Effect of *Bacteroides fragilis* on the human erythrocyte membrane: pathogenesis of Tk polyagglutination

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SYNOPSIS Incubation of normal human erythrocytes with the supernatant of centrifuged culture of some strains of *Bacteroides fragilis* results in the exposure of Tk polyagglutination determinants on the erythrocyte membrane. These determinants are present on non papain-labile structures and are probably exposed by an enzyme mechanism. Other strains of *B. fragilis* were found to produce neuraminidase, haemolysins, and protease. It is suggested that early recognition of Tk determinants may be an aid to the diagnosis of *B. fragilis* infection.

Polyagglutination is the agglutination by a variable number of normal sera of altered, defective or otherwise abnormal erythrocytes. This abnormality may be due either to adsorption of extraneous antigen by the erythrocyte or to structural peculiarities of the erythrocyte membrane itself (Bird, 1971).

In *vivo* polyagglutinability was first described by Levine and Katzin (1938). The membrane determinant responsible was named 'T' by Friedenreich (1930) who showed that it was present in a latent form on all normal human erythrocytes and could be exposed in the presence of 'enzyme-producing bacteria'. It was subsequently shown that the enzyme responsible for T exposure was neuraminidase (Gottschalk, 1960), which specifically cleaves N-acetylneuraminic acid (NANA) from the cell membrane and exposes \( \beta \)-galactosyl determinants (Uhlenbruck et al, 1969). Erythrocytes which carry these determinants are polyagglutinable since anti-T is present in most human adult sera. A number of secondary serological characteristics are associated with T antigen exposure. Loss of NANA results in a reduction in the net electronegative charge carried by the erythrocytes (Cook et al, 1961) and as a result they may not be aggregated by polycations such as protamine sulphate, Polysrene or polylysine which aggregate erythrocytes carrying a normal electronegative charge (Greenwalt and Steane, 1973). Also erythrocytes carrying exposed T antigens are agglutinated by a lectin extracted from peanuts (Bird, 1964).

In 1972 Bird and Wingham described a new form of polyagglutination, Tk. As shown in table I, the serological properties associated with T and Tk polyagglutination are similar but notable differences are observed in the effect of papain and of protamine sulphate. Earlier this year a second example of Tk polyagglutination was reported (Inglis et al, 1975). Preliminary laboratory investigations indicated that the agent responsible for exposure of the Tk determinant was probably the organism *Bacteroides fragilis*. A third example of Tk polyagglutination has since been reported (Bird et al, 1975) which demonstrated a definite involvement of this species.

<table>
<thead>
<tr>
<th>Agglutination by Arachis hypogea lectin</th>
<th>T Acquired</th>
<th>Tk Acquired</th>
</tr>
</thead>
<tbody>
<tr>
<td>Agglutination by Dolichos biflorus lectin</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>Aggregation by protamine sulphate</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>Effect of papain</td>
<td>Some destruction</td>
<td>Enhanced</td>
</tr>
<tr>
<td>Chemical basis of specificity</td>
<td>( \beta )-Galactose</td>
<td>( \beta )-Galactose</td>
</tr>
</tbody>
</table>

**Table I** Differentiation of T and Tk polyagglutination

In this report we present the results of *in vitro* experiments involving incubation of supernatants of cultures of different strains of *B. fragilis* with normal human erythrocytes and give a possible explanation of the mechanism involved in Tk exposure.
Effect of Bacteroides fragilis on the human erythrocyte membrane: pathogenesis of Tk polyagglutination

Material

ORGANISMS
Pure cultures of B. fragilis from a variety of human sources were incubated at 37°C for three days in cooked meat broth. The supernatant from centrifuged cultures was retained and stored at −20°C until tested. Supernatants from pure cultures of other genera were prepared in an identical manner. These included Escherichia coli, Staphylococcus aureus, Streptococcus faecalis, non-haemolytic streptococci, Pseudomonas aeruginosa, Proteus species, and Klebsiella aerogenes.

