Technical method

Histological demonstration of platelet thrombi with a double silver impregnation argyrophil stain

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Thrombi vary in composition with differing percentages of fibrin, platelets, and red blood cells. Of these components, only fibrin is routinely demonstrated by histological staining techniques. We describe the use of a double silver impregnation argyrophil stain to demonstrate platelet thrombi.

Case report

A 74-year-old retired metal worker presented with gangrene in the left 1st, 2nd, and 3rd toes and the left 4th finger. The platelet count was normal (210 × 10^9/1) but spontaneous platelet aggregation was present on aggregometry; the platelet aggregate ratio was low (0-45) using the method of Wu and Hoake; the presence of circulating platelet aggregates was confirmed by the inspection of a peripheral blood film and the β-thromboglobulin concentration was raised (83 ng/ml). These results were considered indicative of intravascular platelet aggregation. The peripheral blood count and other coagulation tests were normal. Cardiac ultrasound examination was normal.

HISTOLOGICAL STUDIES

Light microscopy of the digital vessels in non-gangrenous areas showed luminal occlusion by faintly eosinophilic granular material. This stained negatively for fibrin with both Martius Scarlet Red and phosphotungstic acid haematoxylin.

Examination of the vessels by electron microscopy (Philips EM400) showed that the material consisted of platelet thrombus (Fig. 1) with minimal amounts of fibrin. An interesting feature was the presence of scattered granules in the cytoplasm of many platelets. The presence of electron-dense granules in unstained sections and the identification of calcium with x-ray microanalysis (EDAX system with EPIC computer) indicated that the granules included dense bodies.

As platelet granules are known to contain various amines, including 5-hydroxytryptamine, histamine, and catecholamines, it was considered possible that the granules may be demonstrable by stains normally used for APUD granules. Consequently, paraffin sections of the vessels were stained by the diazo reaction and Masson-Fontana for argentaffin granules and by a modification (as below) of Pascual's double silver impregnation technique for argyrophil granules. Negative staining was obtained with the argentaffin stains but strongly positive granular staining was revealed with silver impregnation method (Fig. 2).

Platelet-rich plasma was prepared from whole blood by centrifugation at 140 g for 10 min. When this was stirred in an aggregometer, aggregation was seen to occur. Ultrastructural examination of these aggregates showed central platelets containing dense bodies and alpha granules; the peripheral platelets were degranulated (Fig. 3). Light microscopy of aggregates from the same aggregometer cuvette showed positive argyrophil staining in the centre of the aggregates.

Material and methods

MODIFICATION OF PASCUAL'S SILVER IMPREGNATION METHOD

1. Take sections to water and rinse well in distilled water.
2. Treat in preheated 5% silver nitrate at 60°C for 45 min.
3. Rinse briefly in distilled water.
4 Reduce in preheated Bodian's reducer (hydroquinone 1g; sodium sulphite crystals 10g; distilled water 100 ml) at 60°C for 5 min.
5 Wash well in tap water; rinse in distilled water.
6 Treat sections for a further 20 min in preheated 5% silver nitrate at 60°C.
7 Rinse briefly in distilled water.
8 Reduce as in step 4.
9 Wash well in tap water.
10 Tone in 1:10 000 gold chloride in distilled water for 10 min.
11 Wash in water.
12 Fix in 5% sodium thiosulphate (hypo) for 2 min.
13 Wash well in water.
14 Stain nuclei in nuclear fast red for 3 min (nuclear red 0.1g; aluminium sulphate 5g; distilled water 100ml; Method: add nuclear fast red and aluminium sulphate to the distilled water, boil for 3 min, cool and filter).
15 Rinse in water, dehydrate, clear and mount in resin.

Results

Argyrophil granules – black
Nuclei – red.

Discussion

The positive argyrophil reaction in this case correlated with the presence of granulated platelets in the thrombi. The latter finding was unexpected as it indicated that the platelets had undergone aggregation but not secretion. Ultrastructural examination of thrombi in other diseases has shown different degrees of platelet degranulation and a corresponding variability in the success of argyrophil positivity. However, consistently positive results have been observed in other cases with thrombi associated with spontaneous platelet aggregation and circulating platelet aggregates. Thrombi containing largely fibrin give a negative reaction indicating that the positive result was not due to non-specific silver precipitation.

The exact cause of the positive argyrophil reaction remains uncertain. However, the negative argentaffin reaction indicates that it is due to amines other than 5-hydroxytryptamine.

Although platelet function can be tested in vitro by both mechanical and biochemical methods, techniques for measuring in vivo platelet function are too invasive to be used in man. There is much indirect evidence that platelets are causally involved in arterial thrombosis and the staining method described here is a histological tool that can demonstrate platelets to be a cause of vascular occlusion.
Letters to the Editor

Word and data processing in histopathology laboratories

As judged by the number of visitors and enquiries about the system in our laboratory, there is a considerable interest around the country in word and data processing facilities for histopathology laboratories. Accounts of our system have appeared elsewhere\(^1\) but we felt that many potential users of such systems might be interested in the sorts of problems which will almost inevitably arise when a computer system is installed and developed.

Installation
There were initial problems over the wiring from the computer to the various terminals. The Department of Medical Physics offered to do this, but the hospital administration felt that this was politically unwise in view of possible union objection. In consequence the hospital Works Department performed the job, which was done admirably, but four weeks after the Department of Medical Physics could have completed it.

The instruction manuals and system documentation accompanying computers and commercial software are seldom complete or written in a concise, comprehensible form. Our word processing manual was, in places, particularly bad and took a lot of understanding. The suppliers now have a much improved version.

Software
Software problems are the main headache in any computer development. The commercially purchased programs are never totally error-free. Of course when one is originally looking around the market, what is available, salesmen are not at pains to point these out. These errors, or bugs, must be overcome by the scientific staff. This staff cannot be expected to have grasped every last limitation of the programs before purchase, just as the clinical staff cannot be expected to have totally realised what they require of the system. These two groups of people must be in constant and close collaboration as the system is developed so that the laboratory actually ends up with a system which is good both in the computer scientist’s eyes and in the pathologist’s eyes. This collaboration is crucial and is why we believe that a good system cannot be developed quickly if the scientific staff are not resident in the hospital. This close liaison has allowed the development of a sophisticated set of coding, data storage, and archiving programs. These, as indicated above, were not devoid of developmental problems. A very simple example is shown in the fact that although 1981 was used as a developmental year, all the histological programs were running and error free, in mid-November, 10 months after the installation of the equipment. Efforts were then concentrated on other programs, only to realise, just after Christmas, that the laboratory specimen numbering format of XXXX81 would produce problems at the change over of the

References

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