Immunoreactive determinants of CA 125 in women with endometriosis

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SUMMARY Among 10 patients with endometriosis CA 125 was increased (> 35 U/ml) in endometriotic cyst fluid in all the patients, but only two had increased serum concentrations. Gel electrophoresis of serum, endometriotic cyst fluid, and endometriotic tissue resolved the CA 125 immunoreactive fragments from the three sources into bands of similar electrophoretic mobilities. Electrophoresis under reducing and non-reducing conditions showed immunoreactive fragments of apparent masses of 55 000 and 140 000 daltons, respectively. Analysis under reducing conditions did not result in loss of activity. CA 125 antigen is thought to be a high molecular weight glycoprotein complex. As far as is known, this is the first report describing lower molecular weight immunoreactive determinants of CA 125.

CA 125 is an antigen recognised by the monoclonal antibody OC 125 obtained through somatic hybridisation of splenic cells of mice immunised with a cell line of OVCA 433 ovarian cancer.1 The antigen is expressed by most common epithelial ovarian carcinomas. In non-malignant tissue the antigen is frequently expressed in benign ovarian tumours of mucinous and non-mucinous origin.2-5 as well as in endometriosis.6-9 The presence of the antigen has also been shown in human milk,10 amniotic fluid,11 cervical mucus,12 central airway and lung tissue13 and seminal plasma.14 These findings suggest that CA 125 may be a secretory product of many normal human epithelia.

The CA 125 determinant has been reported to be associated with a mucin like high molecular weight glycoprotein complex,10,15 but several epitopes of the antigen have been identified,16-17 and the recognition of a different epitope by a monoclonal antibody raised against lung cancer cells16 suggests that different epitopes may be expressed in the various conditions with increased tissue expression of the antigen. After our previous report on the successful immunoscintigraphic localisation of areas of endometriosis18 this study was set up to characterise the epitope bound by OC 125 in endometriosis.

Material and methods

Samples of serum and endometriotic cyst fluid were obtained from 10 women undergoing oocyte retrieval for in vitro fertilisation. Fresh frozen tissue histologically confirmed as endometriosis was obtained from the Nuffield Department of Obstetrics and Gynaecology, Oxford, and a melanoma cell line was obtained from the Sir William Dunn School of Pathology, Oxford. The monoclonal antibody OC 125 was obtained from CIS (UK, Ltd).

The samples were analysed by sodium dodecyl sulphate polyacrylamide gel electrophoresis (SDS-PAGE) under reducing (with DTT and heating) and non-reducing (without DTT and without heating) conditions. In the first run 10 μl of sample fluid and 40 μl of solubilising buffer (2% SDS, TRIS-HCl 10mM, pH 8.0, 0-1 M DTT and 10% glycerol) was boiled for two minutes before running on SDS-PAGE. Tissues were finely minced, rinsed in distilled water, and extracted with 0-2% sodium deoxycholate in TRIS-HCl, 10 mM, pH 8-0, for five minutes at 20°C. Insoluble material was spun off in a microfuge at 9000 g for two minutes and 25 μl of the supernatant taken for analysis on SDS-PAGE. In the second (non-reducing) run the samples were applied in solubilising buffer not containing DTT and without boiling.

SDS-PAGE was performed as previously described19-20 in 4-12.5% gradient slab gels using the LKB vertical gel apparatus run at 4 V cm−1 for 16 hours. The molecular weight standards used were myosin (200 kilodaltons), b-galactosidase (116 kilodaltons), phosphorylase (94 kilodaltons), bovine albumin (67 kilodaltons), bovine albumin (67 kilodaltons) and ovalbumin (43 kilodaltons).
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Fig 1  Conventional SDS-PAGE (4-12.5% gradient) of endometriotic tissue (a), endometriotic cysts fluid (b, c) and serum (d) from patients with endometriosis under reducing (1a) and non-reducing conditions (1b). Approximate mobilities of molecular weight standards are indicated by arrows or arrow heads. The molecular weight standards used were myosin (200 kilodaltons), β-galactosidase (116 kilodaltons), phosphorylase (94 kilodaltons), bovine albumin (67 kilodaltons) and ovalbumin (43 kilodaltons). Other indicated mobilities are approximate.