ERYTHROCYTES
Normal erythrocytes from blood donors were collected into acid-citrate-dextrose (ACD) anticoagulant. The erythrocytes were washed three times in 0.15 M sodium chloride and examined before use for the absence of polyagglutinability.

PEANUT LECTIN
80 g of peanuts were ground in a coffee grinder and suspended overnight at 4°C in 200 ml of 0.15 M sodium chloride. After centrifugation the aqueous layer was retained as stock reagent. Ten dilutions of stock (1/1 to 1/512) were prepared for use in 0.15 M sodium chloride.

PROTAMINE SULPHATE
Salmine sulphate (BDH Ltd) was prepared as a stock solution (100 mg/dl) in 0.15 M sodium chloride. Ten dilutions, 100 mg/dl–10 mg/dl, were prepared for use in 0.15 M sodium chloride.

NEURAMINIDASE
Neuraminidase, prepared commercially from Vibrio cholerae and assayed at 500 units/ml, was purchased from BDH Ltd.

PAPAIN
Papain solution was prepared by the method of Löw (1955) and stored at −20°C before use.

All serological experiments were carried out in 2 × 1 inch (50 × 6 mm) polystyrene tubes (Sterilin Ltd).

Methods and Results

TREATMENT OF ERYTHROCYTES WITH NEURAMINIDASE
Neuraminidase was diluted in 0.15 M sodium chloride to give a range of enzyme activity of 40 units/ml to 0.16 units/ml. One volume of washed, concentrated erythrocytes was added to one volume of each neuraminidase dilution and incubated at 37°C for 15 minutes. After centrifugation, the supernatants were recovered and NANA was assayed by the thiobarbituric acid method of Warren (1959). The erythrocytes were washed three times in 0.15 M sodium chloride and resuspended to a final concentration of 3–5% (v/v). Each erythrocyte suspension was tested with dilutions of neuraminidase in saline and in papain at 4°C for 1 hour (1 vol lectin + 1 vol papain (or saline) + 1 vol erythrocytes); 1 vol of erythrocyte suspension was also tested with 1 vol of protamine sulphate dilutions at 37°C for 30 minutes.

### Table II Erythrocytes treated with neuraminidase—pattern of agglutination with peanut lectin

<table>
<thead>
<tr>
<th>Concentration of Neuraminidase used to treat Erythrocytes (units/ml)</th>
<th>Medium</th>
<th>Medium</th>
<th>Dilution of Peanut</th>
<th>NANA (μ moles per ml, RBCs)</th>
</tr>
</thead>
<tbody>
<tr>
<td>40</td>
<td>Saline</td>
<td>+ + + + + + +</td>
<td>1 1 1 1 1 1</td>
<td>(+) 0.41</td>
</tr>
<tr>
<td>20</td>
<td>Saline</td>
<td>+ + + + + + +</td>
<td>1 1 1 1 1 1</td>
<td>(+) 0.36</td>
</tr>
<tr>
<td>10</td>
<td>Saline</td>
<td>+ + + + + + +</td>
<td>1 1 1 1 1 1</td>
<td>(+) 0.26</td>
</tr>
<tr>
<td>5</td>
<td>Saline</td>
<td>+ + + + + + +</td>
<td>1 1 1 1 1 1</td>
<td>(+) 0.14</td>
</tr>
<tr>
<td>2.5</td>
<td>Saline</td>
<td>+ + + + + + +</td>
<td>1 1 1 1 1 1</td>
<td>(+) 0.07</td>
</tr>
<tr>
<td>1.25</td>
<td>Saline</td>
<td>+ + + + + + +</td>
<td>1 1 1 1 1 1</td>
<td>(+) 0.06</td>
</tr>
<tr>
<td>0.63</td>
<td>Saline</td>
<td>(+) + + (+)</td>
<td>1 1 1 1 1</td>
<td>0.02</td>
</tr>
<tr>
<td>0.32</td>
<td>Saline</td>
<td>(+) + + (+)</td>
<td>1 1 1 1 1</td>
<td>0.01</td>
</tr>
</tbody>
</table>

