Cells containing factor XIIIa and pulmonary fibrosis induced by bleomycin

M Toida, Y Okumura, T Takami

Abstract

To show the clinical importance of cells containing FXIIIa in pulmonary fibrosis induced by bleomycin, the distributions of FXIIIa and collagenous components were investigated immunohistochemically in both normal lung tissues and those affected by bleomycin. In the normal tissues FXIIIa-containing cells were sparse, but they were numerous in the pulmonary fibrotic tissues, especially in the subpleural area and around the blood vessels of alveolar septa, where slight to moderate fibrosis was seen, and in the intra-alveolar fibrinous exudate. In the collagenous scar-like areas, however, these cells were fewer in number and their FXIIIa expression was depleted. These findings suggest that cells containing FXIIIa have an important role in the development of pulmonary fibrosis induced by bleomycin.

Bleomycin is one of the most popular drugs for squamous cell carcinoma of the head and neck regions, but it can cause fatal pulmonary fibrosis. This lesion is distributed predominantly in the lower pulmonary lobes and in subpleural regions. It is non-specific histologically and similar to the lesions induced by many other agents affecting the lung. The histological features include intra-alveolar accumulation of proteinaceous for fibrinous exudate, proliferation of alveolar cells, interstitial and intra-alveolar fibrosis, and squamous metaplasia with epithelial dysplasia of the distal air spaces. FXIIIa has been detected immunohistochemically in certain interstitial cells of various human tissues, though the distribution of FXIIIa was not examined in pulmonary tissues. More recently, it has been suggested that these cells have an important role in the process of fibrosis in various lesions.

Methods

Fifty five specimens of lung tissue, consisting of 24 normal and 31 pulmonary fibrotic tissues were obtained from five necropsies. The tissues were almost intact in two of the cases (a 40 year old man and a 54 year old woman), and the tissues from the other three cases (a 61 year old woman, a 69 year old man, and a 79 year old man) showed pulmonary fibrosis. Specimens about 5 mm thick were cut from the excised lower and immediately fixed in glacial acetic acid 1% (v/v) in 95% ethanol at 4°C for 12 to 24 hours. The specimens were embedded in paraffin wax and cut into 4 μm thick sections. Each section was dewaxed, hydrated, and stained with haematoxylin and eosin and azocarmine and aniline blue (AZAN).

Collagenous components were stained using the Sirius red F3BA method and a collagen stain kit (Collagen Gijutsu Kenshu-Kai, Tokyo, Japan).

Immunoperoxidase staining for FXIIIa was performed by means of the avidin–biotin peroxidase complex (ABC) method with rabbit antiserum against human FXIIIa (Behringwerke AG, Marburg, Germany) diluted 1 in 400 and the rabbit IgG Vectastain ABC kit (Vector Laboratories, Burlingame, California, USA). The specific antigen–antibody reaction product was visualised by 3,3′-diaminobenzidine tetrahydrochloride and hydrogen peroxide. Counterstaining was performed with Mayer's haematoxylin. For control slides, normal rabbit serum at the same dilution was used instead of anti-FXIIIa antiserum.

Details on both Sirius red F3BA collagen staining and immunoperoxidase staining procedures have already been described.
Results
In normal tissues FXIIIa was detected in a few interstitial cells. They were mainly spheroidal or spindle-shaped. Alveolar macrophages also exhibited a weakly positive reaction to FXIIIa. Alveolar epithelium, pleural mesothelium, vascular endothelium and bronchial glandular cells were all negative for FXIIIa.

In fibrotic tissues numerous cell containing FXIIIa were observed, especially in the subpleural area (fig 1) and around the blood vessels of alveolar septa, where slight to moderate fibrosis was seen. These cells were mainly dendritic in shape, and some of them were dust-laden (fig 1). In the alveolar spaces, where proteinaceous or fibrinous exudation and intraalveolar fibrosis were prominent, the alveolar macrophages exhibited a strong positive reaction for FXIIIa (fig 2). In the highly collagenous scar-like areas the interstitial cells containing FXIIIa were fewer in number. They were mainly slender, spindle-shaped, and exhibited a weak positive reaction for FXIIIa.

Discussion
Recently, it has been shown that FXIIIa can catalyse in vitro not only the crosslinking of fibrin monomers to each other, but also the crosslinking of fibronectin monomers to each other and to type I and type III collagens. This may be important not only for the stabilisation of fibrin clots, but also during the process of fibrosis. Recent immunohistochemical studies have also indicated findings suggesting that the cells containing FXIIIa have an important role during the process of fibrosis in various lesions. Our study shows a close relation between the distribution of cells containing FXIIIa and that of collagenous components. It has been suggested that alveolar macrophages have an important role in the development of pulmonary fibrosis by releasing various fibroblast proliferation factors, such as fibronectin, certain types of platelet-derived growth factor and interleukin-1. Apart from these factors, FXIIIa contained in alveolar macrophages as well as in pulmonary interstitial cells may also have an important role in the development of this lesion. On the basis of the findings obtained in this study we speculate that these cells proliferate before fibrosis develops and that the FXIIIa stimulates fibrosis in the early stage of this lesion, and furthermore, that this role of the cells is diminished as the lesion develops to the final stage.

In this study some of the cells containing FXIIIa were identified morphologically as alveolar macrophages. The dust-laden connective tissue cells containing FXIIIa are also interpreted as being of the macrophage type. There is, however, some doubt as to whether all interstitial cells that contain FXIIIa belong to the macrophage type; and also whether macrophages can in fact act as facultative fibroblasts.

We thank Dr Akitsugu Ojima, the Department of Pathology, Gifu University School of Medicine, for his advice; Dr Nobumitsu Oka, and Norichika Tatematsu, of the Department of Oral and Maxillo-Facial Surgery, Gifu University School of Medicine; and Dr Matsuo Hosimoto, of Ihi General Hospital for their help.