The SDS-PAGE gels were subjected to electrophoretic transfer in TRIS-glycine buffer (25 mM and 192 mM, respectively), pH 8.3, containing 20% methanol, using the Transblot system (Biorad Laboratories, Richmond, California). Proteins and glycoproteins were transferred to nitrocellulose membrane filters (Schlecher and Schuell, GmbH, Dassel, West Germany), washed in PBS containing 0.4 M NaCl and 0.2% Tween 20 (PNT), and blocked in 1% bovine serum albumin in PNT for 30 minutes. After rinsing in distilled water the membrane was incubated in OC 125 at a 1 in 20 dilution in PNT for 30 minutes. After being washed three to four times in PNT buffer the membrane was incubated in 125I-labelled anti-mouse IgG for 30 minutes. Excess radioactivity was washed off with 0.1% w/v SDS in PBS. After staining with amido black (Sebia) the membrane was dried and exposed to x-ray film (Fuji Photo Film Company, Japan) at -70°C for 24 hours.

The serum samples and endometriotic cysts fluid were analysed for CA 125 content by an immunoradiometric assay according to the manufacturer’s instructions (CIS, UK).

Results

When the samples were subjected to SDS-PAGE under reducing conditions followed by immunoblotting, OC 125 was reactive with a band of about 55 kilodaltons in all the samples (fig 1a). Under non-reducing conditions and without heating, the immunoreactive fragments were about 140 kilodaltons. The endometriotic cyst fluid from case 9 (table)
Table  Serum and endometriotic cyst CA-125 in 10 patients with endometriosis U/ml

<table>
<thead>
<tr>
<th>Case No</th>
<th>Serum</th>
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<tbody>
<tr>
<td>1</td>
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<td>440</td>
</tr>
<tr>
<td>2</td>
<td>94</td>
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</tr>
<tr>
<td>3</td>
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<td>8</td>
<td>18</td>
<td>296</td>
</tr>
<tr>
<td>9</td>
<td>40</td>
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</tr>
<tr>
<td>10</td>
<td>12</td>
<td>&gt; 1000</td>
</tr>
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</table>

CA 125 antigen is defined by the monoclonal antibody OC 125 which identifies a conformational epitope to which both saccharides and protein contribute. CAMOV2, an oligosaccharide, is only occasionally expressed on the same molecule as CA 125.17 Various reports suggest that CA 125 is expressed independently of both the milk mucin family antigen21 and the CA 19-9 antigen.22

Studies on the antigenic nature of CA 125 show that it is a glycoprotein with a molecular weight in excess of 200 000.15 Davis et al also obtained similar data and suggested that the actual protein which expresses the antigenic determinant may be of a lower molecular weight.22 Our results show clearly that OC 125 binds to lower molecular weight determinants in endometriosis. Heating and analysis of the samples under reducing conditions did not inactivate the CA 125 antigen as previously suggested22 but produced lower molecular weight immunoreactive fragments. The CA 125 antigen seems to consist of, at least in part, protein determinants of 55 kilodaltons and 140 kilodaltons in endometriosis. The failure to obtain the high molecular weight immunoreactive species is difficult to explain, but the nature of the CA 125 determinant in endometriosis may be different. Our studies, however, did not attempt to purify and study homogeneous species and the physical properties of the determinants were not studied.

These data show that OC 125 binds to similar protein immunoreactive determinants present in the serum, endometriotic cyst fluid and endometriotic tissue in patients with the condition. It remains to be seen if the same determinants are present in patients with epithelial ovarian cancers and the other conditions with tissue expression of CA 125. The proteinaceous nature of the determinants also make it possible to institute studies which would determine their level of expression and the mode of genetic regulation in the many conditions with increased expression of the CA 125 antigen.

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References

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