Use of fine needle aspiration cytology for investigating lymphadenopathy in HIV positive patients

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Abstract
The cytological diagnoses of 27 lymph node aspirates were compared with the histological diagnoses or clinical outcome in 23 HIV positive patients. There was agreement between the cytological and histological diagnoses in 14 of the 16 surgically biopsied cases. The clinical outcome in the remaining 11 cases was consistent with the cytdiagnosis. Fine needle aspiration (FNA) is a reliable, minimally traumatic, cost effective method with high specificity. It is suitable for an initial rapid diagnosis in HIV positive patients with lymphadenopathy.

Methods
Twenty seven fine needle aspirates were obtained from 23 HIV positive patients who presented with lymphadenopathy.

The palpable lesions were aspirated in an outpatient setting, without anaesthetic. A 10 ml syringe and 23 gauge needle were used and several passes were made. The hand used to immobilise the lesion was protected by a special Kevlar glove (Du Pont, Geneva, Switzerland), which has needle resistant padding on the finger tips. The gloved hand was moved away before the needle was withdrawn from the lesion. No mask was worn, but special care was taken in expelling the aspirate immediately and very closely to the surface of the labelled glass slides to prevent the creation of an aerosol. The material was spread evenly with another slide. All slides were air dried and fixed with methanol. The needle was rinsed with saline and the content was expressed into a test tube for making further slides if required. The slides, needle washings, and request form were sent to the laboratory in a double pocketed plastic bag bearing a health hazard sticker. The stains routinely used in the laboratory were May-Grünwald Giemsa and Grocott’s. Immunocytochemical stains were performed when required. These included CAM 5-2 (Dako), LCA (CD45) Dako; L26 (Dako); UCHL1 (Dako); k and l (Dako). At the start of the study a Ziehl-Neelsen stain was done only when tuberculosis was suspected clinically or seen on light microscopic examination. Later specimens were routinely stained by the Ziehl-Neelsen method.

The histological material was routinely stained with haematoxylin and eosin Ziehl-Neelsen, and Grocott stains. Immunohistochemical stains were done when required.

All histological and cytological diagnoses were reviewed (by EMB and KS). The relevant clinical data were obtained from the request forms or, where available, from the clinical notes.

Results
Twenty seven lymph node aspirates were obtained from 23 HIV positive patients—22 male homosexuals and one female heterosexual of African origin. The average age at first clinical presentation was 36-4 (range 19 to 72) years. Four patients presented with lymphadenopathy on more than one occasion.

Twenty six aspirates were from palpable
Correlation between cytological and histological diagnoses and clinical outcome

<table>
<thead>
<tr>
<th>Cytological diagnosis</th>
<th>Confirmed on clinical follow up only</th>
<th>Histological confirmation</th>
<th>Other histological diagnoses</th>
</tr>
</thead>
<tbody>
<tr>
<td>Reactive changes</td>
<td>11</td>
<td>4</td>
<td>5</td>
</tr>
<tr>
<td>Tuberculosis</td>
<td>6</td>
<td>2</td>
<td>4</td>
</tr>
<tr>
<td>Other infections</td>
<td>4*</td>
<td>4</td>
<td>0</td>
</tr>
<tr>
<td>Malignant lymphoma</td>
<td>2†</td>
<td>0</td>
<td>2</td>
</tr>
<tr>
<td>Kaposi’s sarcoma</td>
<td>4</td>
<td>3</td>
<td>0</td>
</tr>
<tr>
<td>Total</td>
<td>27</td>
<td>11</td>
<td>14</td>
</tr>
</tbody>
</table>
*Cryptococcus n = 1; ‡Hodgkin’s lymphoma n = 1; ‡Tuberculosis; Histoplasma n = 1; non-Hodgkin’s lymphoma n = 1; Acute abscesses n = 1.

lesions; one aspirate was from an intra-abdominal mass taken under computed tomographic guidance. The site of the lymph node aspirates were: cervical n = 14; axillary n = 4; inguinal n = 7; submandibular n = 1; and intra-abdominal n = 1.

The cytological diagnoses of the 27 aspirated cases were: reactive changes n = 11; tuberculous n = 6; other infections (one cryptococcosis, one histoplasmosis, two acute abscesses) n = 4; malignant lymphomas (one non-Hodgkin’s lymphoma, one Hodgkin’s lymphoma) n = 2; Kaposi’s sarcoma n = 4 (table).

Sixteen corresponding histological lymph node biopsy specimens were available for comparison. The histological diagnosis agreed with the cytological diagnosis in 14 cases and comprised: five reactive changes, four tuberculoses, two malignant lymphoma (one non-Hodgkin’s lymphoma and one Hodgkin’s lymphoma) and three Kaposi’s sarcoma. Disagreement between the cytological and histological diagnoses occurred in two cases, both reported as reactive changes on cytology and as tuberculosis on histology reports.

Eleven FNA cases were subjected to clinical follow up alone. In four of them extranodal histological analyses were available and were identical with the cytodiagnosis (one cryptococcosis, one histoplasmosis, one tuberculoses and one Kaposi’s sarcoma).

