Increased central immunoreactive β-endorphin content in patients with Wernicke-Korsakoff syndrome and in alcoholics

J A Summers, P T Pullan, J J Kril, C G Harper

Abstract

β-endorphin, adrenocorticotrophin, and α-melanocyte stimulating hormone were measured by radioimmunoassay in three areas of human brain at necropsy in seven subjects with Wernicke-Korsakoff syndrome and in 52 controls. Thiamin concentration in six brain areas was also measured. Mamillary body β-endorphin concentrations were significantly increased in those with the syndrome compared with controls, and those controls with high alcohol intake showed increased mamillary body β-endorphin compared with controls with low alcohol intake. Brain thiamin concentration was similar in both groups, with the exception of the brainstem, where it was reduced in subjects with Wernicke-Korsakoff syndrome. Thalamic β-endorphin in controls was inversely correlated with thiamin in frontal white matter, frontal cortex, parietal white matter and parietal cortex, while β-endorphin in the hypothalamus of patients was inversely correlated with thiamin in frontal cortex, parietal white matter, thalamus and brainstem. These results suggest that there is a disturbance of the endorphinergic system in Wernicke-Korsakoff syndrome which may be related to alcohol intake.

Ethanol may affect the metabolism of β-endorphin (β-EP) in the central nervous system (CNS). Ethanol ingestion for two to three weeks led to increased synthesis of β-EP, β-lipotropin (β-LPH), and their precursor molecule, pro-opiomelanocortin (POMC), in the pituitary gland of rats. Acute ethanol in vivo was associated with pituitary release of β-EP. Conversely, β-EP pre-conditioning attenuated the acquisition of ethanol preference, suggesting that the CNS activities of β-EP and ethanol may be interdependent.

Administration of β-EP has repeatedly been shown to affect memory and learning. Low dose β-EP produced complete amnesia for two types of task in rats, whereas naloxone, a specific endorphin antagonist, facilitated memory function. Similar findings have recently been reported in chicks, and continue to be shown in rats. The sum of experimental evidence in animals and man suggests that endogenous opioids have a major physiological role as amnesic agents.

Memory performance in rats is improved by adrenocorticotrophin (ACTH) and α-melanocyte stimulating hormone (α-MSH), which are also derived from POMC. Similar but less predictable effects are found in man, where the predominant changes are on mood and selective attention.

The brain areas which contain the highest concentrations of POMC and its products—that is, the hypothalamus, mamillary body, and thalamic nuclei—are precisely the same areas most affected in Wernicke-Korsakoff syndrome. This disease, caused by thiamin deficiency and found almost exclusively in alcoholics, is characterised by profound memory impairment. We have previously reported decreased hypothalamic ACTH concentrations in rats with long term thiamin deficiency. In view of the colocalisation of central POMC product distribution and Wernicke-Korsakoff lesions, the putative roles of β-EP, ACTH, and α-MSH in memory function; reported effects of ethanol on CNS β-EP metabolism; and the finding of central ACTH depletion in rat thiamin deficiency, this study was undertaken to measure POMC products in the human brain at necropsy and to relate these measurements to the presence of Wernicke-Korsakoff lesions, brain thiamin content, and alcohol intake during life.

Methods

Fifty nine cases were selected for study from a larger series of 120 necropsies. Selected cases had no neurological or neuropsychological abnormalities except those relating to alcohol. Necropsies were performed at the Perth coroner’s department and the Royal Perth Hospital. Brains were collected fresh and the weight and volume determined. The cerebellum and brainstem were removed at the level of the midbrain and the hemispheres divided sagittally. The left hemisphere was fixed for 14 days in 10% buffered saline after which it was embedded in agarose and cut at 3 mm intervals in the coronal plane. The cerebellum and brainstem (after the brainstem block was removed) were also fixed and sectioned. Macroscopic and microscopic examinations, including the cerebellum, midbrain,pons,medulla,hippocampus,thalamus, hypothalamus, mamillary body and numerous areas of the cortex, were performed.

The right hemisphere was dissected fresh and the areas for study placed in preweighed glass phials, reweighed, and frozen at −70°C. Samples of tissue were taken from: the mamillary body—entire right mamillary body, the frontal cortex—superior frontal gyrus,
Brain β-endorphin in alcoholics

frontal white matter—deep white matter underlying the superior frontal gyrus, parietal cortex—precentral gyrus, parietal white matter—underlying precentral gyrus, hypothalamus and thalamus—cut into the basal ganglia at the level of the mamillary body, and brainstem—cross sectional slice of medulla at the inferior olivary nuclei.

