Granulocyte-colony stimulating factor concentrations in a patient with plasma cell dyscrasia and clinical features of chronic neutrophilic leukaemia

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Abstract
In order to study the pathogenesis of plasma cell dyscrasias with associated clinical features of chronic neutrophilic leukaemia, the concentration of granulocyte-colony stimulating factor (G-CSF) was measured in a patient, a 73 year old man, who underwent steroid pulse therapy. High G-CSF concentrations and leucocyte counts prior to treatment declined rapidly on administration of dexamethasone, but rose subsequently. G-CSF was not detected in primary cultures of bone marrow cells, but large amounts of interleukin-6 were found in the culture supernatant. These observations suggest that the neutrophilia observed in the patient represented a reactive response to G-CSF secreted from abnormal plasma cells or stromal cells rather than the existence of a genuine myeloproliferative disorder.

Keywords: chronic neutrophilic leukaemia, myeloma, G-CSF, steroid, regulation.
G-CSF concentrations in a patient with plasma cell dyscrasia

Figure 1  Morphology of bone marrow aspirate at diagnosis. Mature neutrophil expansion and increased numbers of plasma cells were observed (May-Grunwald-Giemsa; original magnification ×200).

Figure 2  (A) Leucocyte counts during the three cycles of steroid pulse therapy. (B) Leucocyte counts and serum G-CSF and IL-6 concentrations during the fourth cycle of steroid pulse therapy. Dex = dexamethasone; m-PSL = methyl-prednisolone; WBC = white blood cells.

Chronic neutrophilic leukaemia (CNL) is a rare myeloproliferative disorder characterised by persistent neutrophilia and hepatosplenomegaly. The important criteria for its diagnosis are the exclusion of underlying diseases capable of provoking a reactive neutrophilia and chronic myelogenous leukaemia by appropriate cytogenetic and molecular studies. Interestingly, despite the rarity of this condition, more than 30 cases of CNL have been reported in association with plasma cell dyscrasias. The high frequency of this unusual association involving both myeloid and lymphoid lineages raises the question whether the neutrophilia represents the existence of a genuine myeloproliferative disorder or is a reactive response. Recently, dysregulation of cytokines has been implicated in the pathogenesis of various haematological disorders. To study a possible role of abnormal cytokine production in the development of this unusual association, we examined serum granulocyte-colony stimulating factor (G-CSF) concentrations in a patient presenting with clinical features consistent with CNL and a plasma cell dyscrasia.

Case report
The patient, a 73 year old man, presented in April 1992 for further evaluation of a leucytosis. A complete blood count revealed a haemoglobin of 11.4 g/dl, a leucocyte count of 28 080/μl, and a platelet count of 137 000/μl (leucocyte differential: neutrophils, 85%; lymphocytes, 9%; monocytes, 3%; eosinophils, 3%). The bone marrow was grossly hypercellular with a predominance of mature neutrophils. An increased number of plasma cells (7.6% of the total nucleated cells) also was observed (fig 1). An IgG, K paraprotein was detected in the serum at a concentration of 32 g/l with depressed levels of IgA (0.8 g/l) and IgM (0.9 g/l). Bence-Jones protein was detected in the urine. The NAP (neutrophil alkaline phosphatase) score was 383 (normal 120). A microbiological screen was negative. Chromosome studies of marrow cells revealed a normal male karyotype, and molecular genetic analysis showed no evidence of bcr gene rearrangement. The serum G-CSF concentration was 475 pg/ml and interleukin-6 (IL-6) was not detected. The patient was diagnosed as having plasma cell dyscrasia associated with CNL and monthly cycles of melphalan therapy (8 mg/day/4 days) were initiated, with no obvious clinical improvement. Treatment with busulfan (2–4 mg/day) was instituted, which controlled the leucocyte count and paraproteinemia until April 1994, when thrombocytopenia (32 000/μl) developed. The patient was then admitted to hospital for steroid pulse therapy.

As shown in fig 2A, treatment with dexamethasone (40 mg/day for four days) rapidly reduced the leucocyte count from 13 150/μl (neutrophils, 86%) to 1650/μl (neutrophils, 12%) by the fifth day of treatment. However, the leucocyte count transiently increased to 39 590/μl on the eighth day before returning to the pretreatment level by the 14th day. Similar changes in leucocyte counts were observed during second and third clinical courses of steroid pulse therapy.

