Distribution of basement membrane antigens in clinical gastric adenocarcinomas: an immunohistochemical study

K NAKAMURA, M MORI, M ENJOJI

From the Second Department of Pathology, Faculty of Medicine, Kyushu University, Fukuoka, Japan

SUMMARY The distribution of laminin and collagen type IV in the basement membranes of 85 gastric adenocarcinomas was studied using immunoperoxidase techniques to check for invasive carcinoma. Lymph nodes with metastases were also studied in 23 cases. Thick and discontinuous staining of the basement membranes was observed in 12 cases of well differentiated adenocarcinoma; thin and discontinuous staining in 26 (12 well and 14 moderately differentiated adenocarcinomas); fragmentary staining in 36 (15 moderately and 21 poorly differentiated adenocarcinomas); and unrecognisable basement membrane staining in the remaining 11 cases of poorly differentiated adenocarcinoma. These patterns were largely related to the histological grade, the nuclear atypism and loss of polarity of tumour cells, and the degree of inflammatory infiltration.

Basement membranes are complex structures lining epithelia on connective tissue stroma. Distribution of these tissues differs in various states, particularly in neoplastic lesions. Laminin and collagen type IV are natural basement membrane components, and their distributions have been well studied in carcinomas of various organs. Recent reviews have shown that the changes in basement membrane distribution are related to the histological grade and metastatic property of tumours; this has prognostic implications. We analysed immunohistochemically the distributions of basement membranes in serial tissue sections, expressed by the reactivity to laminin and collagen type IV in human gastric adenocarcinomas.

Material and methods

Stomachs were resected from 85 patients (61 men, 24 women, age range 23–81 years) with a carcinoma, including 23 with spread to the regional lymph nodes. All but two patients were available for follow up data. Each specimen was fixed in buffered formalin, cut into slices, embedded in paraffin, and cut into three serial 5 μm thick sections. One section was stained with haematoxylin and eosin to confirm the histopathological diagnosis, and the other two were treated with 0.4% pepsin (Sigma P-7012) in 0.01 N hydrochloric acid for two hours at 37°C, washed and reacted with rabbit antibodies against mouse laminin (Bethesda Research Laboratories, Gaithersburg, Maryland, USA) and with mouse antibodies against human collagen type IV (Australian Monoclonal Development, Artarmon, Australia), respectively. Both antisera were used after diluting 1/50 in phosphate buffered saline at a pH of 7.3.

Bound antibody was detected by the avidin-biotin-peroxidase complex (ABC) method of Hsu et al after reaction with 0.2% hydrogen peroxide to remove endogenous peroxidase. Sections were treated with biotinylated antirabbit (for laminin staining) or antimouse (for collagen type IV staining), immunoglobulin antiserum (1/500), avidin (1/1000), and biotinylated horseradish peroxidase complex (Vector Laboratories, Burlingame, California, USA). The peroxidase was developed with 0.01% H2O2 and 0.05% 3,3-diaminobenzene tetrahydrochloride for five minutes. After a light counterstaining with methyl green the preparations were mounted using a solution of veronal buffered glycerol. Control stainings using pre-immune sera were always negative.

Immunohistochemical staining patterns were assessed according to the staining positivity, intensity, and continuity, and were classified as follows: (i) thick and discontinuous, (ii) thin and discontinuous, (iii) fragmentary, and (iv) unrecognisable. Tumour histol-
Basement membrane antigens and gastric adenocarcinoma

Oota and Sobin— that is, well differentiated, moderately differentiated, and poorly differentiated adenocarcinoma. Immunohistochemical and histological correlations were analysed contrasting the serial sections stained with haematoxylin and eosin with those stained for laminin and collagen type IV in each serial triplet.

Results

Similar patterns were obtained with antisera against laminin and collagen type IV using two contiguous sections. The basement membranes with intense stainings in the normal gastric mucosa showed a thick, continuous line beneath the epithelia. No essential differences were noted between the gastric mucosa and the intestinal metaplastic mucosa in the basement membranes. Vessel walls were also labelled, and prominent staining occurred when the epithelial basement membranes were closely apposed to the blood vessels.

In malignant tissue pericellular stainings of the basement membranes for laminin and collagen type IV showed different patterns in different parts of the adenocarcinoma. The typing of immunohistochemical staining was therefore based on the predominant appearance.

