Rapid organism identification from Bactec NR blood culture media in a diagnostic microbiology laboratory

P M Claxton, R G Masterton

Abstract

Aims—To evaluate rapid organism identification on positive blood culture Bactec NR media (phial types 26, 27, 42 and 17), and to assess the usefulness of these procedures in a diagnostic microbiology laboratory.

Methods—Two hundred and sixty, first positive, blood culture bottles from individual patients were tested by rapid identification methods selected on the basis of Gram film organism morphology. Tube coagulase and latex agglutination were applied to presumptive staphylococci; latex agglutination antigen detection methods to suspected pneumococci, *Neisseria* and *Haemophilus* sp; and latex agglutination grouping tests for cultures thought to be non-pneumococcal streptococci.

Results—Media type did not influence test performance ($p > 0.05$ for all comparisons). Misapplication of methods occurred on eight occasions and there were 14 false positive results, nine involving the latex reagents for group C streptococci and pneumococci. The positive predictive values for tube coagulase tests and latex reactions for *H influenzae* type b, and *N meningitidis* groups B and C were 100%. The pneumococcal and staphylococcal latex tests gave positive predictive values of 94.1% and 62.5%, respectively, and the corresponding figure for streptococcal grouping reactions was 75.9%. With the exception of staphylococcal latex testing (80%) all investigation negative predictive values were $> 90\%$.

Conclusions—The performance of the staphylococcal latex agglutination method was unsatisfactory and it is not appropriate for use with the media studied. In view of the cross-reactions observed with the tests used to identify group C streptococci and pneumococci, positive findings must be interpreted with caution. In all other regards the protocol evaluated produced rapid, reliable, clinically useful information and, subject to local experience, is recommended to users of Bactec NR media.

(J Clin Pathol 1994;47:796-798)

detection of bacterial growth. However, the potential clinical benefits of this may be limited in practice as conventionally presumptive organism identification relies on Gram film morphology until culture becomes available, often 18-24 hours later.

Many workers have therefore used a variety of methods¹⁻⁴ to explore rapid organism identification from positive blood culture bottles. These studies have evaluated the performances of specific tests rather than the utility of the approach in general. Only small numbers of blood cultures were included and results have been shown to be influenced by the type of media used.¹

The published work has not included assessment of rapid identification methods in the recently available Bactec NR Plus media. Therefore, this study was designed to evaluate rapid organism identification using the new media, and to assess the usefulness of this approach in a diagnostic microbiology laboratory.

Methods

Adult patient blood culture sets comprised a pair of aerobic and anaerobic Bactec NR Plus media with paediatric samples made up of a Bactec NR Peds Plus bottle and an NR anaerobic bottle (phial types 26, 27, 42 and 17, respectively). The first positive blood culture bottle from any patient was included in the study.

The procedure for carrying out direct tube coagulase tests and staphylococcal and streptococcal latex agglutination reactions involved preparing an organism suspension by double centrifugation.^{1,2} Between 8-10 ml of the positive blood culture was centrifuged at 700 rpm for 10 minutes to remove the red blood cells. The supernatant fluid was recovered and centrifuged at 3000 rpm for a further 10 minutes. The resultant deposit was re-suspended in 0.5 ml sterile saline to produce the in-use suspension.^{1,2}

For latex agglutination antigen detection of *Streptococcus pneumoniae*, *Neisseria meningitidis*, and *Haemophilus influenzae*, 2 ml of the positive blood culture was centrifuged at 3000 rpm for 10 minutes and the supernatant fluid used as the test material.³

The rapid organism identification methods used on each positive blood culture bottle were determined by the organism morphology on the Gram film. Where relevant all methods thought to be appropriate were applied. Cultures thought to be mixed were excluded from the study.

Central
Microbiological
Laboratories, Western
General Hospital,
Crewe Road,
Edinburgh EH2 4XU
P M Claxton
R G Masterton

Correspondence to:
Dr R G Masterton

Accepted for publication
28 February 1994

Automated blood culture systems and improved blood culture media permit early

Table 1 Rapid organism identification by culture medium type

Culture identification	n =	Phial types				No correctly identified by rapid methods
		26	27	42	17	
<i>Haemophilus influenzae</i> type b	19	2	0	17	0	16
<i>Neisseria meningitidis</i> group B	7	0	0	7	0	7
<i>Neisseria meningitidis</i> group C	2	0	0	2	0	2
<i>Streptococcus pneumoniae</i>	66	41	9	8	8	64*
Coagulase negative staphylococci	99					
tube coagulase	96	55	7	29	5	96
latex agglutination	67	45	4	15	3	64
<i>Staphylococcus aureus</i>	31					
tube coagulase	29	24	2	2	1	19
latex agglutination	21	17	1	3	0	5
<i>Enterococcus</i> sp	15	7	1	5	2	15#
Group A streptococcus	5	2	0	2	1	2§
Group B streptococcus	2	0	0	2	0	2
Group C streptococcus	1	0	1	0	0	1†
Group G streptococcus	2	1	1	0	0	2
Non-groupable streptococci	8	2	0	5	1	7‡

n = total number of isolates cultures

*Two not tested with pneumococcal latex reagent but identified as group C streptococci. Additionally, three other pneumococcal latex positive isolates cross-reacted with group C streptococcal latex suspension.

