An optimized semi-automatic rate method for serum glutathione reductase activity and its application to patients with malignant disease

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SYNOPSIS An improved and optimized method for serum glutathione reductase is described. The reference range for normal subjects is 47-79 IU/l. The method is more sensitive than conventional enzyme tests in the detection of malignant disease. It was not raised more frequently in patients with clinical evidence of metastases than in those clinically free of such metastases, and it did not seem to correlate with prognosis among those patients who failed to survive six months from the time the analysis was first conducted.

Glutathione reductase (GR; NAD(P)H: glutathione oxido reductase EC 1.6.4.2) is an important flavo enzyme involved, with glucose-6-phosphate dehydrogenase, in the maintenance of a reduced intracellular environment. The glucose-6-phosphate dehydrogenase provides NADPH which allows the reduction of oxidized glutathione (GSSG) using the following reaction:

\[ \text{GR} \]
\[ \text{GSSG} + \text{NADPH} + \text{H}^+ \rightleftharpoons 2\text{GSH} + \text{NADP}^+ \]

The equilibrium constant strongly favours the formation of reduced glutathione.

Clinical interest in GR has taken several forms. Screening for hereditary erythrocyte GR deficiency has been advocated (Beutler, 1966) although this does not necessarily lead to oxidant-induced haemolytic anaemias (Jaffé, 1968). The prosthetic group of GR is the riboflavin metabolite FAD. The extent of activation of red cell GR by exogenous FAD has been used as a test for riboflavin deficiency (Glatzle et al, 1968; Beutler, 1969; Sauberlich et al, 1972; Heller et al, 1974). Red cell GR is increased in cystic fibrosis patients (Shapiro et al, 1973).

Early work showed that the serum enzyme was elevated in many liver diseases (particularly infective hepatitis), pernicious anaemia, and often to a very high level in malignant disease, especially where liver metastases were present (Manso and Wróblewski, 1958; Kerppola et al, 1959; West et al, 1961; Horn et al, 1962).

As approximately 33% of the patients with cancer in these studies had elevated serum GR levels, and the instrumentation available to the clinical chemist has improved in the decade since these papers were published, the present study was initiated to determine whether, using an automated kinetic determination of GR, the assay was of value in cancer assessment.

Material and Methods

REAGENTS

1 Phosphate buffer (0.15 mol/l), pH 7.2 at 37°C. Store at −20°C
2 EDTA, disodium salt (15 mmol/l). Store at −20°C
3 NADPH, tetrasodium salt (10 mmol/l) Boehringer 15501. Prepare freshly each day
4 GSSG (65 mmol/l) Boehringer 15132. Prepare freshly each day
5 Bulk reaction mixture. This is made up each day and for 100 tests should comprise 110 ml Reagent 1, 5.5 ml Reagent 2, 2.75 ml Reagent 3, and 36 ml deionized water.

PROCEDURE

All assays were performed at 37°C with an LKB 8600 Reaction Rate Analyser (LKB Instruments,
Bromma, Sweden), using an absorbance back-off of 0.3A and a reading time of 1 min. Into each cuvet, 1.4 ml Reagent 5 and 50 μl of serum were added by an autodilutor. After 10 min incubation at 37°C the reaction was initiated by the addition of 50 μl of GSSG. The enzyme activity could be obtained from a recorder tracing by multiplying Δ A/min by a factor of 4823. In the present study automatic print-out of results was provided by the Optilab Multilog System (Bo Philip Instrumentation, Vallingby, Sweden).

PATIENTS
All subjects admitted to one unit of a cancer treatment centre over a six-month period had blood taken before commencement of therapy. This was centrifuged, and the serum was removed and stored at −20°C before analysis. Replicate analysis on 20 samples before and after storage for several months resulted in no significant change in activity. Aspartate transaminase (AST), lactate dehydrogenase (LDH), and alkaline phosphatase (ALP) activities were assayed using the SMA 12/60 (Technicon Instruments Ltd). Reference ranges for this laboratory are 10-50 IU/l, 100-225 IU/l, and 28-100 IU/l respectively.

Careful clinical assessment of the subjects was carried out, and the salient features of their disease are summarized in table I. Only one subject had overt hepatic metastases when first examined. Sites of metastases were superficial glands (6), peritoneal nodes (5), pelvis (4), bones (3), brain (3), bladder (1), lungs (1), and widespread (1).

TISSUE PREPARATION
Purified preparations of human erythrocyte GR were made using the method of Scott et al (1963). The source of human liver GR was a high-speed supernate (100 000 × g for 1 hr) of fresh postmortem liver homogenized 1:3 in ice-cold sucrose (0.25 mol/l).

