Quantitation in inflammatory pleural disease to distinguish tuberculous and paramalignant from chronic non-specific pleuritis

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Abstract

Aim—To determine by morphometry if pleural biopsies with the histopathological diagnosis of “non-specific pleuritis”, malignant, and tuberculous disease could be distinguished morphologically from those with truly non-specific disease.

Methods—Each pleural biopsy was reviewed taking into account three compartments of reference: the visceral/parietal mesothelial compartment, the submesothelial screen compartment, and the submesothelial adipose tissue compartment. Normal connective tissue, granulation tissue, fibrocellular proliferation, fibrin, polymorphonuclear cells, mononuclear cells, and mesothelial cells were measured using conventional point counting procedures in terms of the fractional area occupied by each parameter within each compartment of reference. Ranking was carried out on 164 patients, based on their diagnosis: chronic non-specific disease (n = 57), tuberculosis (n = 27), malignant disease (n = 58), and conditions associated with transudative effusions (n = 22).

Results—Stepwise discriminant analysis of the resulting data showed that biopsies from patients with tuberculosis, malignant disease, and chronic non-specific disease could be distinguished between themselves and normal cases. Statistical differences among the four groups were observed for eight morphometric parameters related to components of inflammation and extension throughout the three pleural anatomical compartments. A robust discriminant function permitted an adequate classification of the three groups of disease in 88.41% of the cases. Pleural biopsies with fibrin incorporated within granulation tissue on the submesothelial screen compartment showed 100% specificity for patients with tuberculosis, while mononuclear cells in a band-like infiltrate on the submesothelial adipose tissue compartment showed 93.1% specificity for patients with malignant disease. The truly non-specific pleuritis was characterised by deposits of fibrin in the subpleural compartment and discrete signs of chronic inflammation and reparatory fibrosis on the submesothelial screen.

Conclusions—Morphometric analysis of pleural biopsies may be a useful supplementary histological procedure to support the diagnosis of pleural tuberculosis and malignant disease.

Keywords: morphometry; pleural biopsy; chronic non-specific pleuritis

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pleural fragment yielded mycobacteria. In another nine patients a second biopsy disclosed granulomata and pointed towards the diagnosis. In six cases, both the second biopsy and culture of the first biopsy fragment were diagnostic. Three patients were diagnosed based upon sputum isolation of mycobacteria.

Patients were considered to be bearers of malignant disease (n = 58) when neoplasms was found with no other identifiable cause for an exudative effusion. Diagnosis was reached by a second pleural biopsy in nine cases, by cytological analysis of the pleural fluid in 11 cases, and by a second biopsy and cytological examination in eight cases. In the remaining 30 cases, thoracic malignant disease was found by means of other diagnostic procedures such as bronchoscopy, mediastinal or transthoracic biopsy, or the discovery of carcinoma at another site such as the breast. In these cases, malignant cells were not found in pleural tissue or fluid, rendering the effusions as paramalignant (associated with malignancy but with no pleural involvement).

After at least three months of follow up subsequent to the second biopsy, 57 patients with exudative pleural effusion showed no evidence of malignancy or tuberculous disease and had a clinical diagnosis (atypical pneumonia, parapneumonic effusions, hepatitis, and splenic infarct) compatible with chronic non-specific disease. This group was classified as chronic pleuritis. Although a follow up of three months is somewhat short, it is of significance when malignancy and/or tuberculosis are considered.

Patients with clinical conditions associated with transudative effusions (congestive heart failure, cirrhosis of the liver, and nephrotic syndrome) and whose follow up of at least three months showed no evidence of malignancy or tuberculous disease composed the fourth group (n = 22).

**MORPHOLOGICAL STUDY**

The same slides containing biopsy specimens in cubes or strips 2–3 mm long, analysed for the initial pathological report (paraffin embedded, haematoxylin and eosin stained) were reviewed blind until the final diagnosis.

Morphometry was performed on the different layers of the pleura. The anatomical layers of the pleura are shown diagrammatically in Fig 1. Histologically, the parietal pleura is composed of two different layers: a complete layer of mesothelial cells and a thin screen of submesothelial connective tissue, with a well developed network of collagenic and elastic fibres, which contains lymphatic and blood vessels. This layer is in continuity with one composed of adipose and skeletal muscle tissues. The tissues in the pleural biopsies were divided into three compartments of reference: (1) the space between the parietal and visceral pleura was designated as the visceral/parietal mesothelial compartment; (2) the mesothelium and the thin submesothelial connective tissue was designated as the submesothelial screen compartment; and (3) the continuous layer of adipose tissue underlying the submesothelial

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**Figure 1** The anatomical layers of the pleura.

