

Dihydropteridine reductase activity in dried blood spots: effects of aging and senile dementia of the Alzheimer type

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SUMMARY Dihydropteridine reductase (EC 1.6.99.7) (DHPR) activity was measured in blood spots from 50 neonates, 52 healthy adults aged 30–62 years, and 21 elderly controls aged 67–97 years, as well as 32 demented patients of whom 25 had senile dementia of the Alzheimer type. Enzyme activity was stable for seven days at 4°C and for at least 14 days at –20°C. No important difference was found between the DHPR activity of venous and capillary blood. DHPR activity was considerably lower in the healthy adult group compared with neonates and the elderly group, and there was no sex difference at any age. The erythrocyte DHPR activity of patients with senile dementia of the Alzheimer type was similar to that of elderly controls. This result differs from that previously reported for leucocytes.

It has been estimated that by the end of the century some 60 million people will suffer from senile dementia of the Alzheimer type.¹ Many aetiological hypotheses have been suggested—for example, neurotoxic factors,^{2,3} a virus,⁴ autoimmune disease,⁵ and genetic factors.⁶

The study of neurochemistry senile dementia in the Alzheimer type seems to be a more hopeful approach with regard to future treatment. Reduced concentrations of choline acetyltransferase activity,^{7–10} noradrenaline,^{11,12} dopamine,¹³ dopamine-B-hydroxylase activity,¹⁴ homovanillic acid,¹⁵ 5-hydroxytryptamine, 5-hydroxy indole acetic acid,¹¹ and somatostatin⁹ have all been reported. The neurochemical deficiencies seem to be greater than can be accounted for simply by cell death caused by dementia.¹⁶

Tetrahydrobiopterin (BH₄) is the essential cofactor for the formation of dopamine, noradrenaline, adrenaline, and 5-hydroxytryptophan from phenylalanine, tyrosine, and tryptophan, respectively.^{17–20} Interest in the neurological role of biopterins was first stimulated by the description of three children with a variant form of phenylketonuria. Despite early diagnosis and dietetic treatment they developed severe neu-

rological dysfunction. Biochemical lesions, later established as causative in this condition, include dihydropteridine reductase (DHPR) (EC 1.6.99.7) deficiency and defective synthesis of BH₄ with normal DHPR activity.^{21–23} The resulting deficiencies of neurotransmitters respond to replacement treatment both with neurotransmitters and intravenous 5,6,7,8 tetrahydrobiopterin²⁴.

Disturbed BH₄ metabolism has been described in several neurological diseases including senile dementia of the Alzheimer type, in which decreased biopterin concentrations have been reported in serum,^{25,26} plasma,²⁷ and cerebrospinal fluid.^{28,29} In addition, decreased synthesis of BH₄ has been shown in the temporal lobe samples removed from patients with senile dementia of the Alzheimer type at necropsy.³⁰ Neopterin (a precursor of BH₄) concentrations in dementia are no different from those in healthy control subjects, and this would support the concept that deficiency is not merely due to cell death.

The reason for reduced BH₄ in senile dementia of the Alzheimer type and other neurological diseases is as yet unknown. In samples from temporal lobe brain DHPR activity of patients dying with senile dementia of the Alzheimer type was not greatly different from that of controls.³⁰ Considerably lowered values of DHPR have, however, been shown in the leucocytes

of patients with senile dementia of the Alzheimer type compared with those in healthy elderly controls.²⁷ The success of BH₄ treatment in children with variant phenylketonuria and thus the possibility of similar treatment of patients with senile dementia of the Alzheimer type indicates the need for further investigation of biopterin metabolism in dementia. The objective of this study was to determine the effect of age on DHPR reductase activity in erythrocytes and whether the values were changed in senile dementia of the Alzheimer type.

The project was approved by the Joint Ethical Sub Committee of the Faculty of Medicine of the University of Southampton and Southampton and South West Hampshire Health Authority.

Material and methods

SUBJECTS AND CONTROLS

Group 1 neonates Capillary blood samples collected on to Guthrie cards for routine screening from 50 neonates between the sixth and 11th day of life were assayed for DHPR activity. No neonate had hyperphenylalaninaemia.

Group 2 healthy adults Venous blood samples (1 ml) were collected from 26 men and 26 women aged 30 to 62 years. With the exception of two women taking oral contraceptives, no volunteer was receiving medication.

Group 3 elderly adults Venous blood was collected from 31 patients aged 67 to 97 years attending a day hospital. This group, comprising 21 women and 10 men with scores of 25 or more on mental test assessment,³¹ was used as a control for comparison with the group with dementia. These patients had various conditions and were almost all receiving multidrug treatment.

Group 4 demented patients Venous blood was obtained from 32 demented patients aged between 69 and 87 years. They were divided into subgroups according to the clinical diagnosis and the Hachinski score, which acts as an indicator of multi-infarct dementia.³² Four had computed tomography brain scans to aid difficult diagnosis. Patients with secondary causes of dementia were excluded. All of them were receiving multidrug treatment.

