Congenital dyserythropoietic anaemia type II (HEMPAS): a family study

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SUMMARY  A family having two affected siblings with congenital dyserythropoietic anaemia type II (HEMPAS) is described. The proband was diagnosed after referral for investigation of haemolytic anaemia. Clinical evaluation and in vivo red cell (RBC) survival and the sequestration studies in the proband indicated that the anaemia was due to a combination of ineffective erythropoiesis and premature destruction of RBCs in the spleen. Scanning electron microscopic examination of peripheral RBCs was undertaken and is reported. The polypeptide composition of RBC membranes was also examined using polyacrylamide gel electrophoresis after solubilisation in sodium dodecyl sulphate. These results are also reported.

Congenital dyserythropoietic anaemias (CDA) are rare familial disorders characterised by the association of refractory anaemia with multinuclearity and bizarre nuclear abnormalities of erythrocyte precursors in the bone marrow. Three types of CDA are well recognised.1 Recently, a new type has been described.2 CDA type II has been called HEMPAS3 because of the positive acidified serum (Hams) test found in these patients. Other serological abnormalities found in HEMPAS are a high agglutination titre with anti-i, an unusual susceptibility to lysis by anti-i and anti-I, and a negative sucrose lysis test. The aetiology of the disease remains unknown but there is considerable evidence available to suggest a RBC membrane defect.3-5 Recently, Vainchenker6 has provided evidence that the defect in HEMPAS is due to an abnormal early erythroid precursor and is not the result of an environmental defect in the bone marrow.

Case report

The proband, GH, a 7-year-old boy, was referred to the haematology department for evaluation of probable haemolytic anaemia. The only prior medical history was that of a heart murmur at the age of 5 years. This was thought to be functional, secondary to a severe anaemia. Physical examination revealed a small child with a lemon yellow tinge to his skin. The forehead was prominent and the spleen was palpable 3 cm below the left costal margin. The liver was palpable 3 cm below the right costal margin.

Investigations showed haemoglobin 7.5 g/dl; MCV 80 fl; WBC 8.5 × 10⁹/l (normal differential), platelets 280 × 10⁹/l, reticulocytes 4%. The blood film showed anisocytosis and poikilocytosis, and pincer cells were seen (Fig. 1). Occasional nucleated red blood cells were also found in the bone marrow aspirate. Electron microscopy showed pincer cells, multinuclear erythroid cells, and pathologically abnormal mitochondria. These findings were consistent with HEMPAS. A bone marrow biopsy showed hyperplasia of the erythroid series with a high proportion of nuclei with abnormal morphology. The nucleated RBC count was 1.58 × 10⁹/l.

Received for publication 14 April 1980

Fig. 1  Peripheral blood showing 'pincer cells' (arrow). Jenner Giemsa × 1000 (oil immersion).
red cells were present. The serum bilirubin was 53 \(\mu\)mol/l and the LDH was 173 IU. Haptoglobins were not detected. The serum B12 and folate were normal and the direct antiglobulin test (Coombs) was negative. The serum iron was 36 \(\mu\)mol/l and the TIBC was 38 \(\mu\)mol/l. The osmotic fragility curve revealed a tail of slightly resistant red cells. A skull x-ray showed well-marked vertical striation of the trabeculae of the skull vault (Fig. 2). The chest x-ray was normal. The bone marrow was strikingly abnormal. Erythroid hyperplasia was present, and the majority of late normoblasts showed evidence of multinuclearity. The nuclear configuration was T H O II M H JH DH PH JH JH.

Fig. 3 Family tree showing proband and one affected sibling.

Material and methods

The annual blood consumption was calculated as outlined for patients with thalassaemia. Red cell survival and sequestration studies using Na\(_2^{51}\)CrO\(_4\) (sodium chromate) were performed using standard techniques. Scanning electron microscopy (SEM) Venous blood was taken without an anticoagulant and added dropwise immediately to a large volume of saline.
of 3% glutaraldehyde, which was constantly shaken. The sample was then processed, as described by Bessis10 and examined using an MSM-5 mini SEM.

RED CELL MEMBRANES
Venous blood was taken into lithium heparin and processed within 1 hour. Red cell ghosts were prepared,11 and electrophoresis after solubilisation in sodium dodecyl sulphate was carried out on a 10% polyacrylamide gel (PAGE SDS).12 The gels were stained with Coomassie brilliant blue R.250.13 The peptide profiles of patients and normal controls were analysed with respect to standard published profiles.14

SEROLOGY
Anti-i agglutination titres were performed at 20°C using washed red cells by Dr S Worlledge, Royal Postgraduate Medical School, London.

Results
Annual blood consumption by the proband was 270 ml/kg per year to give a mean haemoglobin of 9.7 g/dl. The expected blood consumption in thalassaemia major for a similar mean haemoglobin would be 220 ml or less.7

The red cell survival was reduced (Table 2), and there was evidence of some excess splenic sequestration.

SCANNING ELECTRON MICROSCOPY
SEM revealed two populations of red cells. The majority of RBC appeared morphologically normal, and a minority (approximately 10%) showed cytoplasmic projections (Fig. 4), which presumably correspond with the pincer cells seen under light microscopy.