The biopsies were performed by a resident or fellow under staff supervision. Three or four specimens of tissue, obtained with a Cope needle, were used for histological study, mycobacterial culture, and fungal culture. Pleural fluid obtained at the time of the biopsy was submitted routinely for a cell count with white cell differential, protein, glucose, and lactate dehydrogenase measurements, bacterial culture, and cytological examination. A second biopsy was performed whenever the pleural fluid exhibited exudative characteristics and the histological findings were non-diagnostic.

In 147 patients, biopsy led to a specific diagnosis. In the remaining 164 patients, chronic non-specific pleuritis was diagnosed after the first closed pleural biopsy and these patients were classified into four groups according to their follow up diagnosis.

Tuberculous disease was the final diagnosis in 27 patients. In nine cases, culture of the...
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screen was designated as the submesothelial adipose tissue compartment.

Subsequently, specimens were evaluated for:
(1) tissue type (normal connective tissue, granulation tissue, and fibrocellular proliferation); (2) components of inflammation (fibrin, polymorphonuclear cells, and mononuclear cells); (3) mesothelial cells; and (4) extension and distribution of the pathological process throughout the three anatomical compartments.

Morphometric studies were performed by a conventional point counting procedure using a miculated eyepiece (100 points and 50 lines). Counting was performed using a cascade progressive sampling approach. In each case, at a magnification of ×100, five non-coincident microscopic fields were studied to quantify the relative fraction of the three pleural compartments by counting a total of 500 points that covered an area of 1 mm² per biopsy. The error of this procedure was estimated according to Gundersen and was smaller than 5%. Next, the numbers of points overlying fibrin, granulation tissue, and fibrocellular proliferation were counted in five fields, at a magnification of ×400, covering an area of 62 500 μm² in each pleural compartment, actually representing the fractional area occupied by each parameter within each compartment of reference. Finally, five fields, at a magnification of ×1000 (10 000 μm²) were counted in each compartment to evaluate the fractional area occupied by polymorphonuclear cells (neutrophils + eosinophils), mononuclear cells (macrophages + lymphocytes + plasma cells), and mesothelial cells. The relative fraction of area was obtained by dividing the values reached by the corresponding parameter under study. Interobserver and intraobserver variability was 10% and 8%, respectively.

![Figure 3](http://jcp.bmj.com/) Fractional area (%) of tissue types expressed as (A) normal connective tissue, (B) granulation tissue, and (C) fibrocellular proliferation in the four groups.

![Figure 4](http://jcp.bmj.com/) Fractional area (%) of components of inflammation and mesothelial reactivity expressed as (A) polymorphonuclear cells, (B) mononuclear cells (lymphocytes + plasma cells), (C) mesothelial cells, and (D) fibrin in the four groups.

**STATISTICAL ANALYSIS**

Discriminant analysis was used to obtain a statistical classification of the four groups. This method finds the linear (additive) combination of variables that gives the clearest separation from individuals into different groups. This procedure includes identification of the variables that contribute significantly to discrimination. The criterion for inclusion of a variable was an F value of 3.0 or more, roughly corresponding to p = 0.04. Further independent combinations of the same variables were calculated. Plotting of the values from the first two linear discriminant functions combined for each individual, showed almost the separation achieved between the groups. A stepwise procedure was used to select the variables relevant to distinguish the groups. Because the discriminant power would be optimistic when assessed on the same data used to derive the functions, a jack knife (one out) procedure was included in the results. This procedure withdraws one patient (patient 1, for instance) from the analysis, then the model is re-estimated excluding that patient. Afterwards, the
excluded patient is classified according to the new model, and his actual classification compared to the predicted one. Next, patient 1 is again included in the analysis and patient 2 is withdrawn, according to the same procedure, until calculations are completed for all patients included in this study. All statistical procedures were performed using the SPPS (v 6.0) statistical package and the level of significance was 0.5%.

Results

Individual Morphometric Measurements

Figures 2-4 show the morphometric data from the 164 serial biopsies. The submesothelial screen compartment (fig 2B), fibrin (fig 4D), granulation tissue (fig 3B), and polymorphonuclear cells (fig 4A) were higher in tuberculosis, whereas the submesothelial adipose tissue compartment (fig 2C) and mononuclear cells (fig 4B) were found more frequently in the group with neoplasia. Fibrocellular proliferation (fig 3C) and normal connective tissue (fig 3A) characterised chronic pleuritis and normal pleura, respectively.