6-methyl-5,6,7,8-tetrahydrobiopterin, ferricytochrome C type III from horse heart, and nicotinamide adenine dinucleotide (reduced form) were obtained from Sigma. All other chemicals were of Analar grade and obtained from BDH. Filter paper (grade 31 ET CHROMA) was obtained from Whatman Chemical Separation.

A Pye Unicam SP1800 double beam spectrophotometer set at a wavelength of 550 nm and a Pye

Unicam AR 25 linear recorder were used to monitor the change in absorbance at 37°C.

PROCEDURES

The activity of DHPR was determined by the method of Arai *et al.*,³³ modified to increase sensitivity by doubling the concentration of ferricytochrome C and increasing the temperature of the reaction to 37°C. Using the modified assay conditions, it was necessary to confirm that the erythrocyte DHPR activity was still proportional to the concentration of haemoglobin up to 0.5 mg/ml. Venous blood was diluted with distilled water to give a haemoglobin concentration range of 0.05–1.6 mg/cuvette. The DHPR activity at each haemoglobin concentration was measured. The diluted samples were then centrifuged at 2000 rpm for 10 minutes, and the DHPR activity for each sample was assayed again using the supernatants.

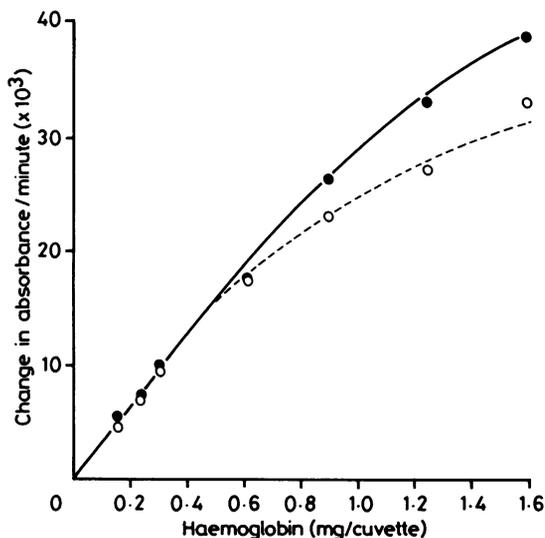
A DHPR assay for use with blood spot samples would provide the most effective method for screening of the elderly. For this reason and also to standardise the procedure for the neonatal and adult samples all the venous blood samples were spotted on to filter paper before analysis. It was therefore necessary to compare enzyme activity in capillary blood with that of venous blood. Both types of sample were collected from each of five healthy volunteers, and DHPR activity was measured in all the samples.

In addition, the whole blood samples and capillary blood spots were stored at 4°C and –20°C and subsequently analysed to determine the stability of the enzyme. Intrabatch precision was determined by analysing 18 replicate samples and interbatch precision was calculated from six analyses for three subjects.

Results

The Figure shows that at haemoglobin concentrations below 0.6 mg/cuvette centrifugation of the haemolysate was not essential. At higher haemoglobin concentrations the relation with DHPR activity was non-linear. Failure to remove red cell membranes led to an apparent further reduction in DHPR activity. As this was a potential source of error, care was taken to ensure that each haemolysate was centrifuged and that the haemoglobin concentration was below 0.6 mg/assay.

Using a paired *t* test no significant differences were found between DHPR activity in capillary or venous blood samples. Whole blood samples and blood spots were stable for seven days at 4°C and for at least two weeks at –20°C. The results for intrabatch precision were mean = 3.68 ((SD) 0.22) with a coefficient of



Effect of haemoglobin concentration on erythrocyte DHPR activity

variation of 6.0%. The coefficient of variation for the interbatch precision, however, varied from 8.6% to 10.2%.

The Table gives the results of enzyme activity in the four groups. There was no sex difference in any group. Enzyme activity was higher in groups 1 and 3 when compared with that of group 2. On further subdivision of group 2 by age (Table), using non-parametric analysis, the results of those aged 40 to 49 years were higher than for those aged 30 to 39 years and significantly higher than for those aged 50 to 62 years ($p < 0.05$). Group 3 was also divided into subgroups by age, but no difference in enzyme activity was observed, nor was enzyme activity associated with the mental test score. Patients with senile dementia of the Alzheimer type had the same enzyme activ-

ity as those in group 3. The seven patients with other types of dementia seemed to have similar values.

Discussion

The method used for the DHPR assay was simple, sensitive, and precise. The observed adult control range of DHPR activity was greater than that reported by Arai *et al.*,³³ and this may be due to the higher temperature used in the analysis.