Thick and discontinuous basement membranes featured predominantly in 12 cases; basement membranes were as thick as those of normal epithelia but had occasional disruptions (fig 1); thin and discontinuous basement membranes were detected in 26 cases in which glandular structures of the tumour cells were surrounded by a thinner and occasionally disrupted line of the basement membranes (fig 2); fragmentary basement membranes, characterised by tiny fractions of the stainings adhering to the irregular nests of tumour cells (fig 3) or glandular structures were seen in 36. The remaining 11 showed tumour tissue without recognisable basement membranes.

The relation between the staining patterns of basement membranes and the degree of differentiation of gastric adenocarcinoma is shown in table 1. In well differentiated adenocarcinomas the thickness of the basement membranes diminished where pleomorphic nuclei, loss of polarity, or papillary structures predominated. Expression of tumoural basement membrane antigens tended to decrease in areas with severe infiltration of inflammatory cells (fig 4) as well as in less well differentiated tumours. In poorly differentiated adenocarcinoma basement membranes stained positive, though this was fragmentary, where the tumour tended to form solid nests or to be composed of gland-like structures. Examples of tumours without recognisable basement membranes were poorly differentiated adenocarcinomas in which tumour cells grew individually or in small clusters of loosely attached cells. Regardless of histological type, distinctive cells located at the periphery of the tumours generally lacked pericellular staining of basement membranes, although these cells rarely contained diffuse and granular intracellular staining of laminin and collagen type IV.

Lymphatic and blood vessel permeations by tumour cells were often observed in the tumours of this series. Clusters of tumour cells floating in the vessels usually lacked pericellular basement membrane staining. There were, however, six cases with intense, pericellular depositions of laminin in the intravascular clusters (fig 5). In these cases the primary tumours were well (n = 4) and moderately (n = 2) differentiated adenocarcinomas, falling in the thick or thin discontinuous basement membrane categories. Pseudoglandular collections of tumour cells floating in the blood vessels or lymphatics were surrounded by fragmentarily stained laminin. These features were not evident for collagen type IV. Metastasised lymph nodes were studied in 23 cases. The staining features of basement membrane antigens were essentially similar to those in their respective primary tumours.

Eighty three of the 85 patients were followed up for five years or longer (table 2). Of 11 with a tumour belonging to the unrecognisable basement membrane group, only three survived for five years or more, the five year survival being 27%. Conversely, in patients with a tumour in one of the other three categories, each five year survival exceeded 60%.

Discussion

Malignant tumour cells are essentially characterised by a loss of intact basement membranes. This has been shown by Barsky et al. who concluded that profound changes of basement membrane expression occur during the transition from a benign lesion to invasive carcinoma in the breast, skin, pancreas and prostate. Recent reviews by Forster et al showed that the presence of basement membranes containing laminin in rectal adenocarcinoma correlated with the
Fig 1  Thick and discontinuous basement membranes around malignant tubules, stained with antiserum for laminin. (Avidin-biotin-peroxidase methyl green.) Inset: low power view of the same area. (Haematoxylin and eosin.)

Fig 2  Thin and discontinuous basement membranes attached to malignant tubules of moderately differentiated adenocarcinoma, stained with antiserum for collagen type IV. (Avidin-biotin-peroxidase methyl green.) Inset: low power view of the same area. (Haematoxylin and eosin.)
Fig 3  Fragmentary basement membranes as staining fractions (white arrow) adjacent to nest of poorly differentiated adenocarcinoma, stained with antiserum for laminin. Vessel walls were intensely labelled (black arrows). (Avidin-biotin-peroxidase methyl green.) Inset: low power view of the same area. (Haematoxylin and eosin.)

Fig 4  Decreased basement membrane staining (arrows) around inflamed crypt of section of well differentiated adenocarcinoma, stained by antiserum for laminin. (Avidin-biotin-peroxidase methyl green.) Inset: low power view of the same area. (Haematoxylin and eosin.)
low histological grade (well differentiated tumours) and increased survival. These investigators did not differentiate between the different staining patterns of basement membrane: the staining was expressed only as positive or negative and the extent of basement membrane development varied with the different tumour. Our grading system for positive staining of basement membrane showed that deficient pericellular basement membranes are related largely to poor differentiation or anaplasia of the tumour.