To determine the alcohol intake, cause of death, nutritional state and vitamin tablet consumption, clinical information was collected from hospital notes, necropsy reports, and questionnaires sent to the relatives of the deceased. No case was included which had any history or evidence of a neurological disease or a head injury. All cases of Wernicke’s encephalopathy were confirmed by examining the contralateral hemisphere histologically.

TISSUE EXTRACTION FOR NEUROPEPTIDE ANALYSIS

The extraction procedures for β-EP, ACTH, and α-MSH were identical and were based on the method of Krieger et al. Tissue samples were homogenised for 10 seconds in 600 µl 100 mM hydrochloric acid. After the addition of 600 µl 80% acetone in 100 mM hydrochloric acid the samples were homogenised for a further 10 seconds. After centrifugation (6000 x g for 10 minutes) 800 µl of supernatant was transferred to a clean polypropylene tube and dried under nitrogen in a water bath at 60°C. The dried extract was reconstituted in 100 µl 100 mM hydrochloric acid before the addition of 900 µl 50 mM sodium phosphate buffer, pH 7.4 (containing 0.25% human serum albumin (Commonwealth Serum Laboratories, Melbourne, Australia), and 0.5% 2-mercaptoethanol (Calbiochem, USA). The pH was checked with phenol red indicator, and if necessary, samples were neutralised with 100 mM NaOH. Average recoveries at 100 pg/assay tube were 107%, 90%, and 84% (n = 5) for β-EP, ACTH, and α-MSH, respectively.

RADIOIMMUNOASSAY

Brain extracts were assayed at appropriate dilutions in three radioimmunoassays based on the methods of Rees et al24 and Jeffcoat et al.25 Separation of bound and unbound ligand was achieved by second antibody precipitation rather than the charcoal used in the original methods. The β-EP antiserum (BSH), raised in rabbits, was used. The final incubation dilution of 1 in 20 000 and cross reacted equally on a weight to weight basis with β-LPH, to a slight degree with ACTH<sub>1-24</sub> (0.005%), α-endorphin (0.003%), and α-MSH (0.0003%), but not at all with ACTH<sub>1-39</sub>, ACTH<sub>1-24</sub>, ACTH<sub>18-39</sub> (CLIP) or met-enkephalin. The ACTH antiserum, raised in sheep and used at a final incubation dilution of 1 in 12 000, was directed at the middle portion of the ACTH molecule, and showed cross reactivity with ACTH<sub>1-24</sub> (100%), α-MSH (0.5%), and β-EP (0.004%). A commercial antibody used for the α-MSH assay (UCB Bioproducts, Belgium 1660/001, Batch 203) at a final dilution of 1 in 600 gave significant cross-reactivity only with ACTH<sub>1-24</sub> (0.15%) and CLIP (0.02%).

Resulting assay sensitivities were 1-0 pg/mg for β-EP and α-MSH in the hypothalamus, 0-4 pg/mg for β-EP in the mamillary body and thalamus, and 0-5 pg/mg for ACTH. All samples were assayed blind.

Measurement of thiamin was performed by an automated method using Lactobacillus fermenti<sup>26</sup> based on an earlier manual microbiological method.<sup>27</sup>

DATA ANALYSES

Results for brain thiamin concentration are expressed as mean ± standard error of the mean, and were compared using Student’s t-test for unpaired data. All other data, which were not normally distributed according to the Kolmogorov-Smirnov one sample test, were transformed using natural logarithms before comparison of geometric means using Student’s t-test. Results are expressed as geometric mean with standard deviation of the geometric mean. Log transformed data were used to calculate Pearson’s partial correlation coefficients, where the subject’s age, body mass index (BMI), and delay after necropsy were held constant.

The concentration of neuropeptides in some brain regions is extremely low in terms of present assay sensitivities; where undetectable results occurred, they were assigned a value corresponding to half the assay sensitivity, for the purposes of calculating mean values. In control subjects this was the case in six of the thalamic β-EP samples, in 19 mamillary body ACTH, and 24 thalamic ACTH samples, and virtually all of the thalamic α-MSH samples, which have been reported as “most undetectable”. Samples from subjects with Wernicke-Korsakoff syndrome were undetectable only twice. Where this occurred values were assigned and treated as described above. All undetectable results were omitted from calculations of correlation coefficients.

