 20 ng units/ml are assayed undiluted.

REPRODUCIBILITY

The intrinsic factor of one gastric juice (measured as intrinsic factor-\(^{57}\)Co vitamin B\(_{12}\) complex) was determined by this assay 12 times in one day. The values ranged from 158 to 163 ng units/ml with a coefficient of variation of 0-8% (Table II). Another gastric juice was stored in multiple aliquots and one of these was assayed in duplicate on eight different

![Graph showing the relationship between volume of diluted gastric juice and amount of intrinsic factor-\(^{57}\)Co vitamin B\(_{12}\) bound to gel.](http://jcp.bmj.com/)

Table I  Assay protocol

<table>
<thead>
<tr>
<th>0.15M NaCl (ml)</th>
<th>Gastric Juice</th>
<th>Control Serum</th>
<th>Antibody Serum* (ml)</th>
<th>Zirconium Gel pH 6.25 (ml)</th>
<th>(^{57})CoB(_{12}) Bound to Gel</th>
<th>(^{57})CoB(_{12}) Bound by Binding Antibody*</th>
</tr>
</thead>
<tbody>
<tr>
<td>1.0</td>
<td>0.01</td>
<td>0.1</td>
<td>10</td>
<td>0.19</td>
<td>0.18</td>
<td></td>
</tr>
<tr>
<td>1.0</td>
<td>0.01</td>
<td>0.1</td>
<td>10</td>
<td>0.01</td>
<td>0.18</td>
<td></td>
</tr>
<tr>
<td>1.0</td>
<td>0.01</td>
<td>0.1</td>
<td>10</td>
<td>0.01</td>
<td>0.18</td>
<td></td>
</tr>
<tr>
<td>1.0</td>
<td>0.01</td>
<td>0.1</td>
<td>10</td>
<td>0.01</td>
<td>0.18</td>
<td></td>
</tr>
</tbody>
</table>

*Only 1 volume of gastric juice-\(^{57}\)Co vitamin B\(_{12}\) is shown; the protocol for other volumes is identical.

*The antibody serum used contained 50 ng units of binding antibody per ml.

*ng units adsorbed onto gel using antibody serum minus the amount adsorbed using control serum. In practice, because of the high degree of reproducibility, duplicates may not be necessary.
days. The values ranged from 145 to 162 ng units/ml with a coefficient of variation of 3.8% (Table II).

<table>
<thead>
<tr>
<th>Gastric Juice A</th>
<th>Gastric Juice B</th>
</tr>
</thead>
<tbody>
<tr>
<td>Date</td>
<td>ng units/ml</td>
</tr>
<tr>
<td>13-8-71</td>
<td>158</td>
</tr>
<tr>
<td>13-8-71</td>
<td>159</td>
</tr>
<tr>
<td>13-8-71</td>
<td>161</td>
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<tr>
<td>13-8-71</td>
<td>163</td>
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<td>13-8-71</td>
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<td>13-8-71</td>
<td>162</td>
</tr>
<tr>
<td>162</td>
<td>158</td>
</tr>
</tbody>
</table>

Table II  Reproducibility of the assay carried out on the same day (gastric juice A) and on eight different days (gastric juice B)

Comparison with other assays

The intrinsic factor content of 21 specimens of gastric juice, as measured by the present assay for intrinsic factor-vitamin B₁₂ complex, was compared with the results of intrinsic factor measurement using zirconium gel at pH 5.0 and the blocking antibody (Hansen et al, 1966). The results are shown in Figure 3. There is good correlation between the two assays (r = 0.99) although the binding antibody method gives a slightly lower value (approximately 5%) than the blocking antibody method.

The present assay was also compared with the albumin-coated charcoal assay (Gottlieb et al, 1965) (Fig. 4). Again there is good correlation between the results of the two assays (r = 0.99), although the binding antibody method gives values about 12% lower.