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Results

The results are given in table II. The concentration of NANA released increased with increasing neuraminidase activity, as did the agglutinating activity of the erythrocytes with the peanut lectin. The erythrocytes treated with 0.63 and 1.25 units/ml neuraminidase were agglutinated by lectin in saline but not by lectin in papain. Erythrocytes treated with 2.5 units/ml neuraminidase or greater were agglutinated by lectin both in saline and in papain. As the concentration of neuraminidase increased, the strength of agglutination of the treated erythrocytes in lectin increased in both media but was always greater in saline than in papain for any given erythrocyte suspension. The capacity of the erythrocytes to aggregate in protamine sulphate decreased as increasing concentrations of NANA were cleaved from the cells.

TREATMENT OF HUMAN ERYTHROCYTES WITH SUPERNATANT OF B. fragilis CULTURE

The previous experiment was repeated substituting dilutions of B. fragilis culture supernatant in 0.15 M sodium chloride for neuraminidase.

Results

The results (table III) demonstrated that a different pattern of agglutination in lectin is obtained with culture-treated cells. Erythrocytes treated with high dilutions of culture supernatant were agglutinated by lectin, but only in the presence of papain. The agglutinating activity of the erythrocytes increased as the dilution of culture supernatant used to treat the cells decreased. By increasing the incubation time of erythrocytes in neat culture supernatant, cells were obtained which were agglutinated by lectin in papain and in saline. For any given erythrocyte suspension, agglutination by lectin was always greater in papain than in saline. NANA was not detected, either in the free state or as sialopeptide, in the supernatant after culture treatment of the erythrocytes. Aggregation of all the treated erythrocytes was normal in protamine sulphate.

INHIBITION OF PEANUT LECTIN BY B. fragilis CULTURE SUPERNATANT

Equal volumes of culture supernatant and dilutions of peanut lectin were incubated for three hours at 4°C. Each dilution of lectin was then tested for inhibition by testing with Tk erythrocytes both in saline and in papain at 4°C. Inhibition of lectin activity was not observed.

TESTS FOR AGGLUTINATION OF B. fragilis ORGANISMS WITH PEANUT LECTIN

A saline suspension of B. fragilis organisms was tested at 4°C with peanut lectin both in saline and in papain. The organisms were not agglutinated.

TESTS FOR OTHER STRAINS OF B. fragilis

Over 50 different strains of B. fragilis were cultured and the centrifuged supernatants examined using the techniques described earlier.

<table>
<thead>
<tr>
<th>Culture Dilution used to Treat Erythrocyte</th>
<th>Medium</th>
<th>Dilution of Peanut</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>1</td>
</tr>
<tr>
<td>Neat (60 min)</td>
<td></td>
<td>+++</td>
</tr>
<tr>
<td>Papain</td>
<td>Saline</td>
<td>+++</td>
</tr>
<tr>
<td>Saline</td>
<td>+++</td>
<td>+++</td>
</tr>
<tr>
<td>Papain</td>
<td>Saline</td>
<td>+++</td>
</tr>
<tr>
<td>Neat (45 min)</td>
<td>Saline</td>
<td>(+)</td>
</tr>
<tr>
<td>Papain</td>
<td>Saline</td>
<td>+</td>
</tr>
<tr>
<td>Neat (30 min)</td>
<td>Saline</td>
<td>-</td>
</tr>
<tr>
<td>Papain</td>
<td>Saline</td>
<td>+</td>
</tr>
<tr>
<td>Neat (15 min)</td>
<td>Saline</td>
<td>-</td>
</tr>
<tr>
<td>Papain</td>
<td>Saline</td>
<td>+</td>
</tr>
<tr>
<td>1/2</td>
<td>Saline</td>
<td>-</td>
</tr>
<tr>
<td>Papain</td>
<td>Saline</td>
<td>+</td>
</tr>
<tr>
<td>1/4</td>
<td>Saline</td>
<td>-</td>
</tr>
<tr>
<td>Papain</td>
<td>Saline</td>
<td>+</td>
</tr>
<tr>
<td>1/8</td>
<td>Saline</td>
<td>-</td>
</tr>
<tr>
<td>Papain</td>
<td>Saline</td>
<td>+</td>
</tr>
<tr>
<td>1/16</td>
<td>Saline</td>
<td>-</td>
</tr>
<tr>
<td>Papain</td>
<td>Saline</td>
<td>(+)</td>
</tr>
<tr>
<td>1/32</td>
<td>Saline</td>
<td>-</td>
</tr>
<tr>
<td>Papain</td>
<td>Saline</td>
<td>(+)</td>
</tr>
<tr>
<td>1/64</td>
<td>Saline</td>
<td>-</td>
</tr>
</tbody>
</table>