One isolate cross-reacted with pneumococcal latex reagent

§Three not tested directly with group A streptococcal reagent

†This isolate cross-reacted in the pneumococcal latex reagent

‡One isolate was not tested directly with streptococcal grouping suspensions, but gave a false positive pneumococcal test.

Gram positive cocci with staphylococcal morphology were investigated by a tube coagulase test using rabbit plasma.⁵ Staphylococcal latex agglutination (Staphaurex, Murex Ltd) testing was introduced later in the trial as an additional procedure. Gram positive cocci with streptococcal morphology were tested by a latex agglutination grouping method (Streptex, Murex Ltd).

Neisseriae were identified by latex agglutination antigen detection (Wellcogen, Murex Ltd) using coated latex particles reactive against *N meningitidis* groups A,C,Y,W135, and group B/*Escherichia coli* K1. Organisms thought morphologically to be haemophili or pneumococci were investigated with the appropriate latex agglutination antigen detection test (Wellcogen, Murex Ltd) where assessment of the former was restricted to the presence or absence of type b antigen.

Each kit was used according to the manufacturer's recommendations. After culture organisms were identified to species level using standard microbiological techniques. The coagulase reaction of staphylococci was determined by the tube method using rabbit serum⁵ and β haemolytic streptococci were grouped by a latex agglutination test (Streptex, Murex Ltd). The identification of

isolates of *S pneumoniae*, *N meningitidis*, and *H influenzae* were confirmed by reference laboratories.

Statistical analyses were performed using Fisher's exact probability test. The sensitivities, specificities, and predictive values of the different methods were calculated using standard formulae.⁶

Results

A total of 260 positive blood cultures were evaluated. One culture yielded mixed growth. The results of rapid organism identification by culture medium type are shown in table 1. A statistical analysis of the true and false reactions by phial type was performed for staphylococcal coagulase, pneumococcal, and staphylococcal latex tests as there were insufficient numbers of the other organisms for valid comparisons (two tailed Fisher's exact test; $p > 0.05$ for all evaluations).

Table 2 shows the results for the individual investigations. The combined findings of the streptococcal grouping reactions produced a sensitivity of 100%, a specificity of 50%, and positive and negative predictive values of 75.9% and 100%, respectively. There were eight misapplications of testing methods involving three staphylococcal and five streptococcal isolates. Of these, two staphylococci produced false positive group D streptococcal reactions, and two pneumococci were positive with group C streptococcus reagent but were not tested against pneumococcal latex suspension. Three group A streptococci were negative to pneumococcal agglutination but were not investigated with streptococcal grouping agent. The final misapplication involved a coagulase negative *Staphylococcus* which was thought morphologically to be a *Streptococcus* and was investigated as such, being negative to pneumococcal and streptococcal grouping reagents.

There were 14 false positive results. A mixed culture, an *Enterococcus* sp, a group C *Streptococcus*, and a non-groupable *Streptococcus* cross-reacted with pneumococcal latex reagent. Five pneumococci gave false positive group C streptococcal reactions. Three of these strains agglutinated pneumococcal latex suspension, the other was tested only with streptococcal grouping reagents. Two staphylococci gave false positive group D streptococ-

Table 2 Individual test performances

Test	n =	TP	TN	FP	FN	% Sensitivity	% Specificity	PPV	NPV
Tube coagulase	125	19	96	0	10	65.5	100	100	90.6
<i>Staphylococcus</i> latex	88	5	64	3	16	23.8	95.5	62.5	80.0
<i>H influenzae</i> type b latex	19	16	N/A	0	3	84.2	N/A	100	N/A
<i>N meningitidis</i> group B latex	9	7	2	0	0	100	100	100	100
<i>N meningitidis</i> group C latex	9	2	7	0	0	100	100	100	100
<i>S pneumoniae</i> latex	88	64	20	4	0	100	83.3	94.1	100
Group A streptococcus	36	2	34	0	0	100	100	100	100
Group B streptococcus	36	2	34	0	0	100	100	100	100
Group C streptococcus	36	1	30	5	0	100	85.7	16.7	100
Group G streptococcus	36	2	34	0	0	100	100	100	100
<i>Enterococcus</i> sp	36	15	19	2	0	100	90.5	88.2	100

n = number of test performances; TP = number of true positives; TN = number of true negatives; FP = number of false positives; FN = number of false negatives; N/A = not applicable as no true negatives tested; PPV = % positive predictive value; NPV = % negative predictive value.

cal reactions, and there were three false positive staphylococcal latex agglutinations.