<table>
<thead>
<tr>
<th>Location of Cancer</th>
<th>Males (%)</th>
<th>Mean Age</th>
<th>No. with</th>
<th>No Detectable Metastases</th>
</tr>
</thead>
<tbody>
<tr>
<td>Respiratory tract</td>
<td>95</td>
<td>60</td>
<td>4</td>
<td>17</td>
</tr>
<tr>
<td>Urogenital tract</td>
<td>56</td>
<td>52</td>
<td>6</td>
<td>10</td>
</tr>
<tr>
<td>Breast</td>
<td>0</td>
<td>59</td>
<td>1</td>
<td>9</td>
</tr>
<tr>
<td>Digestive tract</td>
<td>62</td>
<td>62</td>
<td>3</td>
<td>5</td>
</tr>
<tr>
<td>Reticulo-endothelial</td>
<td>72</td>
<td>49</td>
<td>5</td>
<td>2</td>
</tr>
<tr>
<td>Others</td>
<td>33</td>
<td>46</td>
<td>3</td>
<td>3</td>
</tr>
<tr>
<td>Total</td>
<td>22</td>
<td>46</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Table I   Classification of cancer patients at time Serum GR was first assayed

Results

CHOICE OF BUFFER
Initial studies showed that phosphate was superior to triethanolamine and PIPES (piperazine-N, N’-bis (2-ethane-sulphonic acid)), and that varying the molarity between 0.1 and 0.3 mol/l was without effect.

The inclusion of EDTA in the reaction mixture at a final concentration of 0.5 mmol/l enhanced enzyme activity and raised the pH optimum (fig 1). This shift in pH optimum occurring when EDTA is included in the reaction mixture allows GR from all sources to be analysed at pH 7.2, and this minimizes blank reactions due to spontaneous hydrolysis of NADPH.

Fig 1 Influence of EDTA at a final concentration of 0.5 mmol/l on activity of GR under the conditions of the proposed method upon serum from a patient with cancer and one with acute liver disease, and upon a red blood cell haemolysate (RBC).

Fig 2 Relation of NADPH concentration to serum GR activity, with all other conditions as in the proposed method.
An optimized semi-automatic rate method for serum glutathione reductase activity

CHOICE OF SUBSTRATE CONCENTRATION
NADPH was optimal at final concentrations above 0.12 mmol/l (fig 2). GSSG was optimal at final concentrations above 0.54 mmol/l (fig 3).

EFFECT OF SALTS ON SERUM GR
Increasing concentrations of potassium and sodium chloride were inhibitory (30% and 80% respectively at 0.1 mol/l). Potassium oxalate and acetate at the same concentration caused approximately 20% activation of the serum enzyme. Chloride ions were thus inhibitory to serum GR activity.

EFFECT OF ENZYME ACTIVATORS
Dithioerythritol was included in the reaction mixture to a final concentration of 2 mmol/l and had no significant effect on the activity of the serum enzyme.

FAD has been shown to activate red cell GR (Glatzle et al, 1968; Beutler, 1969). Inclusion of FAD in the reaction mixture up to a final concentration of 10 μmol/l (concentrations above this were slightly inhibitory) had no effect on 37 of 38 randomly selected sera. The single exception was activated by 20% and 65% at 2 and 10 μmol/l respectively. The patient concerned had a prostatic carcinoma, and riboflavin deficiency could not be ruled out.

CHARACTERISTICS OF THE PROPOSED METHOD
Enzyme activity is linearly related to the amount of serum taken, to an absorbance change of 0.310 per min. This corresponds to an activity of 1500 IU/l. Data for within-batch precision are given in table II.

<table>
<thead>
<tr>
<th>No. of Samples</th>
<th>Mean Activity (IU/l)</th>
<th>SD</th>
<th>CV (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>18</td>
<td>52.7</td>
<td>3.72</td>
<td>7.1</td>
</tr>
<tr>
<td>15</td>
<td>140.0</td>
<td>6.6</td>
<td>4.7</td>
</tr>
<tr>
<td>15</td>
<td>252.5</td>
<td>6.3</td>
<td>2.5</td>
</tr>
</tbody>
</table>

Table II Within-batch precision of GR assay

for normal subjects. Comparable data for the other enzymes were AsT 7 (10%), LDH 22 (31%), and ALP 13 (19%). The distribution of enzyme values in all subjects is shown in fig 4. Of the subjects with raised

<table>
<thead>
<tr>
<th>Percentage with Raised Serum Enzymes</th>
</tr>
</thead>
<tbody>
<tr>
<td>GR AsT LDH ALP GR &amp; LDH None</td>
</tr>
<tr>
<td>Dead (36)</td>
</tr>
<tr>
<td>57 20 48 29 19 29</td>
</tr>
<tr>
<td>Alive (32)</td>
</tr>
<tr>
<td>47 0 18 9 19 47</td>
</tr>
</tbody>
</table>

Table III Relation between abnormal enzymes and prognosis after six months

1No. of cases in parentheses
serum GR activity, 15% had an elevated AsT, 51% an elevated LDH, and 20% an elevated ALP. Of the 29 with a normal serum GR activity, only one had an elevated AsT, two an elevated LDH, and five an elevated ALP. Of those with a normal GR and elevated ALP, only one was known to have bony metastases.

Table III shows the outcome of the disease processes six months after the initial assessment. An enzyme elevation indicates a poorer prognosis; in particular, all seven patients with an AsT elevation died within six months of the analysis.

