Morphological heterogeneity in childhood B cell acute lymphoblastic leukaemia

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SUMMARY Considerable heterogeneity of lymphoblast morphology in childhood B cell acute lymphoblastic leukaemia has been observed. One case showed unusual monocytic features and emphasised the need for marker studies in the accurate definition of acute lymphoblastic leukaemia phenotypes. B cell acute lymphoblastic leukaemia is rare but may have been previously underestimated by morphological misinterpretation. Further information is required to determine if the different morphological features of this condition are of clinical and prognostic importance.

In 1976 the French-American-British Cooperative Group classified acute lymphoblastic leukaemia into three morphology groups, L1, L2, and L3, using Romanowski stained films. Different observers consistently identified L1 lymphoblasts but considerable variation between observers occurred in assigning lymphoblasts to L1 and L2 categories. L1 morphology has subsequently been reported as characteristic of childhood B cell acute lymphoblastic leukaemia (B ALL). There is otherwise a poor correlation between lymphoblast morphology and immunological phenotype. We report four cases of B ALL in children. The morphology displayed is more varied than previously recognised.

Material and methods

Blood was taken by venepuncture and placed in plastic tubes containing lithium heparin (LIP Equipment Services Ltd). Marrow was aspirated and placed immediately into lithium heparin tubes containing tissue culture medium (RPMI 1640 Gibco Europe Ltd) or used to make direct films, which were subsequently stained with Wright’s stain for diagnostic morphological examination.

For lymphocyte separation blood was diluted with an equal volume of tissue culture medium at 37°C and layered gently on to 3 ml of Lymphoprep (Nyeguard and Co, AS Oslo) in a conical centrifuge tube (Sterilin Ltd, England) and centrifuged at 400 g for 25–30 min. The interphase containing the mononuclear cells was collected and washed three times with tissue culture medium at 37°C. The washed cells were resuspended in tissue culture medium containing 5% heat inactivated fetal calf serum (Sera-Lab Ltd, Sussex, England) in a final concentration of 2.5 × 10⁶ cells/ml.

Surface membrane marker studies were performed in our own laboratory (see method) unless otherwise stated. The percentage of T cells was determined by the spontaneous formation of rosettes with sheep erythrocytes and that of B cells by demonstrating surface immunoglobulin using fluorescein conjugated antihuman immunoglobulin (Wellcome Reagents Ltd, Beckenham, England). Monoclonal antisera OKT11 (Ortho Diagnostics Systems, New Jersey, USA) and B1 and I2 (Coulter Electronics Ltd, Luton, England) for the detection of the B1 antigen and Ia like or HLA-DR antigens, respectively, were used according to the manufacturer’s instructions.

PATIENTS Table 1 shows the clinical and haematological features of the four patients at presentation. Additional information about these patients is as follows.

Patient 1 died shortly after admission with severe metabolic complications, having also developed a pulmonary fat embolism. Postmortem examination showed diffuse involvement by lymphoblasts of bone marrow, lymph nodes, thymus, lungs, liver, spleen, kidneys, testes, pancreas, and parotids. Histological examination of these organs showed a ‘starry sky’ appearance as seen in Burkitt’s lymphoma.
Patient 2 was admitted with a normal blood count in acute renal failure. When peritoneal dialysis was started, severe intra-abdominal haemorrhage occurred. At laparotomy a large ovarian tumour was found to be the site of haemorrhage. Histological examination of the excised tumour showed a "starry sky" appearance consistent with a Burkitt's lymphoma. The aspirated marrow showed replacement of normal haemopoiesis by L3 type lymphoblasts. The patient died in acute renal failure with peritonitis after eight days of induction chemotherapy.

Patient 3 developed severe metabolic complications with a rapidly rising blood urea concentration, hypocalcaemia, hyperphosphataemia, and hyperglycaemia. Paroxysmal episodes of hypertension were followed by a large non-fatal intracranial haemorrhage, which subsequently required surgical evacuation. At this stage the patient entered remission, but unfortunately she was left with a hemiplegia and partial cortical blindness. She subsequently developed a meningeal relapse, from which she died.

Patient 4 contrasted with the other cases by presenting with a high white cell count (see Table 1) and by having no metabolic problems during induction. A necrotising pseudomonas infection of the vulva was successfully treated with antibiotics and white cell infusions. She remains in remission 18 months after diagnosis following treatment according to the UKALL VIII protocol for children with acute lymphatic leukaemia.