PAGE SDS
Red cell membrane profiles from the proband were compared with normal controls (Fig. 5). Band 3 had a faster electrophoretic mobility than the control.

Discussion
There is considerable evidence that a membrane defect is present in erythrocytes of patients with HEMPAS. Abnormal binding of anti-i and increased lysis with anti-i and anti-I together with a positive Ham test with compatible sera may all result from

Table 1 Anti-i agglutination titre at 20°C

<table>
<thead>
<tr>
<th>TH father</th>
<th>MH mother</th>
<th>GH proband</th>
<th>DH affected sibling</th>
</tr>
</thead>
<tbody>
<tr>
<td>1:32</td>
<td>Nil</td>
<td>1:256</td>
<td>1:256</td>
</tr>
</tbody>
</table>

Fig. 4 SEM. Peripheral blood showing cytoplasmic projection.

Fig. 5 PAGE SDS 10% stained with Coomassie brilliant blue R.250 (bands 1 and 2 Spectrin). A = normal control (bands numbered according to Steck); B = proband; C = normal control. Band 3 has a slightly faster electrophoretic mobility than the control.

<table>
<thead>
<tr>
<th>MRCL (days)</th>
<th>T½Cr (days)</th>
<th>SEI %</th>
<th>LEI %</th>
<th>SLI %</th>
</tr>
</thead>
<tbody>
<tr>
<td>GH</td>
<td>34</td>
<td>17</td>
<td>41</td>
<td>1:21</td>
</tr>
<tr>
<td>Normal</td>
<td>70-150</td>
<td>25-32</td>
<td>1:1-28:7</td>
<td>1:4-10:2</td>
</tr>
</tbody>
</table>

MRCL = mean red cell life; T½Cr = red cell half life; SEI = spleen excess index; LEI = liver excess index; SLI = spleen liver index.
have demonstrated excessive endoplasmic reticulum binding of anti-i. Increased antigen abnormality in the red cell membrane in normoblasts and mature erythrocytes. Our SEM study shows abnormal erythrocyte cytoplasmic projections, and this presumably accounts for the poikilocytes (pincer cells) seen on light microscopy. As the cells were added immediately to glutaraldehyde without the addition of anticoagulant, this appearance is unlikely to have been due to artefact.

The use of polyacrylamide gel electrophoresis after solubilisation of red cell ghosts in sodium dodecyl sulphate (PAGE SDS) is a well recognised method of investigating red cell membrane proteins. The pattern obtained is highly reproducible, and the nomenclature of Steck is widely used. Anselstetter et al. used PAGE SDS to investigate a family with HEMPAS. Although a different numbering system of proteins was used it is evident from comparisons with Steck that an abnormality in band 3 was postulated. Band 3 represents the major polypeptide in the erythrocyte membrane. It is a glycoprotein and is the purported mediator of anion transport. Subcomponents of band 3 are linked to spectrin (bands 1 and 2) and actin (band 5), two proteins forming the cytoskeleton of the red cell. Erythrocyte membranes from the proband and his affected sibling were investigated on three occasions, and in all cases band 3 exhibited a slight but consistently faster electrophoretic mobility than the control. This may represent an altered molecular weight of this glycoprotein or could reflect abnormal glycosylation of band 3 in vivo. It is of interest that Childs et al. reported the presence of blood group I activity associated with band 3.

The red cell abnormality in HEMPAS gives rise to marked ineffective erythropoiesis. If the disease presents in childhood then skeletal abnormalities such as thickening of the diploë of the skull and perpendicular striations between the tables occur and may give rise to the so-called ‘hair on end’ radiological appearances.

Barosi et al. indicated that in some patients there may be a combination of shortened red cell survival together with ineffective erythropoiesis. In our proband, the red cell survival was shortened, and evidence of excessive splenic destruction was present. Modell studied a large number of thalassaemic patients and indicated that the annual blood consumption for patients with ineffective erythropoiesis could be estimated for a given mean haemoglobin value. A consumption in excess of the predicted figure was evidence of hypersplenism. Using these figures as a model for HEMPAS, the proband had an annual blood consumption slightly in excess of the predicted value. This would be in keeping with the modest reduction in red cell survival and slight excess splenic sequestration.

This family study is the first to be reported in Ireland. Although evidence suggests a red cell membrane defect the aetiology of the disease remains unknown. Further investigation of the cytoskeletal membrane proteins using more sophisticated techniques may help to elucidate the aetiology of this uncommon disease and to clarify whether the electrophoretic abnormalities of band 3 represent an alteration in the polypeptide composition or carbohydrate components of the major intrinsic protein of the erythrocyte membrane.

We thank Dr Anne O’Meara, Our Lady’s Hospital for Sick Children, Crumlin, Dublin for referring the proband for investigation; Dr S Worledge, Royal Postgraduate Medical School, London, for kindly carrying out the anti-i agglutination titre; Miss Margaret Milner for typing the manuscript; and Mr P Walsh for technical assistance with the scanning electron microscopy.

This work was supported in part by the Medical Research Council of Ireland.

References

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