Isoelectric points of urinary light chains in myelomatosi s: analysis in relation to nephrotoxicity

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SUMMARY Measurements of the isoelectric point (pI), sialic acid content, and polymerisation were made of 43 light chains isolated from the urine of patients with myelomatosis excreting large amounts of light chains. The pIs ranged from 3.5-9.5, and 23 were > 7.0. Sialylation was detected in 62% of light chains, and 69% showed microheterogeneity of charge. There was no clear association between the pI, sialylation, or polymerisation and the presence of renal failure, as assessed by serum creatinine concentrations. Light chains with pI spectrotopy in the basic range, however, were found to produce more proximal tubular dysfunction than acidic light chains.

Free light chains in the plasma are removed from the circulation by filtration through the glomerulus and catabolised in the proximal tubular cells; their clearance and catabolism is similar to several other serum low molecular weight proteins. In multiple myeloma the concentration of free light chains cleared through the glomeruli often exceeds the tubular reabsorption threshold so that they appear in abnormal concentrations in the urine (Bence-Jones proteinuria). The presence of light chains in the urine is accompanied by an impairment of proximal tubular reabsorption of other low molecular weight proteins and this is accentuated as the glomerular filtration rate falls when indicated by a rising concentration of serum creatinine. Some patients show severe renal impairment with a relatively low urinary free light chain concentration. Conversely, a few patients excrete very large amounts of free light chain with little or no impairment of glomerular function.

It has been suggested that the isoelectric point (pI) of light chains may be an important factor for determining the nephrotoxic potential of these proteins, although studies of the cause of nephropathy, using a light chain producing tumour transplanted into rats, have cast doubt on this hypothesis. Our preliminary studies of high performance chromatography of Bence-Jones proteins indicated that individual monoclonal proteins could be heterogeneous with respect to their charge, and this has recently been confirmed by isoelectric focusing with immunofixation.

In this study we identified various physicochemical characteristics of urinary free light chains in patients with myelomatosis who were excreting large amounts of light chain at the time of presentation. The objective was to examine whether there is a correlation between the character of the light chains in the urine and impaired renal function.

Patients and methods

Forty three untreated patients with myelomatosis whose excretion of urinary light chains exceeded > 3 U/l (IU = lg polyclonal light chains standard; this standard is known to underestimate or overestimate the weight of light chains in certain monoclonal light chains) were selected for study from the Medical Research Council’s Vth Myeloma Trial. They were divided into the functional groups based on those used in an earlier study; (A) creatinine < 150 µmol/l (1.69 mg/100 ml) at presentation and remaining below this level; (B) creatinine > 150 µmol/l at presentation falling to < 150 µmol/l on treatment; (C) creatinine > 150 µmol/l improving on treatment but remaining > 150 µmol/l; and (D) creatinine > 150 µmol/l rising or remaining unchanged despite treatment. In addition, patients who died within 100 days of presentation were analysed separately and excluded from groups A–D.

The relevant data about the urinary light chain type and concentration, serum paraprotein concentration, and serum creatinine concentration were obtained using the methods previously described. The urinary α2-microglobulin concentration was measured by radial immunodiffusion.

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ISOLATION OF LIGHT CHAINS

The urinary free light chains were isolated using a Pharmacia AB (Uppsala, Sweden) fast protein liquid chromatography (FPLC) system. Anion-exchanged chromatography was performed on a Mono Q HR5/5 column, and cation-exchange chromatography was performed on a Mono S HR5/5 column.

The Mono Q column was equilibrated with 6-25 mM bis Tris propane buffer, pH 7-5 (A) and the elution buffer (B) was 6-25 mM bis Tris propane buffer containing 0-35M sodium chloride (pH 9-5). A uniform elution gradient from 0% A to 100% B was produced over 20 ml. The Mono S column was equilibrated with 20 mM sodium succinate (pH 4-5), and the elution buffer was 20 mM sodium succinate containing 0-35M sodium chloride: the gradient was the same for the Mono Q. Both these columns were run at a flow rate of 1-0 ml/minute.

CHROMATOGRAPHIC SEQUENCE

The urine samples were desalted by chromatography on a Sephadex G25 column using the equilibration buffer for the Mono Q column. An aliquot was then chromatographed on the Mono Q column and the fractions collected. Most of the light chains were retained on the column, although some, usually the more basic types eluted in the void volume. Those that were not retained on the Mono Q column were rechromatographed on a Mono S after dialysis against the Mono S start buffer. The k and l light chains in chromatograms with complex peaks were identified by immunoprecipitation using specific antisera on Ouchterlony plates.

AFFINITY CHROMATOGRAPHY

Light chains that had complex patterns on the ion exchangers were also isolated by affinity chromatography using immobilised monoclonal anti-κ light chain antibodies linked to Sepharose 4B or monoclonal anti-l light chain antibodies, both affinity gels. Desalted urine proteins were applied to the column in 0-1M Tris (pH 8-0) containing 0-5M sodium chloride and 0-1% sodium azide and eluted with 3M potassium thiocyanate. Comparison of the FPLC and affinity methods of isolation showed that the same light chains gave similar pl spectrotypes with either method of isolation.

