Relation between antithrombin III and clinical and serological parameters in systemic lupus erythematosus

MARK P JARRETT, * DAVID GREEN, † CHUNG-HSIN TS’AO‡

From *Section of Arthritis-Connective Tissue Disease and the †Atherosclerosis Program, Department of Medicine, and the ‡Department of Pathology, Northwestern University Medical School, Northwestern Memorial Hospital and the Rehabilitation Institute, Chicago, Illinois, USA

SUMMARY The increased frequency of thromboembolic events in patients with systemic lupus erythematosus (SLE) has been attributed to reduced or dysfunctional antithrombin III (At-III). We analysed At-III values, measured by three different assay techniques, in SLE patients, patients with rheumatoid arthritis, and normal and hospitalised controls. In addition, attempts were made to correlate At-III activities of SLE patients with specific clinical and serological parameters such as disease activity, renal involvement, previous thrombosis, degree of proteinuria, and serum complement concentrations. Our results failed to show a significantly reduced At-III in SLE with any method. At-III titres did not correlate with disease activity, concentrations of serum complement or albumin (both only minimally reduced in most patients), or a previous history of thrombosis. At-III deficiency does not appear to be an inherent feature of SLE, and reduced activities should only be anticipated when there are specific aetiological factors present, such as massive proteinuria, extensive hepatic disease, or active thrombosis.

The activity of antithrombin III (At-III) has been shown to be highly correlated with a thrombotic diathesis. Congenital deficiency of this protein is associated with recurrent episodes of arterial and venous thrombosis. Acquired deficiency is found in a number of disease states. Systemic lupus erythematosus (SLE) is associated with an increased risk of thromboembolic complications, even in the presence of thrombocytopenia and circulating anticoagulants. Two recent studies described reduced or functionally abnormal but immunologically normal At-III in this disease. Earlier, Angles-Cano et al reported normal activities of At-III in SLE. In order to gain further insight into the At-III activities in SLE and their possible variation with disease activity and medication, we determined At-III in a group of SLE patients, using both functional and immunological techniques. Hospitalised non-connective tissue disease patients and patients with rheumatoid arthritis (RA) were studied concomitantly.

Subjects and methods

Subjects

Normal subjects

Plasma samples from healthy volunteers of both sexes were used to establish standard curves and to obtain normal At-III values. Sixteen subjects were examined with functional assays and 25 with an immunological method. None of the women donors was taking oral contraceptives for at least one month prior to assay, and none was pregnant.

SLE patients

Thirty-one samples were obtained from 26 patients with SLE who met at least four criteria of the American Rheumatism Association. The group consisted of 24 women and 2 men; 22 were white and four black. Their mean age was 37 ± 13 yr. None of the female patients was taking oral contraceptives. Types of medication, serum complement concentrations, serum albumin, presence of renal disease or protein-
uria, and previous history of thromboembolic diseases were recorded.

**RA patients**
Fifteen patients with definite or classical rheumatoid arthritis (10 women; 13 white and two black) were also studied. Their mean age was 58 ± 9 yr. Twelve of the patients were rheumatoid factor-positive. Thirteen patients were taking non-steroidal anti-inflammatory agents, five on gold, four on penicillamine, three on hydroxychloroquine, and one on azathioprine.

**Hospitalised patients**
Twenty-eight consecutive hospitalised patients had their At-III activities determined as part of their diagnostic evaluation.

**Blood sampling**
Blood, drawn into citrate-containing vacutainers (Becton-Dickinson, Rutherford, NJ), was centrifuged at 1100 g for 15 min at 4°C. The supernatant plasma was frozen at −20°C and assayed for At-III within 2 wk.

**AT-III assays**

**Amidolytic assay**
The assay is based on the inhibition by diluted plasma of the hydrolysis of a fluorogenic synthetic substrate (D-phenylalanine-proline-arginine-5-aminoisophthalic acid, dimethylester) by a known amount of thrombin in the presence of optimal concentrations of heparin. Reagents for this assay were purchased from the American Dade Corporation, (Miami, Florida). Standard curves were not required for the amidolytic assay. At the beginning of each run, the fluorometer (Protopath, American Dade Corp) was calibrated with a solid reference standard. Then, measurements were made with a blank, representing 0% At-III activity, and pooled normal human plasma (PNP), representing 100% activity. At-III activities in test plasma were calculated according to the formula:

\[
\text{% thrombin inhibited} = \frac{\text{blank reading} - \text{PNP reading}}{\text{blank reading}} \times 100
\]

\[
\text{% At-III test plasma} = \frac{\text{PNP reading} - \text{test plasma reading}}{\text{PNP reading}} \times 100
\]

**Anticlotting assay**
This was a two stage assay. Stage 1 involved neutralisation of a quantity of known thrombin by At-III in defibrinated (56°C, 5 min) test plasma. In stage 2, the neutralised sample was mixed with human fibrinogen and the clotting time determined. The longer the clotting time, the higher the At-III activity in the defibrinated plasma. Reagents for this assay were obtained from Ortho Diagnostics Inc (Raritan, New Jersey). Standard curves for the assay were established with each kit of reagents, using a standard human plasma provided with the kit.