In view of previous emphasis on LDH as an indicator of possible malignant disease (Ticktin and Trujillo, 1970), a comparison was made of the data for this enzyme and for GR in the same patients (fig 5). In addition, an analysis was conducted on the regression and correlation between the two estimations in subjects grouped according to presence or absence of clinically detectable metastases (table IV). This showed that, on the whole, there was not a close relationship between the two. The factors leading to their elevation in cancer subjects therefore appear to be different, and this implies that more information can be obtained by assaying both than one alone.

Discussion

During the last decade there have been many attempts to find a laboratory test capable of identifying those patients with malignant disease in an apparently normal population. Several of these have involved the measurement of serum enzymes or isoenzymes, the two most popular in this group being LDH (Ticktin and Trujillo, 1970) and the Regan isoenzyme of ALP (Fishman et al, 1968). These suffer the disadvantages of poor specificity and a low incidence respectively.

Early work on serum GR showed that one-third of patients with neoplastic disease had elevated activity. The present study was initiated to determine the clinical value of this assay using improved methodology, since current methods proved to be suboptimal, unsuitable for application to reaction rate analysers, or optimized for the erythrocyte enzyme. The use of a reaction rate analyser for the estimation of erythrocyte GR has been described (Brewster et al, 1974). The method is optimized for very small sample volumes, since high concentrations of haemolsytes give unacceptably high initial absorbances, and is not directly applicable to serum.

A colourimetric method based upon the quantitative liberation of chlorpromazine from a preformed coloured palladium-chlorpromazine complex by the reduced glutathione produced in the reaction has been described (Lee et al, 1975). This method was independent of the initial absorbance and has been applied to the estimation of both serum and erythrocyte GR activities.

The inclusion of EDTA in the reaction mixture has long been part of erythrocyte GR assays, and its inclusion in the serum assay is justified by the increase in activity and the shift in pH optimum to a more alkaline region with improved stability of NADPH. Earlier methods employed conditions differing from those we found to be optimal, and a comparison of these with our own is presented in table V.

The observation that the serum enzyme is not activated by FAD, unlike the erythrocyte enzyme (Glatzle et al, 1968; Beutler, 1969), may reflect a higher extracellular FAD concentration. The GR activity of rat liver homogenates has been shown to be unaffected by exogenous FAD (Menendez et al, 1974).

The results obtained from patients with cancer demonstrate the superiority of serum GR estima-
tions over LDH in the diagnosis of malignant disease: these two enzymes are much more likely to be raised than either AsT or ALP. Analysis of the individual raised serum GR values showed them to be randomly distributed throughout the various categories of malignancy given in table I. It could not be used as an index of possible metastases.

The level of serum GR activity was of no value in assessing the eventual outcome, and in fact GR was a worse prognostic index than any of the other enzymes assayed. AsT was the most promising in this regard since all the patients who had an elevated AsT died within six months of the sample being taken. However, the high incidence of raised serum GR among cancer patients demonstrated in this study suggests that it may be of value in population screening. Since it is raised in liver disease and blood dyscrasias (Manso and Wróblewski, 1958; Kerppola et al., 1959; West et al., 1961; Horn et al., 1962), serum GR cannot be regarded as a cancer specific test, but it deserves to replace LDH in multiphasic biochemical profiling. A true assessment of its diagnostic value will emerge only when it is applied to a general hospital population rather than to the restricted case material of a cancer-treatment centre.

We thank Dr Helen Munro and Dr Zoe Randall for help in this study.

References


Table V  Comparison of existing methods for estimation of GR

<table>
<thead>
<tr>
<th>Reference</th>
<th>Sample</th>
<th>pH</th>
<th>Reagents¹</th>
<th>Buffer²</th>
<th>EDTA</th>
<th>NADPH³</th>
<th>GSSG</th>
<th>Temperature (°C)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Horn et al (1965)</td>
<td>Serum</td>
<td>6-6</td>
<td>54</td>
<td>—</td>
<td>0.40</td>
<td>0.5</td>
<td>37</td>
<td>25</td>
</tr>
<tr>
<td>West et al (1961)</td>
<td>Serum</td>
<td>7-5</td>
<td>8.7</td>
<td>0.07</td>
<td>5.7</td>
<td>37</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Long and Carson (1961)</td>
<td>Red cells</td>
<td>7-6</td>
<td>139*</td>
<td>34</td>
<td>5.3</td>
<td>37</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Brewster et al (1974)</td>
<td>Red cells</td>
<td>8-0</td>
<td>165*</td>
<td>2</td>
<td>3.3</td>
<td>35</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Lee et al (1975) (colourimetric assay)</td>
<td>Red cells and serum</td>
<td>7-4</td>
<td>34</td>
<td>1</td>
<td>2.3</td>
<td>37</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Present study</td>
<td>Serum</td>
<td>7-2</td>
<td>100</td>
<td>0.5</td>
<td>2.2</td>
<td>37</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

¹Final concentrations (mmol/l)
²Phosphate except where marked *, these being tris
³NADPH concentrations >0.25 mmol/l yield absorbances of >1.55, the maximum acceptable on the LKB