Statistical Analysis

Using discriminant analysis, different combinations of the morphometric data selected eight variables capable of distinguishing the groups. These are shown below, with a p value relating to removal from the model: (1) submesothelial adipose tissue compartment, p < 0.001; (2) submesothelial screen compartment, p < 0.001; (3) visceral/parietal mesothelial compartment, p < 0.001; (4) fibrin, p < 0.001; (5) granulation tissue, p < 0.001; (6) fibrocellular proliferation, p < 0.001; (7) polymorphonuclear cells, p < 0.001; and (8) mononuclear cells, p < 0.001. These relevant variables were used to construct the model presented in fig 5, where a plot of the values of the first two linear discriminant functions for each individual shows almost the separation into the groups. Discriminant analysis allowed us to define four distinct patterns of pleural involvement.

Tuberculous pleuritis—the fraction of the area occupied by the submesothelial screen compartment was significantly higher than in the other conditions (fig 2B) and the morphometric quantification of polymorphonuclear cells (fig 4B), fibrin (fig 4A), and vascularised connective tissue (fig 3B) was statistically higher in tuberculosis than in the other groups. Fibrous inflammation was the typical morphological reaction and this exudate was incorporated into the submesothelial screen, resulting in vascularised connective tissue (fig 6). Twenty-seven tuberculous patients (100%) were correctly classified as having tuberculosis pleuritis (table 1).

Paramalignant pleuritis—the fraction of the area occupied by the submesothelial adipose tissue compartment (fig 2C), mononuclear (fig 4C), and mesothelial cells (fig 4D) was statistically higher in malignant or paramalignant pleuritis than in tuberculosis or chronic pleuritis. Here, the submesothelial screen was normal or showed non-specific findings, such as clusters of lymphocytes around the vessels. The submesothelial adipose tissue was infiltrated characteristically by lymphocytes and plasma cells in a band-like pattern (fig 7). Eosinophils were scattered throughout the mononuclear cells. Mesothelial hyperplasia

Figure 5  Graphical representation of the results of stepwise discriminant analysis.

Figure 6. Tuberculous pleuritis: a fibrinous inflammation (F) is the typical morphological reaction seen in the submesothelial screen compartment (SMSC). It is characterised by exudation of plasma proteins and precipitation of masses of fibrin, forming a tangled eosinophilic meshwork to which neutrophils and macrophages are attached. In the bottom of this fibrinous deposit the exudate is organised into vascularised connective tissue (V) (haematoxylin and eosin; original magnification ×63).
Table 1 Percentages of biopsies that were correctly classified by the model (incorporating jack-knife)

<table>
<thead>
<tr>
<th>Actual groups</th>
<th>Number of cases</th>
<th>Chronic pleuritis</th>
<th>Tuberculosis</th>
<th>Neoplasia</th>
<th>Normal</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chronic pleuritis</td>
<td>57</td>
<td>48 (84.2%)</td>
<td>0 (0%)</td>
<td>0 (0%)</td>
<td>9 (15.8%)</td>
</tr>
<tr>
<td>Tuberculosis</td>
<td>27</td>
<td>0 (0%)</td>
<td>27 (100%)</td>
<td>0 (0%)</td>
<td>0 (0%)</td>
</tr>
<tr>
<td>Neoplasia</td>
<td>58</td>
<td>1 (1.7%)</td>
<td>54 (93.1%)</td>
<td>3 (5.2%)</td>
<td></td>
</tr>
<tr>
<td>Normal</td>
<td>22</td>
<td>3 (13.6%)</td>
<td>0 (0%)</td>
<td>3 (13.6%)</td>
<td>16 (72.7%)</td>
</tr>
<tr>
<td>Total</td>
<td>164</td>
<td>60 (36.5%)</td>
<td>80 (48.8%)</td>
<td>12 (7.3%)</td>
<td>12 (7.3%)</td>
</tr>
</tbody>
</table>

The analysis of the results allowed us to identify four distinct patterns of histological pleural involvement.