**Results**

The morphological appearances of the blast cells from each patient are shown in Figs. 1-4. The large blast cells from patient 1 (Fig. 1) show the typical features of L3 lymphoblasts, intensely basophilic cytoplasm due to the high content of RNA, and the prominent vacuolation of the lymphoblasts. Those from patient 2 (Fig. 2) also fulfil the criteria for L3 type. The blast cells from patient 3 (Fig. 3), although displaying a deeply basophilic cytoplasm, lack the characteristic vacuolation of L3 cells; therefore, the B cell nature of the acute lymphoblastic leukaemia could have been overlooked. The morphology of the blast cells from case 4 (Fig. 4) shows no L3 features; there is abundant pale blue/grey cytoplasm with minimal vacuolation and a large oval nucleus with open chromatin and prominent nucleoli. Our first impression was that these cells morphologically resembled monoblasts. Cytochemical esterase activity, a characteristic of monoblasts,* was absent.

Surface marker studies confirming the B cell nature of the four cases are summarised in Table 2.

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**Table 1** Presenting clinical features and haematological findings of four cases of B cell acute lymphoblastic leukaemia in childhood

<table>
<thead>
<tr>
<th>Patient no</th>
<th>Sex</th>
<th>Age (yr)</th>
<th>Lymphadenopathy</th>
<th>Jaundice</th>
<th>Hepatomegaly (cm)</th>
<th>Splenomegaly</th>
<th>Ascites</th>
<th>Other features</th>
<th>Haemoglobin concentration (g/dl)</th>
<th>Platelet count (x 10⁹/l)</th>
<th>White cell count (x 10⁹/l)</th>
<th>Blast cell count (x 10⁹/l)</th>
<th>Histology</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Boy</td>
<td>3</td>
<td>Axillae</td>
<td>Nil</td>
<td>5</td>
<td>6</td>
<td>Nil</td>
<td>Renal enlargement</td>
<td>16-8</td>
<td>35</td>
<td>35-5</td>
<td>1-7</td>
<td>Diffuse organ involvement*</td>
</tr>
<tr>
<td>2</td>
<td>Girl</td>
<td>13</td>
<td>Nil</td>
<td>Nil</td>
<td>Nil</td>
<td>Nil</td>
<td>++</td>
<td>Ovarian tumour</td>
<td>11-3</td>
<td>45</td>
<td>4-2</td>
<td>0-3</td>
<td>Ovarian tumour at laparotomy*</td>
</tr>
<tr>
<td>3</td>
<td>Girl</td>
<td>8</td>
<td>Axillae Cervical</td>
<td>Nil</td>
<td>++</td>
<td>6</td>
<td>++</td>
<td>CNS disease</td>
<td>10-8</td>
<td>11-3</td>
<td>4-2</td>
<td>0-3</td>
<td>Nil</td>
</tr>
<tr>
<td>4</td>
<td>Girl</td>
<td>10 months</td>
<td>Axillae Cervical</td>
<td>Nil</td>
<td>+ +</td>
<td>Nil</td>
<td>Nil</td>
<td>Pseudomonas infection</td>
<td>8-1</td>
<td>35</td>
<td>5</td>
<td>Nil</td>
<td>Nil</td>
</tr>
</tbody>
</table>

*Histological pattern identical to that of Burkitt's lymphoma.
++ = moderate; +++ = gross.

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**Table 2** Presenting lymphocyte surface marker studies

<table>
<thead>
<tr>
<th>Patient no</th>
<th>ER (%)</th>
<th>SM1g (%)</th>
<th>B1 (%)</th>
<th>I2 (%)</th>
</tr>
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<tbody>
<tr>
<td>1</td>
<td>41</td>
<td>25</td>
<td>NT</td>
<td>NT</td>
</tr>
<tr>
<td>2</td>
<td>3</td>
<td>80</td>
<td>83</td>
<td>NT</td>
</tr>
<tr>
<td>3</td>
<td>55</td>
<td>50</td>
<td>55</td>
<td>80</td>
</tr>
<tr>
<td>4</td>
<td>2</td>
<td>98</td>
<td>NT</td>
<td>NT</td>
</tr>
</tbody>
</table>

ER = erythrocyte rosettes.
SM1g = surface immunoglobulin.
NT = not tested.