The degree of histological differentiation in gastric carcinomas varies within the same tumour. Our observations show that the pattern of basement membrane staining also varied in the same way according to the histological degree of differentiation. Grigioni et al. reported that the basement membrane staining was related to the local architectural arrangement of the neoplastic cells. In our study the fragmentary basement membranes appeared in areas of the poorly differentiated adenocarcinoma where the tumour cells tended to adhere and the intracytoplasmic laminin and collagen type IV were visible in the invading anaplastic cells, arranged in small strands or cords. These observations suggest that the expression of basement membranes depends on the intensity of cellular attachment rather than on the degree to which the tumour cells have invaded the tissue.

Mechanism of basement membrane loss in invasive carcinoma remains unclear. It has been suggested that the loss is due to a decreased synthesis, decreased assembly, and increased turnover of basement membrane components by the tumour cells. Our results further showed that the pericellular basement membrane stainings were influenced not only by the tumour itself but also by stromal factors, particularly inflammation. Uitto et al. reported that leucocytes contain enzymes of degrading basement membrane collagen, which may account for the reduction of basement membrane in relation to inflammation. Infiltration of inflammatory cells into the stroma affects the distribution of basement membranes as well as the structure or shape of tumour cells. Acute or active inflammation of tissues of the gastrointestinal tract leads to morphological changes (of a regenerative or dysplastic nature) on the surface and pit lining epithelium. The in vivo and in vitro studies by Ingber et al. showed that anisotropic tumour cells of pancreatic acinar carcinoma, when in contact with basement membranes, had a reorganised location, form, and orientation, intracellular organelles, and a cell-cell contact. They presumed that acinar tumour cells possessed the intracellular machinery required to change the form needed for a normal differentiation of histology but as a result of neoplastic transformation had lost the trigger required for initiation of this structural cascade.

In this study intravascular tumour cells were rarely accompanied by pericellular basement membrane antigens for laminin and collagen type IV and, only rarely were thick or thin discontinuous basement membranes deposited pericellularly, shown by the

### Table 2: Relation between staining pattern of basement membranes and five year survival

<table>
<thead>
<tr>
<th>Staining Pattern</th>
<th>No</th>
<th>Five year survivals</th>
<th>Unknown</th>
<th>Five year survival %</th>
</tr>
</thead>
<tbody>
<tr>
<td>Recognisable:</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Thick and discontinuous</td>
<td>12</td>
<td>8</td>
<td>1</td>
<td>67%</td>
</tr>
<tr>
<td>Thin and discontinuous</td>
<td>26</td>
<td>17</td>
<td>1</td>
<td>68%</td>
</tr>
<tr>
<td>Fragmentary</td>
<td>36</td>
<td>21</td>
<td>1</td>
<td>60%</td>
</tr>
<tr>
<td>Unrecognisable</td>
<td>11</td>
<td>3</td>
<td></td>
<td>27%</td>
</tr>
</tbody>
</table>
staining patterns for laminin but not for collagen type IV. These findings suggest that both laminin and collagen type IV would have a role similar to that of basement membranes if the tumour cells were accompanied by anchorage stroma, and that laminin alone would sometimes appear where the epithelium-stroma interface was lost. Nevertheless, the possibility of technical artefact cannot be ruled out because of the relatively small number studied and of the few published.

The prognostic factors of gastric carcinoma were studied by Okada et al. who found that depth of penetration, invasion of the duodenum, growth pattern of the cancer, lymphatic or vascular invasion, and fibrosis around the cancerous area were clinically important factors. The most pertinent finding in our study was that tumours with unrecognisable basement membranes carried a significantly lower five year survival than tumours with recognisable basement membrane groups for which the five year survival exceeded 60%. Therefore, the tumours can be divided into two groups on this basis.

This study was presented in part at the 75th Annual Meeting of the Japanese Pathological Society on 10 April 1986, Sendai. We thank M Ohara for comments on the manuscript.

References

Requests for reprints to: Dr M Enjoji, Second Department of Pathology, Faculty of Medicine, Kyushu University 60, Maidashi 3-1-1, Higashi-ku, Fukuoka 812, Japan.