T and B lymphocyte markers in effusions of patients with non-Hodgkin’s lymphoma

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SUMMARY

T and B cells were sought in effusion fluids of 13 patients with lymphoma. In T cell lymphomas (four cases) morphologically abnormal cells that formed E rosettes were present. In B cell lymphomas (nine cases) morphologically abnormal cells were present in only two cases, however immunological studies showed a reduction in T cells and monoclonal light chain immunoglobulin expression in six of nine cases. Patients with lymphocytic effusions often present a diagnostic problem. In particular the distinction of neoplastic from inflammatory lymphoid infiltrates may be difficult. In only about 10% of effusions from patients with lymphoma can abnormal cells be detected by cytological examination. Recently Domagala et al. have suggested that T and B lymphocyte enumeration may be useful in the diagnosis of lymphomatous infiltrates in effusions. In this report we give the results of studies on T and B lymphocytes in effusions of 13 patients with non-Hodgkin’s lymphoma.

Material and methods

Pleural or ascitic effusions were obtained from patients undergoing diagnostic taps or drainage for relief of symptoms. A total of 13 patients with lymphoid malignancy were studied. In cases 1–12 diagnoses were made by lymph node biopsy. In case 13 diagnosis was made by haematological examination of blood and bone marrow. A control group of six patients with non-lymphoid malignancy was also studied (two lung carcinoma, one metastatic seminoma, one pneumonia, one pulmonary embolus, one pericarditis). Samples were collected into sterile plastic universal containers containing EDTA (2%) or heparin (10 U/ml). Mononuclear cells were separated from effusion fluids by centrifugation over Ficoll-Hypaque. Mononuclear cells from the effusion Ficoll interface were washed twice in TC 199 (Flow Laboratories). Cytospin preparations were made for routine morphological assessment (May-Grünwald-Giemsa stains) and cytochemistry (non-specific esterase, acid phosphatase and chloroacetate esterase). T cells were identified by rosetting with sheep red blood cells and B cells by immunofluorescent staining with anti-human IgM, kappa and lambda antisera.

Results

The results of immunological studies on effusions are shown in the Table.

In cases of T cell lymphoma morphologically abnormal lymphoid cells were present in effusions of all the cases examined. Cases 1–3 were diagnosed as lymphoblastic lymphoma of T cell type on the basis of clinical features and histological and immunological investigations of lymph node, blood and bone marrow. In cases 1, 2 and 3 neoplastic lymphoblastoid cells in effusions formed E rosettes and exhibited focal acid phosphatase activity (Figs. 1, 2). Case 4 was diagnosed as an immunoblastic lymphoma by lymph node biopsy. In pleural fluid large immunoblastic cells that formed E rosettes were seen (Fig. 3).

In cases 5–12 diagnosis was established by histological examination of lymph nodes and immunological studies of lymph node and blood mononuclear cells. These were all B cell neoplasms. Of the B cell lymphomas, in only two cases (cases 5 and 7) did the effusion contain morphologically abnormal lymphoid cells. In case 5 these were large non-cleaved lymphoid cells (Fig. 4). In case 7 typical cleaved cells were present (Fig. 5). In both cases the neoplastic cells exhibited monoclonal surface immunoglobulin. In the other cases the effusions contained morphologically normal small lymphocytes. However even in these cases immunological investigations of the effusions showed an increased...
T and B lymphocyte markers in effusions of patients with non-Hodgkin’s lymphoma