Table III Erythrocytes treated with Bacteroides fragilis culture—pattern of agglutination with peanut lectin
Effect of Bacteroides fragilis on the human erythrocyte membrane: pathogenesis of Tk polyagglutination

Results

Five different reaction patterns were observed:
(1) Haemolysis of the erythrocytes
(2) Exposure of T determinants
(3) Exposure of Tk determinants
(4) Exposure of (Tk + T) determinants
(5) Reduction of the electronegative charge without exposure of polyagglutination determinants, i.e., similar to protease treatment of erythrocytes.

Discussion

A concept of layers of antigen activity within erythrocyte membranes has been postulated by Uhlenbruck and Wintzer (1970), who found that N-acetylmuraminic acid can be detected both on glycoprotein and on glycolipid structures and that these structures are very differently distributed in the membrane. The data presented in table II support a layer hypothesis for T determinants and suggest that there are at least two areas of T activity, one on the outer surface of the membrane, carried on papain-labile structures, and the other on a lower layer of non-papain-labile structures. The concentration of neuraminidase available governs which determinants are exposed, papain-labile structures appearing to carry the dominant substrate since only these are exposed by low concentrations of neuraminidase. The conflicting reports in the literature of papain lability of T determinants are also explained by this model.

The results in table III indicate that Tk determinants are found only on non-papain-labile structures. When only small numbers of determinants are exposed, the use of papain is necessary to detect them. By sufficiently increasing the site density of exposed Tk determinants, the erythrocytes may be agglutinated by lectin without the aid of papain. The addition of papain enhances agglutination, suggesting that removal of surface peptide facilitates the reaction with lectin.

Tk determinants are similar to but not identical with T determinants. Both have similar β-galactosyl determinants which are quite different from other β-galactosyl determinants on the red cell membrane surface (Pardoe et al, 1971) so that erythrocytes carrying exposed T or Tk determinants are agglutinated by the peanut lectin. However, Bird and Wingham (1972) have shown that normal human serum absorbed with neuraminidase-treated erythrocytes can agglutinate Tk erythrocytes. Non identity is further demonstrated by the fact that exposure of Tk determinants is not accompanied by the release of NANA from the erythrocyte membrane.

It is probable that the mechanism responsible for Tk exposure is enzymatic since an attempt to inhibit the serological activity of peanut lectin by incubation with culture supernatant was unsuccessful, as was the attempt to identify Tk determinants on B. fragilis organisms themselves.

Not all strains of B. fragilis induce Tk exposure on normal erythrocytes. Some produce a neuraminidase and expose T determinants on normal erythrocytes. Others expose both T and Tk determinants. Haemolysis and protease-like activity were also observed in a few strains.

Although only a relatively small number of genera were tested, B. fragilis was the only species capable of inducing Tk exposure on erythrocytes. On both occasions in which Tk polyagglutination was associated in vivo with B. fragilis infection, death resulted. It may therefore be that Tk polyagglutination indicates a poor prognosis. On the other hand, since only a minority of patients with infection require transfusion therapy, the Tk phenomenon may not be as rare as statistics indicate, in which case early identification of Tk exposure may be useful as an aid to the diagnosis of B. fragilis infection.

References


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