The influence of alcohol intake on mamillary body β-EP was studied by dividing control subjects into three categories, depending on whether daily alcohol intake in grams of pure ethanol was < 20 g/day (low), 20–80 g/day (moderate), or > 80 g/day (alcoholic). Five control subjects were excluded due to there being no information available regarding their alcohol intake. The remaining subjects were divided into the remaining six groups on the basis of the ratio of alcohol intake in grams of pure ethanol to body mass index (BMI) for the alcohol intakes of each subject. No difference was found between the BMI for groups of BMI less than 20 % and those with BMI greater than 20 %.

Results

Of the 59 subjects, seven had lesions diagnostic of Wernicke-Korsakoff syndrome, of which five were chronic and two acute on chronic. Mean age of the remaining subjects was 51.8 (2.3) years, with those with Wernicke-Korsakoff being 62.0 (6.0) years (NS). The mean BMI of both groups was in the normal weight range. The control group comprised 42 males
and 10 females; the group with Wernicke-Korsakoff syndrome was exclusively male. Mean necropsy delay was the same in both groups (controls: 34.5 (2.9) patients; 28.5 (2.2) hours).

Mamillary body β-EP was increased in subjects with Wernicke-Korsakoff syndrome by a factor of 2 (p < 0.01) (table 1). Mean mamillary body β-EP compared with alcohol intake is shown in the figure. There was a significant difference between the mean mamillary body β-EP values of the low intake group compared with the alcoholic group (p < 0.05). The results of the group with Wernicke-Korsakoff syndrome were higher compared with the low (p < 0.01) and moderate alcohol intake (p < 0.05) groups. There were no other significant changes to POMC products in subjects with Wernicke-Korsakoff syndrome compared with controls.

Mean thiamin concentrations in the brain regions tested were not significantly changed in subjects with Wernicke-Korsakoff syndrome, with the exception of the brainstem, where it was reduced (p < 0.02), (table 2). Thalamic β-EP was negatively correlated with thiamin in the frontal white matter (r = 0.31, p = 0.01), frontal cortex (r = 0.30, p = 0.02), parietal white matter (r = 0.38, p < 0.005) and parietal cortex (r = 0.25, p < 0.05) of control subjects. In patients, negative correlations between brain thiamin and β-EP concentrations were also seen in this case between β-EP in the hypothalamus and thiamin in frontal cortex (r = 0.71, p < 0.05), parietal white matter (r = 0.72, p < 0.05), parietal cortex (r = 0.67, p = 0.01), thalamus (r = –0.82, p < 0.01) and brain stem (r = –0.82, p < 0.02).

No further significant effects on brain POMC products, whether related to cause of death, vitamin tablet consumption, or any other measured variables were found.

**Discussion**

Although the highest concentrations of brain POMC products are contained in the arcuate nucleus of the hypothalamus, this study showed no differences in hypothalamic concentrations of the three measured peptides between control subjects and patients with Wernicke-Korsakoff syndrome in this region. This contrasts with our earlier findings in thiamin deficient rats, where hypothalamic ACTH concentration was half that of controls.

An earlier study of 51 cases of subjects with Wernicke-Korsakoff, however, showed that the mamillary body was the major locus of structural damage, being shrunken, brown, and spongy in appearance in all but one instance. Changes in the third ventricular wall and thalamus were seen less often. It is therefore of interest that mamillary body β-EP concentration was significantly affected in our study. That mamillary body β-EP seemed to increase the higher the dose of alcohol, is of further interest, and suggests that although the causative factor of Wernicke-Korsakoff is thiamin deficiency, alcohol, too, may have a large role in the changes in CNS metabolism which have such devastating effects on memory function.

An unexpected finding was that subjects with Wernicke-Korsakoff syndrome were well nourished, with normal BMIs, and that there was evidence of thiamin deficiency only in the brainstem. The value of thiamin estimation in subjects with Wernicke-Korsakoff syndrome is uncertain however, because it is routine emergency department practice to administer a large dose of thiamin to suspected cases of Wernicke-

### Table 1

Necropsy concentrations of β-endorphin, ACTH, and α-MSH in three midbrain areas of 52 controls (C) and seven subjects with Wernicke-Korsakoff syndrome.

<table>
<thead>
<tr>
<th></th>
<th>Hypothalamus</th>
<th>Mamillary body</th>
<th>Thalamus</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>β-EP</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Controls</td>
<td>99.48 (0.35)</td>
<td>8.95 (0.64)</td>
<td>8.67 (1.22)</td>
</tr>
<tr>
<td>Wernicke-Korsakoff</td>
<td>91.84 (0.33)</td>
<td>17.64 (0.65)*</td>
<td>8.17 (1.26)</td>
</tr>
<tr>
<td><strong>ACTH</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Controls</td>
<td>8.08 (0.74)</td>
<td>3.35 (1.32)</td>
<td>1.30 (0.50)</td>
</tr>
<tr>
<td>Wernicke-Korsakoff</td>
<td>11.70 (0.91)</td>
<td>2.34 (0.76)</td>
<td>1.22 (0.32)</td>
</tr>
<tr>
<td><strong>α-MSH</strong></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Controls</td>
<td>17.81 (0.40)</td>
<td>Not measured</td>
<td>Most</td>
</tr>
<tr>
<td>Wernicke-Korsakoff</td>
<td>17.29 (0.48)</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Results are expressed as μg/mg wet weight of brain tissue.