Use of assay in measuring intrinsic factor-¹⁵⁷Co vitamin B₁₂ in intestinal juice

Two volunteers, with normal vitamin B₁₂ absorption and normal secretion of intrinsic factor, were intubated orally with a fine radioopaque polyvinyl tube. When the distal end of the tube was in the upper ileum, a 1 μg dose of ¹⁵⁷Co vitamin B₁₂ (1 μc) was given by mouth. Commencing 45 minutes later, ileal juice was aspirated at intervals with a syringe and the radioactivity of the aspirates monitored. The two specimens containing the most radioactivity were centrifuged at 4°C at 10,000 g for one hour and the supernatants removed. None of the sediments contained any significant amounts of radioactivity. The radioactivity in the supernatants was present in
at least three forms: 'unbound' radioactivity which was measured and removed by treatment with albumin-coated charcoal; intrinsic factor-$^{57}$Co vitamin $B_{12}$ complex which was measured by the present assay; and $^{57}$Co vitamin $B_{12}$ bound to unknown non-intrinsic factor binders, which was estimated by subtracting the other two quantities from the total amount of radioactivity in the supernatants. Table III shows the results of this study on two specimens of ileal juice from each of two patients and Fig. 5 the results of the intrinsic factor-vitamin $B_{12}$ complex assay on one of the specimens.

Table III  Results of study of vitamin $B_{12}$ binding on ileal juice obtained after feeding radioactive vitamin $B_{12}$

<table>
<thead>
<tr>
<th>Time Specimen Obtained (hr after dose)</th>
<th>Total $ng^{57}CoB_{12}$/ml Aspirate</th>
<th>Unbound $^{57}CoB_{12}$ Removed by Albumin-coated Charcoal</th>
<th>Intrinsic Factor-$^{57}CoB_{12}$ Complex</th>
<th>$^{57}CoB_{12}$ Attached to Non-intrinsic Factor Binders</th>
</tr>
</thead>
<tbody>
<tr>
<td>Patient A</td>
<td>1-5</td>
<td>9-8</td>
<td>18% of Total</td>
<td>52% of Total</td>
</tr>
<tr>
<td></td>
<td>2-5</td>
<td>1-8</td>
<td>11% of Total</td>
<td>55% of Total</td>
</tr>
<tr>
<td>Patient B</td>
<td>1-0</td>
<td>1-6</td>
<td>19% of Total</td>
<td>69% of Total</td>
</tr>
<tr>
<td></td>
<td>1-5</td>
<td>0-2</td>
<td>25% of Total</td>
<td>65% of Total</td>
</tr>
</tbody>
</table>

Discussion

A detailed study of the relationship between the binding antibody and intrinsic factor-vitamin $B_{12}$ complex does not appear to have been previously undertaken. The results shown in Fig. 1 are typical of the combination of an antigen to an antibody which occurs in varying proportions. Using the initial part of the zone of antibody excess, the results given in this paper indicate that the reaction can be used to assay intrinsic factor-vitamin $B_{12}$ complex. On a given day, the assay has a high degree of reproducibility. When carried out on different occasions, on aliquots of the same specimen, the variation is slightly greater. The reasons for this are not yet clear but the variations are not such as to interfere with the value of the assay.

The intrinsic factor content of gastric juices measured by the present assay tends to be a little lower than the results obtained using the two assays employing the blocking type antibody although there is an excellent correlation between the assays. Bardhan et al (1967) also noticed that the results of the charcoal assay using the blocking antibody were higher than their results using the binding antibody with the electrophoretic retention technique. Schade, Feick, Imrie, and Schilling (1967) have shown that, at room temperature, blocking antibody can displace some of the vitamin $B_{12}$ already bound to intrinsic factor. It is possible that in the present assay the blocking antibody present in the binding antibody serum displaced some of the radioactive vitamin $B_{12}$ from its combination with intrinsic factor, and so reduced the amount of radioactivity bound to the gel. This would result in a lower intrinsic factor measurement with the binding antibody. It is also perhaps not surprising that the binding and blocking assays give slightly different results, since they depend on two different antibodies, the one active at the site of vitamin $B_{12}$ attachment to intrinsic factor and the other at some other site(s) on the intrinsic factor molecule. Of the two assays with the blocking antibody, the charcoal method gave the higher values. The explanation for this is not known. It was observed, however, that minor variations in the assay conditions, such as changing the brand of charcoal, or the amount of non-intrinsic factor.
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References