Discussion
The cytological diagnosis of reactive changes also included the progressive generalised lymphadenopathy (PGL) seen in AIDS. The diagnostic criteria for tuberculosis were based on the presence of acid-fast bacilli in the aspirate. These cells included a spectrum of small and large lymphocytes some of which were in mitosis. Fragments of reactive follicular centre cells, including tingible body macrophages, syncytia of dendritic reticulum cells, and some plasmacytoid cells were also seen.

PGL is defined as unexpected enlargement of two or more extra-inguinal lymph nodes of at least three months’ duration in individuals at risk for AIDS. The histological appearance varies, depending on the duration of the lesion, with florid reactive hyperplasia in the early stage to advanced lymphocytic depletion and “burnt-out” follicle centres in the late stage. This was reflected in the cytology in some cases, but the changes of PGL could not always be differentiated from reactive lymphomas with different aetiology. The FNA material obtained from lymph nodes with tuberculous lymphadenitis showed caseation necrosis or epithelioid granulomas identical in appearance with those seen in HIV negative patients. The granulomata were composed of syncytial aggregates of oval or fusiform epithelioid histiocytes with “blunt-ended” and sometimes bent or curved nuclei. Langhans’ type giant cells were very occasionally seen. Definitive diagnosis of tuberculosis was made only when acid fast bacilli were demonstrated on Ziehl-Neelsen stain. A Grocott stain was also carried out on every case where material was available to exclude other organisms. In the absence of acid fast bacilli or other organisms, the presence of caseation necrosis or granulomas in lymph node aspirates in HIV positive subjects was reported as consistent with tuberculosis unless proved otherwise.

The specificity of the FNA cytology in our study was 87.5%. This compares favourably with a study by Bottles et al from San Francisco, the only similar study of which we are aware. The authors studied 121 lymph node FNA’s from 113 men with AIDS. The cytological diagnoses included: 60 hyperplasias, 24 non-Hodgkin’s lymphomas, 21 mycobacterial infections, 12 Kaposi’s sarcomas, one Hodgkin’s disease, one giant cell carcinoma, one nasopharyngeal carcinoma, and one squamous cell carcinoma. Histological or microbiological confirmation of the cytological diagnosis was available in 59 specimens. The specificity of the cytology in these cases was 93%. Five cases were reported on FNA as hyperplasia and as malignant on histology (Hodgkin’s disease three cases, non-Hodgkin’s lymphoma one case, and Kaposi’s sarcoma one case). The review of the cytological material showed no evidence of malignancy and therefore these five false negative results were interpreted as sampling error.

In our study two cases were reported on cytology as non-specific reactive changes alone (including PGL). Histological examination showed these to be tuberculosis (false negative rate 12.5%). The aspirate of the first case, which was performed at the early stages of the study, contained insufficient material for carrying out special stains. The second aspirate probably represented a sampling error as no acid fast bacilli were shown on the Ziehl-Neelsen stain. The false negative rate reported by Bottles et al in the San Francisco study was 10%, which is similar to ours.

No false positive results were reported in our study and no malignancy was missed. Our diagnostic difficulties were initially mainly in the area of differentiating tuberculosis from reactive changes which were later remedied by ensuring that sufficient material for routine staining for acid fast bacilli was available in every case. We therefore recommend routine staining for acid fast bacilli on...
every lymph node aspirate from HIV positive patients.

Bottles draws attention to the possibility of sampling errors in the cases of Hodgkin’s disease where FNA may miss the affected part of the lymph node. Classic Reed-Sternberg cells and Reed-Sternberg cell variants are required for the diagnosis of Hodgkin’s disease. In our only case of Hodgkin’s disease occasional classic Reed-Sternberg cells were recognised and these allowed the correct diagnosis to be made. We agree, however, that if there is a discrepancy between the cytopathological diagnosis of non-specific reactive changes and clinically suspicious lesions for malignancy or infection, a repeat aspirate or surgical biopsy should be performed. Close communication between the pathologist and the clinician is essential.

In our series Kaposi’s sarcoma did not present us with major diagnostic difficulties. All four cases of Kaposi’s sarcoma had similar morphology of tightly packed rather monotonous atypical elongated cells with overlapping nuclei, and showing little pleomorphism and mitotic activity.

Of the 11 cases that had only clinical follow up at the time of the lymph node aspiration, four cases had identical extra-nodal pathology. In the remaining seven cases the cytopathological diagnosis was accepted as definitive. The clinical confidence in the cytopathological diagnosis increased with time and this led to a subsequent reduction in the number of biopsies performed. There are limitations to the FNA method, however, such as sampling error, which may result in false negative results. These diagnostic limitations could be overcome in many cases by close communication between cytopathologist and clinician. Repeat aspiration is advised when there is a discrepancy between the cytopathological diagnosis and the clinical impression. FNA does not entirely exclude the need for histological methods and in difficult cases biopsy is still advised.

Although FNA is a relatively simple procedure we emphasise that certain precautions should be observed especially when dealing with HIV positive patients. The aspirator should wear a gown and special gloves to avoid needlestick injuries. Care must be taken in obtaining and transporting the specimens to the laboratory according to standard health regulations for dealing with hazardous material.

In conclusion, we recommend the fine needle aspiration (FNA) of lymph nodes in HIV positive patients as a reliable method for the diagnosis and follow up of cases with lymphadenopathy. The method is accurate, cost-effective and acceptable to patients.