Table 1

<table>
<thead>
<tr>
<th>Patient</th>
<th>Diagnosis</th>
<th>% E+</th>
<th>sIgM+</th>
<th>κ+</th>
<th>λ+</th>
<th>Polyclonal</th>
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<tr>
<td>1</td>
<td>Lymphoblastic lymphoma</td>
<td>92</td>
<td>1</td>
<td>-</td>
<td>-</td>
<td>Polyclonal</td>
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<tr>
<td>2</td>
<td>Immunoblastic lymphoma</td>
<td>76</td>
<td>1.5</td>
<td>0</td>
<td>0</td>
<td>Polyclonal</td>
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<tr>
<td>3</td>
<td>Immunoblastic lymphoma</td>
<td>86</td>
<td>0.5</td>
<td>0</td>
<td>0</td>
<td>Polyclonal</td>
</tr>
<tr>
<td>4</td>
<td>Centroblastic lymphoma</td>
<td>86</td>
<td>1</td>
<td>0</td>
<td>0</td>
<td>Polyclonal</td>
</tr>
<tr>
<td>5</td>
<td>Centroblastic lymphoma</td>
<td>79</td>
<td>2</td>
<td>0</td>
<td>0</td>
<td>Polyclonal</td>
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<tr>
<td>6</td>
<td>Centrocytic - diffuse</td>
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<td>0.5</td>
<td>0</td>
<td>0</td>
<td>Polyclonal</td>
</tr>
<tr>
<td>7</td>
<td>Centrocytic - diffuse</td>
<td>19</td>
<td>0.5</td>
<td>0</td>
<td>0</td>
<td>Polyclonal</td>
</tr>
<tr>
<td>8</td>
<td>Centrocytic - diffuse</td>
<td>19</td>
<td>0.5</td>
<td>0</td>
<td>0</td>
<td>Polyclonal</td>
</tr>
<tr>
<td>9</td>
<td>Centrocytic - follicular</td>
<td>19</td>
<td>0.5</td>
<td>0</td>
<td>0</td>
<td>Polyclonal</td>
</tr>
<tr>
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<td>Centrocytic - follicular</td>
<td>19</td>
<td>0.5</td>
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<td>0</td>
<td>Polyclonal</td>
</tr>
<tr>
<td>11</td>
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<td>0.5</td>
<td>0</td>
<td>0</td>
<td>Polyclonal</td>
</tr>
<tr>
<td>12</td>
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<td>0.5</td>
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<td>0</td>
<td>Polyclonal</td>
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<td>19</td>
<td>0.5</td>
<td>0</td>
<td>0</td>
<td>Polyclonal</td>
</tr>
<tr>
<td>Controls</td>
<td>(n = 6) Mean ± SD</td>
<td>84</td>
<td>6.5</td>
<td>2</td>
<td>6</td>
<td>Polyclonal</td>
</tr>
</tbody>
</table>

Cases 7, 11 were ascitic fluids, the remainder were pleural fluids. Monoclonality of light chain expression was determined from calculation of κ:λ ratio. A κ:λ ratio of > 4:1 or λ:κ of > 2:1 was considered abnormal (5, 6).

Discussion

Although in this study only a small number of cases have been examined the data show the potential of immunological assessment of effusion cells for the diagnosis and immunological typing of non-Hodgkin’s lymphoma and the distinction between lymphomatous and other lymphocytic infiltrates.

Fig. 1  Effusion cells from case 3 showing large lymphoblastic cells, some with convoluted nuclei. Cytospin preparation stained with May-Grünwald-Giemsa. Original magnification × 1000

Fig. 2  Effusion cells from case 3 showing focal acid phosphatase staining in lymphoblasts and strong perinuclear staining in a serosal cell. Cytospin preparation stained for acid phosphatase. Original magnification × 1000
In non-lymphomatous effusions the predominant lymphoid cell is a morphologically normal T lymphocyte with a small percentage of B cells. The percentages of T and B lymphocytes in the control group in this study are similar to those previously reported.\(^{3,7,8}\)

In patients with T cell lymphoma normal percentages of T cells were present, these however were morphologically abnormal in all the cases examined. In two cases of lymphoblastic lymphoma (2, 3) effusion cells showed high levels of Tdt activity further indicating an immature abnormal T cell population.\(^9\) In cases of T cell lymphoma less than 20% of the lymphocytes expressed surface immunoglobulin.

All cases of non-follicular B cell lymphoma showed a reduction in the percentage of T cells and increased B cells with monoclonal light chain expression, even in the absence of morphologically abnormal cells. These findings are similar to those of Domagala et al\(^{10}\) who in a study of four cases of chronic lymphocytic leukaemia and one centrocytic lymphoma found no morphologically abnormal cells but a marked reduction in T cells and increase in B cells compared to non-lymphomatous controls. In all the cases of diffuse B cell lymphoma we found over 20% of cells expressing surface IgM.

In the four cases of follicular centrocytic lymphoma studied only one case showed a reduction of T cells and an increased monoclonal B cell population. In the three other cases normal numbers of T cells were present, with B cells showing polyclonal staining for surface Ig. In only one of these three cases was the percentage of B cells above 20%. These findings suggest that the effusions may not have been neoplastic in origin.

This study has established that identification of T and B cells in effusions is useful in the diagnosis of lymphoma. This is especially the case in B cell lymphomas where morphologically abnormal cells may not be present but where immunological studies may clearly show evidence of a neoplastic infiltrate in effusion fluid. In patients with known lymphoma the distinction by immunological methods between lymphomatous and other infiltrates in effusion fluids

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Fig. 3  **Effusion cell from case 4 showing a large immunoblastic cell forming an E rosette. Cytospin preparation stained with May-Grünwald-Giemsa. Original magnification × 1000**

Fig. 4  **Effusion cells from case 5 showing (left) a large neoplastic lymphoid cell with cytoplasmic vacuolation and (right) a normal small lymphocyte forming an E rosette. Cytospin preparation stained with May-Grünwald-Giemsa. Original magnification × 1000**

Fig. 5  **Effusion cells from case 7 showing numerous cleaved cells (centrocytes). Cytospin preparation stained with haematoxylin. Original magnification × 1000**
may be of importance not only in staging of disease but also in subsequent clinical management.

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References


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