**Immunological assay**
The technique of rocket electroimmunoassay was used. The personnel who performed these tests had no knowledge of the functional assay results. At-III activities of individuals determined by three assay techniques. Bars indicate mean and standard deviation for each group of subjects.
Antithrombin III in SLE

Comparison of At-III activities in SLE patients with or without certain clinical and serological manifestations

<table>
<thead>
<tr>
<th>Disease activity</th>
<th>Proteinuria</th>
<th>Serum complement</th>
<th>Renal involvement</th>
<th>Previous thrombosis</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>At-III Mean ± SD (%)</strong></td>
<td>122.8 + 119.3 + 115.8 + 111.7 + 114.2 + 138.8 + 119.7 + 118.6 + 114.5 + 132.2 + 117.8 + 137.8</td>
<td></td>
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<tr>
<td>± 22.6 + ± 17.5 + ± 10.3 + ± 10.6 + ± 15.0 + ± 20.7 + ± 19.2 + ± 19.8 + ± 15.9 + ± 20.1 + ± 17.7 + ± 17.5</td>
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<tr>
<td><strong>At-III Mean ± SD (%)</strong></td>
<td>85.8 + 95.7 + 86.3 + 91.0 + 88.1 + 98.4 + 89.7 + 90.0 + 87.7 + 90.0 + 88.1 + 94.4</td>
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<tr>
<td>± 4.4 + ± 9.7 + ± 6.3 + ± 8.6 + ± 15.3 + ± 12.0 + ± 17.5 + ± 13.6 + ± 12.2 + ± 15.3 + ± 12.9 + ± 16.3</td>
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</tr>
</tbody>
</table>

| No of patients | 10 | 9 | 4 | 3 | 21 | 12 | 14 | 17 | 9 | 22 | 4 |

* Determined by an amidolytic assay.
† Determined by an immunologic assay.

Antisera were purchased from Calbiochem-Behring Co (La Jolla, California). Standard curves for the assay were established using plasma pooled from 20 normal donors.

**Statistical Analysis**

Data were analysed by Student’s t test, the χ² analysis and linear regression.

**Results**

Individual At-III activities of all four groups of subjects, and the means and standard deviations as determined by the three techniques are presented in the Table. For both the normals and hospitalised patients, there was a significant correlation among At-III values assayed by these methods (r = 0.60, p < 0.01). In the SLE group, the two functional assays correlated with each other (r = 0.82, p < 0.001), but neither correlated with the immunological assay (r = 0.33, p > 0.10). In the RA group, either functional assay correlated with the immunological assay (r = 0.71, p < 0.001), but there was no correlation between the functional assays (r = 0.16, p > 0.10). Nevertheless, all values derived from the functional assays for RA patients were within the range of normal controls.

In determining whether there might be a positive relation between At-III activities and specific clinical problems in SLE patients, we compared At-III values with various clinical and serological parameters. Results are shown in the Table. There was no correlation between disease activity and the At-III activities. Only one patient with a low immunological value (70%) had + + + disease activity. At-III values of 14 patients with reduced complement titres were comparable to the normocomplementaemic group. Nine patients with documented renal disease (active urinary sediment or positive renal biopsy), and 5 patients with proteinuria (0.3 to 5.9 g/24 h urine), had At-III values similar to those without renal involvement or proteinuria. Furthermore, there was no correlation between serum albumin concentrations and At-III. Four patients with a history of thromboembolic disease also had normal levels of At-III. None of the SLE patients had active thromboembolic complications at the time of the assay. Some SLE patients were receiving prednisone (up to 60 mg/day; mean dose of 10.5 ± 15 mg/day). There was a significant correlation between the steroid dose and immunological activities of At-III (r = 0.63, p < 0.001).

One patient with RA had a low immunological At-III (66%). However, there were no unusual clinical or serological features in this patient, and she had no history of thromboembolic disease.

**Discussion**

At-III, an α₂-globulin, is a major inhibitor of activated clotting factors, and is responsible for heparin cofactor activity. Deficiency of this globulin, either congenital or acquired, leads to a state of hypercoagulability. SLE is often associated with such a state. According to one report, almost 25% of the 35 SLE patients developed clinically significant thrombosis. Another report puts the figure around 10%. The precise mechanism underlying the development of thrombosis in SLE is not clear. Angles-Cano et al attributed this tendency to an increased factor VIII-related antigen level in plasma. On the other hand, the finding that some SLE patients with proteinuria showed reduced At-III titres, and one SLE patient had dysfunctional At-III, imply a quantitative or qualitative defect in At-III.

Because of the critical antithrombotic role of At-III, we elected to re-examine the plasma activities of this protein in a group of SLE patients using three different assay techniques. Such a broad approach would provide us with a more accurate picture of At-III values by minimising the shortcomings inherent in a single procedure. It is particularly warranted in
light of reports of discrepant results obtained by functional and immunological techniques in SLE and in other diseases. We also determined At-III titres in a group of rheumatoid patients to see whether the defect in At-III would be specific for SLE, or also be present in closely related connective tissue disorders. We assayed At-III in a group of hospitalised patients to ensure that each of the techniques we employed was capable of detecting reduced activities of At-III.

Statistically, our assay methods did not always correlate among themselves for all groups of subjects. Nevertheless, a sample showing reduced At-III by one method was usually low by another method. Within a given method, the mean At-III value for SLE patients was not significantly different from that of normal controls. In addition, with the exception of steroid intake, the At-III values of SLE patients appeared to have no relation to any of the clinical or serological parameters we measured, regardless of whether these values were obtained by an amidolytic or an immunological assay. It should be pointed out that none of our patients had active thrombosis at the time of the assay. The highest protein content in the urine of the five positive proteinuria patients was 5.9 g/24 h, far less than the protein concentrations in urine samples of most of the At-III deficient patients described by Kaufmann et al. None of our patients had serum albumin concentrations below 2.9 g/l. We therefore conclude that At-III deficiency does not appear to be an inherent feature of SLE. Reduced activities should probably be anticipated only when specific aetiological factors are present. These factors may include severe proteinuria, extensive hepatic disease and active thrombosis.

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References


Requests for reprints to: Professor C. Ts'ao, Department of Pathology, Northwestern Medical Center, Wesley Pavilion, 433 East Superior Street and Fairbanks Court, Chicago, Illinois 60611, USA.