**Table 3** Extended lymphocyte surface marker studies* on lymphoblasts of patient 4

<table>
<thead>
<tr>
<th>T cell markers</th>
<th>B cell markers</th>
<th>Early lymphoid markers</th>
</tr>
</thead>
<tbody>
<tr>
<td>Sheep rosettes</td>
<td>Anti IgG 70%</td>
<td>Anti-common ALL 0%</td>
</tr>
<tr>
<td>Anti-T 0%</td>
<td>Anti κ 74%</td>
<td>Tdt -ve</td>
</tr>
<tr>
<td>Anti κ 0%</td>
<td>HLA-DR 56%</td>
<td></td>
</tr>
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</table>

*From Dr Greaves, The Imperial Cancer Research Institute, London.
Figs. 1-4  The morphological appearances of the blast cells from patients 1-4.
In view of the atypical morphology in patient 4 and the finding of surface immunoglobulin on 98% of cells, the marker studies were repeated and extended by Dr M Greaves at the Imperial Cancer Research Institute. These results confirmed the monoclonal B cell phenotype of the blast cells (Table 3).

**Discussion**

Although the lymphoblast morphology is often of L3 FAB type in adults with B ALL, heterogeneous morphology with cases of L1 and L2 blast cells has been reported. Cases with atypical morphology have included patients with acute leukaemic transformation to B ALL from chronic lymphocyte leukaemia and secondary marrow involvement with non-Hodgkin's lymphomas.

In children, B ALL represents only 1–2% of cases of acute lymphoblastic leukaemia. Patients with L3 morphology have been reported consistently as having B ALL. Lymphoblasts of L3 type are not, however, diagnostic of B ALL in children. Between the years 1976 and 1983 we have seen five cases with L3 lymphoblasts that have typed as non-B, non-T acute lymphoblastic leukaemia, and similar cases have been reported by others. The marrow cytology and trephine biopsy histology of L3 cells is indistinguishable from those described in cases of Burkitt's lymphoma. In contrast to the African cases of Burkitt’s lymphoma, cases from non-endemic Europe and North America often have marrow involvement. The L3 morphology of marrow lymphoblasts seen in patients 1 and 2 was accompanied by histological features indistinguishable from Burkitt’s lymphoma on lymph node and ovarian tissue. The morphological and histological features of these two cases are representative of the majority of children with B ALL.

In contrast, the lymphoblast morphology in patient 4 has not been reported previously in association with B cell phenotype. The morphology of these cells is more characteristic of acute monocytic leukaemia. Two previously reported cases of acute monocytic leukaemia in infancy, however, had characteristic intracytoplasmic esterase activity present. In both these cases surface immunoglobulin was detected, but unlike our patient (see below) κ and λ light chains were found. This observation is consistent with the passive absorption of immunoglobulin from serum, rather than being the product of a neoplastic clone. In case 4 only κ light chains were shown, which confirms the monoclonal B cell phenotype of the leukaemia.

Surface immunoglobulin was shown in all four patients, with cells from patients 1 and 3 initially showing a population of positive T cell markers. Serial analysis of markers on marrow aspirates from patient 3 before chemotherapy showed a considerable reduction in OKT11 positive cells from 26% to 11% and a further increase in all B cell markers after 24 h. Analysable chromosomal spreads were unfortunately obtained from case 1 only and in this patient’s lymphoblasts the 14q+ anomaly previously reported with B ALL and Burkitt’s lymphoma was found. Further differentiation of cases of B ALL in children based on blast cell morphology may be of clinical and therapeutic importance. Recognition of cases with and without L3 morphology may be important. A low or absent blast cell count in the blood is usual in B ALL (the so-called Burkitt’s lymphoma cases with marrow disease). Cases 1 and 2 with L3 cells had low blast cell counts in the blood while blast cells were not seen in the blood from patient 3. In contrast, patient 4, with morphological features more like a monocytic leukaemia, had a high blood blast cell count (104 × 109/l).

Furthermore, patients with L3 morphology are at risk of severe metabolic complications secondary to acute tumour lysis, particularly during induction chemotherapy. Such features were noted in our first three patients, but patient 4, despite the initial high white cell count, encountered no such problems. This 10 month old baby girl responded well to induction chemotherapy with prednisolone and vincristine and remains in complete remission 18 months later on a UKALL VIII maintenance regimen.

In conclusion, we emphasise the importance of accurate identification of the lymphoblast phenotype in all cases of childhood leukaemia. Further cases with B cell phenotype and atypical morphological features may subsequently be recognised. The clinical and prognostic features of these cases may be different from previously reported cases of B ALL in childhood.

We thank Dr Mann for permission to include data on her patients in this study, Dr MG Greaves for extending the marker studies in patient 4, Dr AH Cameron for histopathology data, and Mrs P Mann for typing the manuscript.

**References**


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