Letters to the Editor

Clinical versus laboratory strains of Group B streptococci and human neutrophil chemiluminescence

Group B streptococci are implicated in serious infection, particularly during the neonatal period. Considerable attention has therefore been focused on the opsonisation and phagocytosis of these organisms by neutrophils in vitro since these are the principal means of host defence against group B streptococci. Neutrophil chemiluminescence, which provides an indirect measurement of membrane generated oxidative activity, is a sensitive technique for investigating opsonic requirements and phagocytosis of group B streptococci.1 Furthermore, a positive correlation has been shown between chemiluminescence and phagocytosis for a limited number of strains of serotypes Ic, II, and III,2 and in a separate report, serotype III.3

In a recent study we showed a significant, positive correlation between peak chemiluminescence values and the rate of phagocytosis of the five serotypes of group B streptococci, using the standard NCTC reference strains.4 Differences in opsonic requirements between strains of the same serological type have also recently been reported.5 The purpose of this study was to compare clinical isolates of group B streptococci with the NCTC reference strain, in terms of their ability to induce neutrophil luminol dependent chemiluminescence in an established assay system. The details of the experimental procedure have been described previously,6 and since type Ia (NCTC strain 090R) and type III (NCTC 11080) had consistently produced the lowest and highest chemiluminescence values respectively, eight clinical strains of type Ia and 10 of type III were tested.

The results are shown in the Table, with the peak chemiluminescence values, originally measured as millivoltage, expressed as a percentage of the maximum value (100%) produced within each serotype. All the clinical isolates of type Ia generated chemiluminescence far in excess of that for the NCTC strain, which always stimulated low chemiluminescence levels in our test system.4,4 Three of the strains were clearly very reactive, with the remaining five variably so. The range for the type III clinical strains was similar, but in this case the NCTC strain value was around the midpoint. To permit a comparison between the two serotypes, the 100% value for the Ia was 950 mV and that for the III was 1110 mV.

In commenting on these results several points should be made. In all previous experiments we have shown a clear separation in the chemiluminescence induced by NCTC strains of group B streptococci serotypes Ia and III,4 which was subsequently correlated with their phagocytosis.4 For the clinical strains tested here this distinction was no longer apparent, and the overlap between values for the two serotypes was considerable. In terms of relating such results to the likely pathogenesis of group B streptococci infections, this clearly casts doubt on the relevance of using laboratory strains for such in vitro experiments. We have not attempted to correlate chemiluminescence with phagocytosis of clinical strains as yet, although in one study the relation has been shown to hold for four clinical isolates of type III.1 In our first report we suggested that a failure to induce neutrophil chemiluminescence, and therefore by inference oxygen dependent microbicidal mechanisms, may be indicative of a bacterial strain which is pathogenic relative to one which stimulates higher levels of chemiluminescence, and thus a more successful host response.* This hypothesis remains to be tested by further investigation and comparison of the laboratory findings with clinical data.

M J KOWOLIK
CG CUMMING
Department of Oral Medicine and Oral Pathology,
University of Edinburgh,
High School Yards,
Edinburgh EH1 1NR

References

Inhibition of growth of Staphylococcus aureus on sensitivity testing agar by Streptococcus faecalis var zymogenes

In this laboratory the blotting paper strip method1 is used to obtain a semiquantitative estimate of the number of bacteria in urine. The method is both economical and rapid but can provide problems in isolating individual colonies for antibiotic sensitivity testing.

A catheter specimen of urine received in this laboratory was cultured by the blotting paper strip method and yielded numbers of colonies indicative of appreciable bacteriuria on both blood agar and Cled medium (Oxoid). Growth on both media was semiconfluent but in each case appeared homogenous.

Antibiotic disc sensitivity tests were carried out on DST agar (Difco), and after incubation for 18 h at 37°C sensitivity to the antibiotics was determined by the comparative method.2 The appearance of the sensitivity plate obtained is shown in the Fig.

The pattern was produced by the use of a mixed culture on the sensitivity plates. The opaque pigmented colonies, present only as discrete bands around some antibiotic sensitivity discs, was confirmed to be...