To elucidate the relation between leucocyte counts and serum G-CSF concentration, we monitored both during a fourth course of steroid therapy and simultaneously performed in vitro analysis using primary cultures of bone marrow cells aspirated prior to treatment.
Cytokine concentrations in the serum and the supernatant of the cultures were assessed by enzyme immunoassay (Amersham Life Science, Tokyo, Japan). On administration of dexamethasone the serum G-CSF concentration rapidly declined from 137.0 pg/ml to 15.7 pg/ml on the fifth day in parallel with the leucocyte count. However, the serum G-CSF concentration rose to 525.4 pg/ml on the ninth day, which was followed by a fall to 275 pg/ml on the 15th day. IL-6 was not detected (<10 pg/ml) in serum prior to the treatment, but a serum IL-6 concentration of 107 pg/ml was measured on the 15th day (fig 2B). Meanwhile, in vitro studies revealed that large amounts of IL-6 (26479 ± 329 pg/ml/2 × 10⁶ cells) were found in the supernatant of cultures on day 6. Unexpectedly, G-CSF was not detected (<7 pg/ml), although small amounts of granulocyte-macrophage colony stimulating factor (2 pg/ml) were found in the supernatant of these cultures.

**Discussion**

Two interesting observations were made in this study. First, high concentrations of G-CSF were found in the serum of a patient with clinical features consistent with both CNL and a plasma cell dyscrasia. Steroid pulse therapy rapidly reduced serum G-CSF concentrations, which coincided with a decline in the patient’s leucocyte counts. Interestingly, this downregulation was counteracted by upregulated IL-6 secretion from the patient’s leucocytes. These observations suggest that the neutrophilia observed in this patient was dependent on the serum G-CSF concentration. Furthermore, the rapid and transient alterations in G-CSF concentrations suggest that steroids inhibit G-CSF synthesis or secretion from producer cells rather than killing them. Although the mechanism of genetic regulation of G-CSF synthesis is not understood fully, a recent study has identified a binding site for NF-IL-6, which is a nuclear transcriptorial factor for IL-6 synthesis, in the promoter region of the G-CSF gene. There may be a similar inhibitory mechanism involving IL-6 synthesis, where steroids antagonise the NF-IL-6 binding region and inhibit G-CSF synthesis at the level of transcription.

The second interesting observation was that high concentrations of IL-6 were present in the supernatant of primary cultures of bone marrow cells, although we could find no detectable concentrations of IL-6 in the serum prior to treatment. Previous studies have suggested that serum IL-6 concentrations may not reflect those in the bone marrow. In our patient, it is likely that abnormal plasma cells or activated stromal cells secreted IL-6 and supported the clonal expansion of plasma cells via an autocrine or paracrine mechanism.

 Unexpectedly, however, we failed to detect G-CSF in the culture supernatant, despite the high serum concentrations of G-CSF. One possible explanation is that the secreted G-CSF bound rapidly to its receptors expressed on the numerous immature myeloid cells present in the culture and disappeared from the supernatant. This raises the question of the cell of origin of this G-CSF overproduction. Recent studies have demonstrated that not only IL-6, but G-CSF mRNA is expressed in most bone marrow cells of patients with myeloma. It is possible that abnormal plasma cells in the patient secreted G-CSF by themselves. Alternatively, IL-1 and tumour necrosis factor, which is produced by myeloma cells, may in turn activate the stromal cells to produce G-CSF. To resolve this question, in situ hybridisation using a specific G-CSF probe might be helpful.

In conclusion, the persistent neutrophilia in this patient seems to represent a proliferative reaction to the overproduction of cytokines rather than a distinct myeloproliferative disorder. In support of this hypothesis, Standen et al. have reported the case of a patient with CNL and myeloma that clearly demonstrated the polyclonal nature of myeloid cells on analysis of restriction fragment length polymorphisms using an X-linked probe. However, additional studies are required to delineate the nature of the myelopoietic stimulus in this unusual subgroup of patients with CNL.