*p < 0.01.

Results are geometric mean with standard deviation of the geometric mean in parentheses.

### Table 2

Mean (SEM) thiamin concentrations in six brain areas of 52 controls and seven patients.

<table>
<thead>
<tr>
<th></th>
<th>Controls</th>
<th>Patients</th>
</tr>
</thead>
<tbody>
<tr>
<td>Frontal white matter</td>
<td>1.00 (0.06)</td>
<td>1.05 (0.17)</td>
</tr>
<tr>
<td>Frontal cortex</td>
<td>1.60 (0.08)</td>
<td>1.35 (0.19)</td>
</tr>
<tr>
<td>Parietal white matter</td>
<td>0.97 (0.06)</td>
<td>0.86 (0.13)</td>
</tr>
<tr>
<td>Parietal cortex</td>
<td>1.68 (0.08)</td>
<td>1.40 (0.18)</td>
</tr>
<tr>
<td>Thalamus</td>
<td>1.90 (0.10)</td>
<td>1.71 (0.19)*</td>
</tr>
<tr>
<td>Brainstem</td>
<td>1.24 (0.08)</td>
<td>0.94 (0.09)*</td>
</tr>
</tbody>
</table>

Results are expressed as μg/g wet weight of brain tissue.

*p < 0.02.
Korsakoff syndrome. Although none of them regularly took vitamin tablets, three of them did so occasionally. The observed negative correlations between brain thiamin and β-EP concentrations, although suggestive of an interplay between the two factors, should therefore be viewed with caution.

It should be noted that the β-endorphin assay used in this study does not distinguish between it and the related, but behaviourally inactive peptide β-LPH, nor was the methodology used here able to distinguish whether the observed increase in β-endorphin was due to increased synthesis or disturbed metabolism. In view of the lack of effect on ACTH and α-MSH, which derive from the same precursor, the latter seems the more likely mechanism.

The hypothesis that β-endorphin may have a role in the pathogenesis of Wernicke-Korsakoff syndrome, has been put forward in several publications, the authors proposing that an increase in β-endorphin concentrations in Wernicke-Korsakoff syndrome may account for some of the amnesic features of the syndrome. The only other published studies of central POMC products in alcoholics contradict this point of view, deficits of cerebrospinal fluid β-endorphin having been reported. The assumption in these studies is that CSF peptide concentrations reflect their concentration in the brain itself, whereas in reality the situation may be more complex. In view of the present findings one possible interpretation is that ethanol interferes with normal β-EP release and metabolism, causing its concentration in the mamillary body to rise, with a corresponding fall in cerebrospinal fluid concentrations.

Our results seem to point towards there being an aetiological role for the endorphinergic system in Wernicke-Korsakoff syndrome, and that the gradual rise in β-EP with increasing alcohol intake provides further evidence that alcohol may exert some of its reinforcing effects through endogenous opiates.

In summary, the preceding data lend support to the hypothesis that excessive β-EP activity is a contributory factor to the memory deficit in Korsakoff’s psychosis, and that this excess is related to high alcohol intake. This is the first time, as far as we are aware, that an abnormality of brain neuropeptide concentrations has been reported in this disorder.

We thank Professor J H Rees and Dr P J Lowry (now Professor Lowry, University of Reading), Department of Chemical Endocrinology, St Bartholomew’s Hospital, London, for their generous gift of antibodies to β-endorphin and ACTH, and Dr D Davis and Mr G Icke, Haematology Department, Royal Perth Hospital, Western Australia, for performing thiamin estimations. Mr J Daly assisted with the collection of specimens.


4 Sandi C, Borrelli J, Guzau C. Beta-endorphin administration interferes with the acquisition and initial maintenance of ethanol preference in the rat. Physiol Behav 1989;45:87–92.


6 Izquierdo I, Dias RD. Effect of ACTH, epinephrine, β-endorphin, naloxone and of the combination of naloxone or β-endorphin with ACTH or epinephrine on memory consolidation. Psychoneuroendocrinology 1983;8:91–7.