Tuberculous pleuritis was characterised by a fibrinous inflammation of the submesothelial screen compartment in an absence of granulomata. Our results show how fibrin becomes incorporated into the submesothelial screen, an event that may precede the development of the granulomatous reaction. Pathologists experienced in reviewing pleural biopsies may believe that these results are not realistic. Indeed, fibrin is associated with most effusions (at least in biopsies), but Paterson’s morphometric model for the development of tuberculous pleuritis, which correlates well with what is seen in clinical practice, supports our findings. Both described how the pleura was replaced extensively by a layer of granulation tissue in which tuberculous granulomas were dispersed. These findings could explain those specimens where the needle reached the tissue between the granulomas and only granulation tissue was found. Another important point is that even in the absence of an intact mesothelial lining, pathologists can distinguish between fibrin in granulation tissue in the submesothelial screen compartment (specific for tuberculosis) and fibrin in the visceral/parietal mesothelial compartment (specific for truly non-specific pleuritis). In truly non-specific pleuritis the fibrin lies in the visceral/parietal compartment as a detached pseudomembrane, which has not been incorporated into the submesothelial screen compartment. This compartment shows very discrete signs of chronic inflammation and reparative fibrocellular proliferation instead of fibrinous inflammation.

Paramalignant pleuritis, characterised by a band-like inflammatory reaction of the submesothelial adipose tissue compartment, was the typical morphological reaction seen in pleural biopsies of patients with malignant conditions, but without malignant cells in the pleural tissue at the time of biopsy. Paramalignant pleuritis may be caused by different malignancy-associated conditions that do not result from direct pleural involvement. The inflammatory changes in the submesothelial adipose tissue compartment might be an immunological reaction of the pleura to tumour produced antibodies. A similar situation occurs in the post-cardiac injury syndrome, in which an autoimmune phenomenon produces antibodies, causing an exuberant pleuroperticarditis rich in mononuclear cells. Exuberant mononuclear infiltrates are a well documented immunological phenomenon characterising peripheral reactions to tumours, such as gastric cancer. Also, this band-like cellular infiltrate on the submesothelial adipose tissue might be the result of lymphatic obstruction by tumours. A similar band-like infiltration by mononucleated cells in the submesothelial adipose tissue compartment (associated with non-tuberculous pleuritis) was found by Nagata et al. Despite the diversity of events that occur in tumour pleural relationships, the present study indicated that a band-like inflammatory reaction is a common feature of cases with paraneoplastic pleuritis.

Discussion
In this study, we have shown that by means of a simple and quick method, with no additional costs to the patient, it is possible to characterise non-specific pleuritis further and to suggest a specific diagnosis with reasonable accuracy.

Figure 7 Paraneoplastic pleuritis: characteristically, the submesothelial adipose tissue compartment (SMATC) is infiltrated by lymphocytes, plasma cells, and eosinophils in a band-like pattern (I). Mesothelial hyperplasia is seen re-covering the adipose tissue. Note that the submesothelial screen compartment (SMSC) is normal or shows small clusters of lymphocytes around the vessels (haematoxylin and eosin; original magnification ×63).
involved. Therefore, this lymphoproliferative response of the pleura to a distant tumour may point to the need for a thoracoscopy or other diagnostic procedure.

Pathologists would argue that chronic inflammation and its sequelae could interfere with the recognition of the border between the submesothelial screen compartment and the submesothelial adipose tissue compartment. In this study, we felt that chronic inflammation and its sequelae in the submesothelial adipose tissue compartment were characterised by a mononuclear infiltrate around the vessels and replacement of the screen by a thin layer of collagenous tissue (fibrocellular proliferation), which extended from the submesothelial connective tissue deep into the external elastic lamina (fig 1). The submesothelial screen compartment could be stripped easily from the submesothelial adipose tissue compartment, presumably because the anatomical compartments of the pleura were not destroyed by the original inflammatory process and because the internal elastic was not affected. The density of the mononuclear infiltrate in a band-like pattern on the submesothelial adipose tissue compartment allows the recognition of the border between the submesothelial screen compartment and the submesothelial adipose tissue compartment.

In the third pattern of pleural involvement, apart from improving diagnostic characterization of chronic non-specific pleuritis or benign non-tuberculous pleuritis, the use of quantitative information led to the identification of patients who should continue to be under diagnostic investigation.

A normal pattern of pleural involvement was found in the fourth group of patients with clinical conditions associated with transudative effusions. However, 5.2% of patients with malignancies fell into this group. Therefore, the finding of a normal pleural pattern and a transudate does not exclude the diagnosis of malignancy.

We concluded that morphometric analysis of pleural biopsies could be useful as a supplementary histological procedure in the diagnosis of pleural tuberculosis and malignant disease. Because of its simplicity, efficiency, and low cost the use of morphometric tools in the routine diagnosis of pleural biopsies should be encouraged in the management of patients with pleural effusions.

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