ASSESSMENT OF LIGHT CHAIN GLYCOSYLATION AND POLYMERISATION

The size of purified light chains was assessed by sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE) using 10% vertical slab gels prepared according to the method of Laemmlli.11 Samples were dissolved in Tris-SDS sample buffer and boiled for five minutes with or without 1% β2-mercaptoethanol. Low molecular weight standards (Pharmacia) were run with the samples. The gels were stained for protein with 0-1% PAGE blue 83 (British Drug Houses, Poole, Dorset) dissolved in methanol:acetic acid:water (3:1:6). The sialic acid content was measured by the thiobarbiturate method of Warren.12

MEASUREMENT OF URINARY LIGHT CHAIN PI SPECTROTYPE

Isoelectric focusing was performed in 1 mm thick 5% polyacrylamide gels containing 6-4% Pharmalyte 3-10. The gels were fixed in a mixture of 10% trichloroacetic acid and 5% sulphosalicyclic acid for one hour and washed overnight in destain (methanol: acetic acid:water 5:1:14). The protein bands were stained with PAGE blue 83, 0-1%, in destain. The isolectric points of the light chain bands were determined from the positions of the marker proteins (broad pl calibration kit, Pharmacia) was run in the same gel.

Results

Fig. 1 shows the spectrotypes of urinary light chain pl's. Sialic acid was detected in 26 of 42 (61-9%) samples. Fig. 1 shows the level of sialylation of the light chains and indicates that sialylation cannot be the only cause of the charge of heterogeneity.

Parallel studies of the same light chain isolated by an ion exchange chromatography and immuno-sorbent chromatography gave similar levels of sialylation and microheterogeneity of pl.

LIGHT CHAIN PI AND THE PRESENCE OF RENAL FAILURE

The results were ranked by serum creatinine, assessed before starting cytotoxic chemotherapy but after 48 hours of hydration. There was no obvious association between this measure of renal failure and light chain pl. This series contained three cases of IgDk, which is an exceptionally high incidence of this rare form of myelomatosis. These patients, however, were among the high light chain excretors from the 400 patients examined.

LIGHT CHAIN PI AND PROXIMAL TUBULAR DYSFUNCTION

It has been shown previously that patients with light chain proteinuria show objective evidence of selective proximal tubular dysfunction. This evidence included failure to reabsorb the low molecular weight protein α1-microglobulin [α1-m].4 There is a correlation between urinary α1-m and light chain excretion.10 Some patients, however, have less impairment of tubular reabsorption of α1-m than others with
Isoelectric points of urinary light chains in myelomatosis

Fig. 1 Data base of study. First column denotes immunoglobulin type A, G, or D of corresponding paraprotein; second column light chain type where κ or λ are shown alone it is a Bence-Jones proteinuria only myeloma. SA = sialic acid, LC = urinary light chain concentration, U = units (1 unit is equivalent to 1 g of polyclonal LC standard); α1-m = α1-microglobulin concentration.
equivalent levels of light chain production. To see if this heterogeneity was limited to the pl of the light chains, the mid point of the pl spectrotypes for each patient was plotted against pl-1-microglobulin-excretion. Figs. 2a and b show correlations for patients who were not in renal failure at presentation; there was an association between urinary pl-1-m concentrations and urinary light chain pl in these patients. No such association, however, was seen in the patients presenting with renal failure. Renal failure increases serum pl-1-m concentrations and hence the concentration of pl-1-m in glomerular filtrate. This complicates the analysis in patients in renal failure.

**Discussion**

Observations of the effects of injecting human myeloma light chains on glomerular function in rats have suggested that the light chains with a basic charge are nephrotoxic. Coward et al. reported a correlation between the light chain pl and glomerular filtration rate in a study of 23 patients excreting light chains in the urine (6–635 mg/mol creatinine). Light chains with a basic pl were associated with the greatest renal impairment; but Smolens et al. showed that in rates transplanted with a k-producing lymphoma, the pl of the urinary k chains, which varied from 4.3 to 7.6, did not correlate with the incidence of nephropathy.

Our study has provided evidence of the microheterogeneity of light chains, which might influence their physicochemical behaviour. Evidently, they can exhibit considerable microheterogeneity of charge. A spread of bands on isoelectric focusing of >0.5 pH units was present in 21 of 43 (48.8%) samples. Conversely, a pl restricted to a single band was seen only in eight of 43 (18.6%) samples. The microheterogeneity varied from two bands separated by a few tenths of a pH unit to four bands spread over 1.5 pH units. A comparable spread of pl and the presence of multiple bands has been reported by using immunofixation to identify the Bence-Jones proteins after isoelectric focusing. The density of sialic residues/mol of light chains suggests that they may suffer random loss after synthesis.

Our study does seem to indicate greater tubular
dysfunction in those patients whose light chains are more basic. This effect will tend to lower the level of light chain required to induce impairment of proximal tubular dysfunction. All patients in this study, however, had massive light chain proteinuria. This effect may be more pronounced and clinically important in patients with lower levels of light chain in the urine.

No association was seen between the degree of resolution of renal failure and light chain pI. This seems to indicate that pI is of relatively little importance in the induction of irreversible damage to nephrons in this disease. Some patients show progressive renal failure despite high fluid intake. In these patients the mechanism of renal failure seems likely to differ from that seen in most patients. None of the factors analysed in this study identified a physical characteristic that relates to this progressive form of light chain associated renal damage.

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References


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Table Distribution of light chain polymerisation

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<thead>
<tr>
<th>No renal failure</th>
<th>Renal failure</th>
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<tr>
<td></td>
<td>Death at 100 days</td>
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<tr>
<td>( \kappa )</td>
<td>3d, 2m, 1d + m</td>
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<tr>
<td>( \lambda )</td>
<td>7d, 1d + m</td>
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\( d \) = dimer; \( m \) = monomer; \( d + m \) = equal dimer and monomer.