There was no direct correlation of enzyme activity with age. Patients in group 2 showed lower activity than those in groups 1 and 3. It is unlikely that the increased values in group 3 (compared with group 2) were due to a simple age effect. The probable causes include multipathology and the effects of drugs. The commonest types of drug taken by the elderly patients in this study were tricyclic antidepressants, benzodiazepines, non-steroidal anti-inflammatory analgesics, diuretics, and cardiac glycosides. Tricyclic antidepressants and phenothiazines produce no effect on lymphocyte DHPR activity *in vivo*.²⁷ *In vitro* studies have concluded that tricyclic antidepressants, diazepam, chlorpromazine, trimethoprim, and sulphamethoxazole have no effect on DHPR and that methotrexate, lithium, folic acid, and ferrous sulphate inhibit the enzyme.³⁴ The effects of multipathology, however, together with combined drug treatment on DHPR activity have yet to be defined.

Interestingly, when group 2 was subdivided, those aged 40–49 years seemed to have higher values. There were more women in this subgroup, and perhaps the menopause affects erythrocyte DHPR activity.

There was no reduction in erythrocyte DHPR activity in patients with senile dementia of the Alzheimer type when compared with non-demented patients of similar age. This contrasts with the decreased DHPR in lymphocytes of patients with senile dementia of the Alzheimer type reported pre-

Erythrocyte DHPR activities (mean (SD))

Enzyme activity (nmol cytochrome C reduced/minute/mg Hb)

Group	Female	Male	Total
1 6–11 days	4.81 (1.32) (n = 24)	4.84 (0.94) (n = 26)	4.82 (1.13)* (n = 50)
2 Total group	4.11 (0.75) (n = 26)	3.76 (0.74) (n = 26)	3.93 (0.76)*† (n = 52)
30–39 years	3.96 (0.92) (n = 10)	3.75 (0.77) (n = 11)	3.85 (0.83) (n = 21)
40–49 years	4.42 (0.57) (n = 11)	4.06 (0.84) (n = 4)	4.32 (0.64)‡ (n = 15)
50–62 years	3.72 (0.58) (n = 5)	3.66 (0.70) (n = 11)	3.68 (0.65)‡ (n = 16)
3 67–95 years	4.41 (1.23) (n = 21)	4.60 (0.74) (n = 10)	4.47 (1.09)† (n = 31)
4 Senile dementia of the Alzheimer type	4.74 (1.50) (n = 14)	4.28 (1.02) (n = 11)	4.54 (1.31) (n = 25)
Multi-infarct dementia	3.97 (n = 1)	3.88 (1.39) (n = 3)	3.90 (1.14) (n = 4)
"Mixed type dementia"	4.31 (1.08) (n = 3)		4.31 (1.08) (n = 3)

* $p < 0.05$ group 2 compared with group 1;

† $p < 0.05$ group 2 compared with group 3;

‡ $p < 0.05$ age group 40–49 years compared with age group 50–62 years.

viously.²⁷ The methods used in both studies were similar. The DHPR assay on dried blood spots, however, is more precise than that for lymphocytes, with an interassay coefficient of variation of 9-10%, compared with 24% for the lymphocyte assay. It is possible that neither lymphocyte nor erythrocyte DHPR values reflect the enzyme's activity within the brain, but evidence suggests that this is not the case. Analysis of DHPR in leucocytes, erythrocytes, liver, brain, and skin from patients clinically diagnosed as having malignant hyperphenylalaninaemia, all showed a total enzyme deficiency.^{22 33 35} In addition, DHPR in human liver, leucocytes, erythrocytes, and fibroblasts has similar immunochemical characteristics.^{33 35} An alternative explanation, therefore, should be sought. Other clinical conditions and the medication of the elderly patients may have had important influences on DHPR activity and could account for the differing study results. A recent report showed that the neurotransmitter deficit in younger patients with senile dementia of the Alzheimer type dying in their seventh and eighth decades was more widespread and severe than that of the older patients with this condition.³⁶ Separating the data from this study—those under 80 years of age and those who were older—showed no difference in DHPR activity between the groups. It is impossible to draw conclusions from the other groups with dementia we studied due to the small number of samples analysed. All the results seemed to fall within the observed range for the elderly control patients, suggesting that there is no obvious DHPR deficit in multi-infarct and mixed type dementia.

It has been suggested that there is a defect in the synthesis of BH₄ in senile dementia of the Alzheimer type.^{26 27} Brains from subjects with senile dementia of the Alzheimer type have lowered total bipterin concentrations but retain their ability to synthesise neopterin and have normal DHPR activity.³⁰ This indicates a specific loss of ability to convert dihydro-neopterin triphosphate to BH₄ during de novo biosynthesis. The normal erythrocyte DHPR activity shown in the group with senile dementia of the Alzheimer type in this study supports these findings, and, as suggested previously,²⁷ indicates a possible therapeutic role for BH₄ in this disease.

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