Discussion

The purpose of rapid blood culture isolate identification is to produce the earliest possible clinically meaningful information and it is particularly relevant where the data can be used to guide management. To this end most of the published work has investigated staphylococci and streptococci, especially pneumococci, with some attention also to *Neisseria meningitidis* and *Haemophilus influenzae*. With the exception of one small study, Gram negative bacilli in general have not been explored.² This probably reflects less pressing diagnostic needs and the lack of influence such results are likely to have on treatment where antibiotic modification usually awaits sensitivity testing.

The performance of rapid blood culture isolate testing has limitations. The manufacturers of streptococcal latex agglutination products all recommend that to minimise false reactions the method should only be applied to β haemolytic cultures. This is impossible when using the tests for direct investigations. Furthermore, organism identification is normally performed only on pure cultures and where initial investigations have indicated the genus. In rapid blood culture identifications the former cannot be guaranteed and the latter is based on Gram film morphology alone. Additionally, the ability to produce rapid results would be undermined if all possible tests were applied to each isolate, a protocol which in any event would be wasteful of human and consumable resources.

Our findings reinforce previous work on *N meningitidis* and *H influenzae*,^{2,3} showing rapid testing to be highly reliable with all positive results confirmed and only three false negative reactions with strains of *H influenzae* type b. The situation is less satisfactory with *S pneumoniae* where the problem of cross-reaction with group C streptococcal reagent is well recognised. In one study this reached 92% (11/12) of clinical blood cultures tested.⁷ Using our selective protocol where not all pneumococci were examined with streptococcal grouping reagent, at 7.6% (5/66) of confirmed identifications, our cross-reaction rate was much lower but still troublesome. Gram film morphology and blood lysis in bottles provided useful supporting identification information. Conversely, when applied to the

non-pneumococcal streptococci, there was a 18.2% (4/22) false positive reaction rate for pneumococcal antigen detection. Inevitably, given the comparatively low incidence of group C streptococcal bacteraemia, this performance makes the test positive predictive value for that organism very poor. However, in the main the direct application of latex grouping methods proved satisfactory in terms of identifying β haemolytic streptococci and enterococci as well as in excluding non-groupable strains.

Rink² commented on the poor performance of the staphylococcal latex identification method applied directly to blood cultures and Rappaport *et al*¹ reported 77.3% sensitivity and 100% specificity for the same reagent used in this study. Our findings indicate it to be less reliable and we recommend that it should not be used as a rapid method with Bactec NR media. A positive tube coagulase is, however, highly predictive, though negative reactions still leave a 10% chance of the organism being *S aureus*.

Despite the limitations inherent in rapid identification of isolates in positive blood cultures, we have shown that the selection of tests based on Gram film morphology and the application of such methods as primary investigations to be robust tools. Although small organism numbers prevent firm conclusions, no media effect was observed. Rapid clinically useful information is produced cost effectively and, while recognising local experience to be essential, we recommend these procedures to any diagnostic laboratory using Bactec NR media.

We are grateful to Murex Ltd for their donation of the Staphaurex used in the trial.

- 1 Rappaport T, Sawyer KP, Nachamkin I. Evaluation of several commercial biochemical and immunologic methods for rapid identification of Gram-positive cocci directly from blood cultures. *J Clin Microbiol* 1988;26:1335-8.
- 2 Rink MJ. Rapid identification of organisms from blood cultures. *Med Lab Sci* 1990;41:6-9.
- 3 Williams RG, Hart CA. Rapid identification of bacterial antigen in blood cultures and cerebrospinal fluid. *J Clin Pathol* 1988;41:691-3.
- 4 Wellstood S. Evaluation of Phadebact and Streptex kits for rapid grouping of streptococci directly from blood cultures. *J Clin Microbiol* 1982;15:226-30.
- 5 Rossney AS, English LF, Keane CT. Coagulase testing compared with commercial kits for routinely identifying *Staphylococcus aureus*. *J Clin Pathol* 1990;43:246-52.
- 6 Strike PW. Test interpretation. In: *Statistical methods in laboratory medicine*. Oxford: Butterworth-Heinemann Ltd, 1991:336-46.
- 7 Lee P, Wetherall BL. Cross-reaction between *Streptococcus pneumoniae* and Group C Streptococcal latex reagent. *J Clin Microbiol* 1987;25:152-3.